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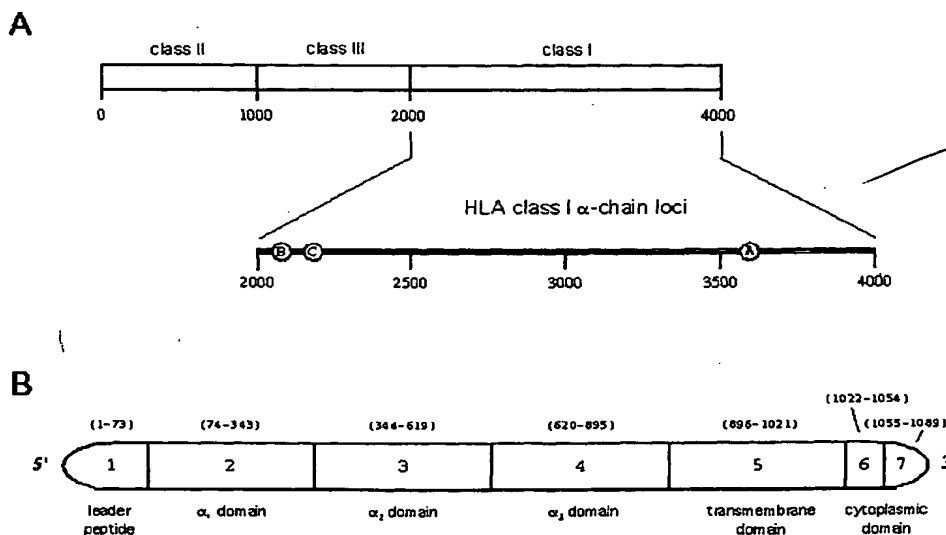
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(54) Title: METHOD AND APPARATUS FOR THE PRODUCTION OF ANTIGENS AND USES THEREOF



(57) Abstract: The field of the invention relates in general to at least one method and apparatus for the production of soluble MHC antigens and more particularly, but not by way of limitation, to at least one method and apparatus for the production of soluble Class I and II HLA molecules. The field of the invention also includes such produced soluble Class I and II HLA molecules and their use. According to the methodology of the present invention, the soluble Class I and II HLA molecules can be produced from either gDNA or cDNA starting material.

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METHOD AND APPARATUS FOR THE PRODUCTION OF ANTIGENS AND USES THEREOF

CROSS REFERENCE TO RELATED APPLICATIONS

This application claims priority under 35 U.S.C. § 119(e) of provisional U.S. Serial No. 60/256,410, filed December 18, 2000, entitled "HLA PRODUCTION FROM GENOMIC DNA," and provisional U.S. Serial No. 60/256,409, filed December 18, 2000, entitled "HLA PROTEIN PRODUCTION FROM CDNA," the contents of which are hereby expressly incorporated in their entirety by this reference.

This application is also a continuation-in-part of U.S. Serial No. 09/465,321, filed December 17, 1999, entitled "METHOD AND APPARATUS FOR THE PRODUCTION OF SOLUBLE MHC ANTIGENS," the contents of which are hereby expressly incorporated in their entirety by this reference.

In addition, this application is a continuation-in-part of U.S. Serial No. 09/974,366, filed October 10, 2001, entitled "COMPARATIVE LIGAND MAPPING FROM MHC POSITIVE CELLS," the contents of which are hereby expressly incorporated in their entirety by this reference.

STATEMENT REGARDING FEDERALLY FUNDED RESEARCH

At least a portion of the invention was developed under funding from the National Institute of Health ("NIH") under contract Nos. No1-A1-45243 and No1-A1-95360. As such, the U.S. Government may own certain rights in and to this

BACKGROUND OF THE INVENTION

1. Field of the Invention

The field of the invention relates in general to at least one method and apparatus for the production of soluble MHC antigens and more particularly, but not by way of limitation, to at least one method and apparatus for the production of soluble Class I and II HLA molecules (i.e. sHLA). The field of the invention also includes such produced soluble Class I and II HLA molecules and their uses. According to the methodology of the present invention, the soluble Class I and II HLA molecules can be produced from either gDNA or cDNA starting material. One such exemplary, but non-limiting, use is the formation of a tetrameric sHLA (or other multimeric complex) complex which may be used to test immunogenicity of peptide ligands of interest - i.e. will a peptide ligand of interest provoke a CTL response and/or preferentially bind a CTL.

2. Brief Description of the Background Art

Class I HLA molecules are polymorphic human glycoproteins that endogenously bind and then extracellularly present peptide ligands to CD8⁺ T lymphocytes. Polymorphisms within the class I peptide binding groove are positioned to moderate ligand binding and presentation to such immune system cells. To date, the small quantities of natural ligands available to those of skill in the art has limited the understanding of precisely how polymorphism alters

peptide binding and in turn, vaccine development and, more basically, if a peptide ligand of interest will provoke a CTL mediated immune response. In order to address the impact of polymorphism upon antigen presentation, the inventors developed the novel approach disclosed herein – i.e. that ligand presentation overlaps exist across the polymorphisms and that these overlaps distinguish divergent class I peptide binding grooves. Utilizing this novel approach and coupling it with a unique hollow-fiber cell culture scheme and utilizing a mass spectrometric ligand mapping approach, large quantities of peptides eluted from soluble class I and class II molecules can be obtained for detailed analyses, vaccine development, and functional testing.

Initially, peptide ligands were extracted from five different HLA-B15 allotypes and subsequently examined. Mapping and characterizing the ligands obtained from these allotypes demonstrated that they: (i) vary in length from 7 to 12 residues; (ii) are more conserved at their C termini than at their N-proximal residues; and (iii) are presented as overlaps contingent on C-terminal preferences. These results provide insight into class I and class II ligand loading not available via other methods, demonstrating that an elemental role is played by a peptide ligand's C terminus during endogenous binding and provides the starting material for a multimeric complex to be used to test the functionality of a peptide ligand of interest. The data obtained, and disclosed herein, validates, and illustrates the unique methods disclosed herein for the

production of sHLA from either gDNA or DNA starting material and the uses to which this sHLA material may be put.

Class I and class II MHC molecules, designated HLA class I and class II, respectively, in humans, bind and display peptide antigens upon the cell surface. The peptides they present are derived from either normal endogenous proteins ("self") or foreign proteins ("nonself"). Nonself proteins include items such as the products of malignant transformation or intracellular pathogens. In this manner, class I and class II molecules convey information regarding the internal fitness of a cell to CD8⁺ CTLs which are activated upon interaction with "nonself" peptides. Such activation may lead the CD8⁺ CTLs to kill and/or suppress a cell which is malignant or contains intracellular pathogens.

Examination of HLA by serologic and molecular methods by those of ordinary skill in the art continues to demonstrate that the class I HLA-A, B, and C molecules are encoded by the most polymorphic genes in mammals. Translating class I polymorphism into the tertiary structure of the class I molecule indicates that residues positioned to affect class I peptide presentation to T lymphocytes are most frequently affected by the mutagenic events which diversify class I loci. Throughout the world, HLA class I molecules exhibit a high degree of polymorphism that is generated by systematic recombinatorial events and collectively allows for the presentation of a vast array of different peptides. Depending upon allelic composition, two individuals' molecules may not

necessarily bind the same peptides with equal affinity or even at all.

While the general structure and function of MHC class I molecules has been reasonably well studied and established, their polymorphic nature and how they specifically influence the capacity of class I in peptide binding and presentation remains an issue of persistent inquiry by those of ordinary skill in the art. The nature of precise overlaps in peptide binding specificity to HLA class I is particularly ill-defined at the current time due to the complexity of peptides bound. For example, this and other issues must be clarified in order to effectively pursue vaccines capable of eliciting protective CTL responses across an extensive population range. Unraveling the functional significance of class I polymorphism is an important issue that requires an understanding of how the mutagenic events diversifying the class I binding groove differentially moderate the presentation of peptide ligands.

The heavy chains of class I molecules are encoded within the MHC and, upon assembling into heterodimers with the light chain, β_2m , are responsible for selectively gathering endogenously processed peptides. Once peptides are collected, mature class I molecules transport the bound peptides to the cell surface where receptors on CD8⁺ T lymphocytes engage the class I molecules to inspect the ligands. CTLs may then be triggered by class I molecules bearing virus or tumor-derived peptides.

With respect to the background art, hereafter numerous references are

disclosed which detail one or more aspects of the background art as it relates to the novel methods and uses of the present invention. As such, each reference listed should be understood as being wholly incorporated by reference herein in their entirety as though the reference were fully transcribed herein. In this manner, one of skill in the art given the present specification would be fully informed and could truly appreciate the novel and unprecedented nature of the invention(s) disclosed and taught herein. The full and complete citations for each reference are appended hereto after the detailed description and before the claims.

The class I molecules expressed upon the nucleated cells of all vertebrate systems studied to date (for example, Ennis et al. 1988; Winkler et al. 1989; Kaufman et al. 1990; Lawlor et al. 1990; Trowsdale 1995; Prilliman et al. 1996; Antao et al. 1999) are heterodimers composed of a glycosylated 45 kDa heavy chain (α -chain) and a 12 kDa light chain (β_2m). In humans, heavy chains are encoded at 3 loci (B, C, and A) within the MHC on the short arm of chromosome 6 (FIG. 1A). FIG. 1B illustrates each α -chain comprised of α_1 , α_2 , and α_3 domains, as well as a transmembrane domain, which tethers the molecule to the cell surface and a short C-terminal cytoplasmic domain (Björkman and class I location and heavy chain coding region).

X-ray crystallography (Björkman et al. 1987a; Madden et al. 1991; Saper et al. 1991; Madden et al. 1992; Collins et al. 1995; Reid et al. 1996; Smith et

al. 1996a; Smith et al. 1996b; Glithero et al. 1999) has illustrated details of the structural relationship of the extracellular α -chain domains and β_2m (FIG. 2). The membrane-proximal domains, α_3 and β_2m , associate in an immunoglobulin fold structure. The membrane-distal α_1 and α_2 domains together create a closed basket-like structure that sits atop the α_3 and β_2m structure (FIG. 2A). It consists of two α -helical "walls" with a "floor" created by eight anti-parallel β -sheets (Björkman et al. 1987a; Madden 1995; and FIG. 2, B and C). In the earliest studies, detection of electron density situated in the α_1/α_2 groove helped to clarify the experimentally-suspected occupancy of peptide fragments 8-10 residues long and thus the function of class I molecules in presenting such peptides upon the cell surface (Björkman et al. 1987b). Initial crystallographic studies designated six subsites, A-F (FIG. 3), or "specificity determining pockets," that constitute the peptide binding groove (Garrett et al. 1989; Saper et al. 1991; Fremont et al. 1992; Matsumura et al. 1992; Elliott et al. 1993; Young et al. 1995). In addition to crystal structure analyses, thermodynamic stability studies indicate that networks of hydrogen bonds to structural residues lining the A- and F-pockets, which lie at opposite ends of the groove, serve to fasten a peptide by its N and C termini, respectively (Bouvier and Wiley 1994); these two pockets are thus implicated in the fixed orientation of a peptide within the binding groove.

Class I molecules primarily associate with peptide fragments, thus

forming α -chain/ β_2 m/peptide trimolecular complexes, via an endogenous processing pathway during their assembly (Germain 1994; Heemels and Ploegh 1995; Lehner and Cresswell 1996; York and Rock 1996; Pamer and Cresswell 1998); in fact, the very binding of peptides is essential for the stabilization and expression of these molecules (Ljunggren et al. 1990; Townsend et al. 1990; Elliott 1991). The class I α -chain and β_2 m are cotranslationally translocated into the ER lumen (Townsend et al. 1990; Germain and Margulies 1993; Neefjes et al. 1993), where the α -chain remains anchored to the ER membrane and is stabilized prior to association with β_2 m and/or peptide through interactions with various chaperone proteins, including BiP, ER-60, calnexin, calreticulin, and tapasin (Nöbner and Parham 1995; Sadasivan et al. 1996; Suh et al. 1996; Spee and Neefjes 1997; Harris et al. 1998; Lindquist et al. 1998). Although several alternative proteolytic processing and transport pathways, certainly exist (for example, Hsu et al. 1991; Henderson et al. 1992; Kozlowski et al. 1992; Roelse et al. 1994; Snyder et al. 1994; Ferris et al. 1996; Craiu et al. 1997; Luckey et al. 1998; Mosse et al. 1998; Wang et al. 1998; Young et al. 1998), it is believed that the majority of peptides the nascent class I molecules interact with are delivered to the ER in a distinct series of events.

Proteins in the cytoplasmic compartment are first enzymatically degraded into peptides of relatively uniform length by an ATP-dependent proteasome

complex (Coux et al. 1996). Some proteasome components include the IFN- γ inducible subunits LMP2 and LMP7; these are themselves encoded within the MHC (Gaczynska et al. 1994). The typically nonameric fragments produced are then actively conveyed across the ER membrane via a dimer of TAP1/TAP2, an MHC-encoded ATP-binding cassette transporter (Monaco et al. 1990; Parham 1990; Grandea III et al. 1995). Once inside the ER, a processed peptide can be captured within the α_1/α_2 groove of a class I molecule and a stable trimer is formed (Germain and Margulies 1993; Smith et al. 1995). This trimeric α -chain/ β_2m /peptide complex is then transported through the Golgi complex and ultimately to the extracellular surface. The processes of class I assembly and transport are summarized in FIG. 4.

Following cell surface localization, mature complexes of class I bearing peptide antigens become available for interaction with receptors on monocytes, B and T lymphocytes, and NK cells (Townsend and Bodmer 1989; Yokoyama 1993; Lanier and Phillips 1996; Borges et al. 1997; Cosman et al. 1997). Their primary, and to date most thoroughly examined, natural receptor appears to be the TCR of T lymphocytes bearing the CD8 heterodimer. Site-directed mutagenesis and crystallographic studies indicate that the $V\alpha$ and $V\beta$ domains of heterodimeric TCRs associate in a diagonal fashion across the top surface of the structure formed by MHC α_1 and α_2 (Hogan et al. 1988; Lombardi et al. 1991; Moots 1993; Tussey et al. 1994; Garboczi et al. 1996; Garcia et al.

1996; Parham 1996; Björkman 1997; Smith and Lautz 1997; Manning et al. 1998). The hypervariable CDRs of V_{α}/V_{β} contact specific regions of this interface (FIG. 5). Both precursor and effector CTLs are defined as being class I-restricted in that they are only capable of recognizing and responding to antigens displayed in the context of these molecules (Zinkernagel and Doherty 1974).

Since the antigens presented to $CD8^{+}$ T lymphocytes are predominantly obtained through the processing of intracellular proteins as previously described, class I molecules figuratively serve as external banners that advertise the inner contents of the cells. Indeed, these antigens of peptide ligands indicate to the immune system as a whole which cells are to be eliminated and/or protected. Malignancies and/or pathogens effectively use this system to camouflage their existence and thereby escape detection and elimination by $CD8^{+}$ CTLs, for example.

Thymic education of lymphocytes prevents activation in response to characteristic cell-derived peptides (Robey and Fowlkes 1994), but peptides acquired through the degradation of atypical proteins are recognized and induce cytotoxicity (Townsend et al. 1985; Gotch et al. 1988; Walker et al. 1988; Clark et al. 1995). Therefore, it is not surprising that $CD8^{+}$ T lymphocytes play a critical role in controlling and/or eliminating infected and neoplastic cells.

$CD8^{+}$ T lymphocytes are implicated in immunity to pathogens such as

viruses, which are intracellular invaders that utilize the host cell's biosynthetic machinery to produce their own foreign proteins (Yap et al. 1978; Lin and Askonas 1981; Jamieson et al. 1987; Harty and Bevan 1992; Riddell et al. 1992; Kulkarni et al. 1995; Heslop et al. 1996; Schmitz et al. 1999). CTL responses are likewise extended to include stimulation by aberrant proteins such as those associated with malignancy (Vose and Bonnard 1982; Muul et al. 1987; Coulie et al. 1992; Melief 1992; Kittlesen et al. 1998; Shichijo et al. 1998). In fact, class I molecules are capable of binding and presenting to CTLs any protein introduced into the endogenous processing pathway by either natural or artificial means (Gooding and O'Connell 1983; Moore et al. 1988; Yewdell and Bennink 1990; Bertoletti et al. 1991; Donnelly et al. 1993; Ikonomidis et al. 1994; Ballard et al. 1996; Day et al. 1997; Goletz et al. 1997; Kim et al. 1997). This knowledge serves as a motivating factor behind the development of both protein/peptide-based vaccines and other therapeutics intended to elicit protective CTL responses to microbial pathogens and other abnormalities, which otherwise remain cytoplasmically concealed from detection.

Fully understanding the role of class I molecules in ligand presentation as described above is complicated by α -chain polymorphism. Class I structural differences resulting from genetic variation confer extreme heterogeneity upon regions of the molecule that interact with peptides. The knowledge of how

polymorphism specifically impacts the natural presentation of peptide epitopes upon the cell surface is consequently limited.

Class I MHC polymorphism was first documented in mice (Gorer 1936; Gorer 1937; Nathenson et al. 1981) and next studied in humans by serology (Dausset 1958; Payne and Hackel 1961; van Rood 1969); however, first protein and then DNA sequencing studies precisely demonstrated this genetic variability to be most concentrated throughout the exons of the α_1 and α_2 heavy chain domains at positions affecting amino acid residues that line the walls and floor of the previously described peptide binding groove (Orr et al. 1979; Tragardh et al. 1979; Rojos et al. 1987; Parham et al. 1988; Parham et al. 1989; Parham et al. 1995). It is the more centrally-located binding pockets (B-E), together with specific residues within the F-pocket, that appear to be the most polymorphic (Chelvanayagam 1996; Kostyu et al. 1997). Changes in the physicochemical properties of amino acid side chains within the groove can influence the stability with which given peptides interact during the assembly of α -chain/ β_2m /peptide trimers (Matsui et al. 1993; Rohren et al. 1993; Salter 1994; Young et al. 1995). Therefore, despite the overall structural conservation illustrated among class I α -chains (Björkman and Parham 1990; Madden 1995), their peptide binding grooves can vary drastically from one allelic form to another; as a result various isoforms are capable of associating with distinct arrays of peptides (Elliott et al. 1993; Smith et al. 1995; Smith et

al. 1996b).

The characteristic polymorphism observed among class I molecules is thought to originate primarily through recombination and gene conversion (Kuhner et al. 1991; Parham et al. 1995); point mutations are believed to contribute more rarely to the pool of new alleles continually arising (Parham et al. 1989). Individuals inherit a set of three class I genes from each parent, and since their expression is codominant, a single person may therefore display up to six different HLA class I molecules upon his or her nucleated cells. From these alleles, new forms evolving progressively within populations can be passed on to subsequent generations and likewise serve as templates upon which yet further diversity may be introduced. This occurs through events such as single or double recombination (Parham et al. 1988) or nonreciprocal exchanges between *cis*-oriented gene segments during gene conversion (Parham 1992). Serological cross-reactivity studies, as well as locus-specific PCR amplification and sequencing analyses, have verified the existence of allelic subtypes, or allotypes, of closely related alleles that appear to have arisen from a common ancestral template by these molecular mechanisms (for example, Payne et al. 1978; Ooba et al. 1989; Hildebrand et al. 1994; Prilliman et al. 1996). While both inter- and intra-locus genetic events may give rise to polymorphism, the latter is most commonly observed; alleles at a locus generally tend to be more closely related to one another than to those present

at other loci (Parham et al. 1988; Parham et al. 1995). The forces driving HLA class I polymorphism are believed to be those of overdominant or balancing selection (Hughes and Nei 1988; Hughes and Yeager 1998); this is based upon values of $d_N > d_S$ within the coding regions of α_1 and α_2 for specificity determining pocket residues positioned to interact with the peptide binding groove, while the contrary ($d_S > d_N$) is observed among the remaining α_1/α_2 residues.

Considering the manner by which class I structural polymorphisms are generated and maintained demonstrates that HLA genetic variability affords both advantages and disadvantages. It is beneficial in ensuring that at least a small portion of the human population will possess class I molecules capable of: (i) binding immunogenic peptides derived from any given pathogen; (ii) presenting those peptides to CTLs; and (iii) evoking a protective immune response. In short, annihilation of the species is guarded against by molecular diversity (Parham 1992). The concept of heterozygote advantage through polymorphism as a mechanism for effectively allowing broader peptide binding abilities and thus broader CTL recognition of pathogenic peptides has been strongly emphasized from a statistical perspective (Hughes and Nei 1988; Nei and Hughes 1992). For example, HLA heterozygosity has been correlated with diminished progression to disease following HIV infection (Carrington et al. 1999). In addition, at the level of individual allotypes the "nonrandomness"

with which certain polymorphisms are maintained within populations following their evolution supports positive natural selection. This might occur in response to specific pathogenic pressures, as seen in the strong association of the West African allele HLA-B*5301 with resistance to malaria (Hill et al. 1991; Hill et al. 1992), a parasitic illness endemic to West Africa. As mentioned previously however, the polymorphic nature of class I molecules results in divergent allotypes binding and presenting different peptides. CTLs thus focus on distinct portions of any given pathogen from one individual to another. Therefore, dissecting disease susceptibilities and resistances requires a grasp of how binding groove-localized amino acid variations specifically alter ligand presentation.

Numerous previous research endeavors have been directed toward understanding the structural and functional nature of peptides bound by HLA complexes; though some progress has been made in analyzing the manner that peptide binding is specifically influenced through α_1/α_2 substitutions, this knowledge remains limited and sometimes inconsistent. The full extent that polymorphisms dictate the degrees of ligand binding ability, stringency, and/or degeneracy (and subsequently cell surface presentation) has, as a result, not been adequately resolved. Similarly, the occurrence of overlapping ligands, or identical peptides presented across the binding groove polymorphisms of multiple distinct allotypes, remains to be explored.

Ideally, characterizing functional overlaps would provide an advantage not only to explore the general effects of binding groove architecture but more specifically to understand the similarities and/or differences of what is presented to CTLs by the class I molecules of genetically diverse individuals. In the search for answers to ligand binding influences by α -chain polymorphisms, methods including pooled Edman sequencing, mass spectrometric analysis, and binding/reconstitution assays have been employed. However, each approach bears its own strengths and limitations and none so far has been significantly successful in comparatively evaluating levels of functional overlap across class I polymorphisms. The importance of understanding peptide associations with polymorphic class I molecules at a level of complexity not necessarily afforded by the currently-defined strategies is thus underscored: epitope predictions based upon methods that fail to accurately assign possibilities for natural binding groove occupancy by either aberrant or low-abundance peptides of varying binding affinities interfered with detecting presentation overlaps among various HLA class I allotypes.

Early investigations of class I peptide ligands focused on simplifying the effects of polymorphism through establishing "motifs" (Rötzschke and Falk 1991; Rammensee et al. 1993; Engelhard 1994). Motifs have typically been established by purifying surface-expressed class I molecules from detergent lysates of either transformed cells or transfectants and extracting the bound

ligands with either TFA or acetic acid. The resulting peptide pools are then subjected to consecutive cycles of N-terminal Edman degradation (Falk et al. 1991; Jardetzky et al. 1991).

The resultant motifs from N-Terminal Edman degradation are invariably nine amino acids in length and describe, as based upon relative yield increases per cycle, conserved "anchor" residues, or sites of stereochemical preferences, for peptides that are bound by a class I molecule. These anchors, typically appearing to involve both P2 and P9 of the ligands, are considered to be allele-specific and thus common among nearly all of the peptides bound by a given class I allotype. The remaining positions demonstrate no overriding amino acid preferences, although the motifs of a few molecules demonstrate anchors at P3 or P5 (Rammensee et al. 1997). The P2 anchor is assumed to associate with the B-pocket of the binding groove, while P9, the C-terminal residue assignment, associates with the F-pocket. "Auxiliary" or "secondary" anchors (alternative positions frequently defined by the occupancy of chemically similar residues) are additionally included in the motifs of some class I molecules (Rammensee et al. 1997). In general, a common interpretation of this type of data is that endogenous peptide binding and/or loading requires a nonamer with particular P2 and P9 anchors. As will be discussed, many searches (and consequent failed attempts) for putative viral or tumor class I-presented epitopes have subsequently been predicated upon nonameric templates with P2

and P9 anchor assignments.

Broad efforts have been focused upon establishing and analyzing motifs from natural class I ligands (for example, Huczko et al. 1993; Fleischhauer et al. 1994; Kubo et al. 1994; Barber et al. 1995; Steinle et al. 1995; Barber et al. 1996; Tzeng et al. 1996; Tieng et al. 1997; Yagüe et al. 1998). However, motifs fail to reflect the true complexity of peptides presented by divergent class I molecules. Drawbacks are evident in that numerous characterized peptide sequences that bind class I are greater than 9 residues long, with 14 being the largest identified to date (Engelhard 1994); The binding of longer ligands likely results from stable associations at anchor positions coupled with central protrusion of the peptide outward from the groove (Fremont et al. 1992; Guo et al. 1992; Madden et al. 1993). Furthermore, other examinations of specific peptides naturally presented by class I MHC of both humans and mice have indicated that some fail to comply with their respectively defined motif anchors (Calin-Laurens et al. 1993; Henderson et al. 1993; Sadovnikova et al. 1993; Kawakami et al. 1994; Urban et al. 1994; Malarkannan et al. 1996; Mata et al. 1998), thus suggesting that both length variance and "nonanchor" residues in the peptide could play significantly more prominent roles in binding than strictly accounted for by a given pooled motif alone. Studies have also indicated that low copy-number peptides, presented by only a small proportion of the total class I molecules expressed upon the cell surface, can successfully

elicit CTL responses (Cox et al. 1994; Malarkannan et al. 1996; Wang et al. 1997). Issues such as these clearly reflect the limitations posed in applying Edman sequencing to the complex mixtures of peptides extracted from class I molecules (Stevanovic and Jung 1993). As a result, pooled Edman sequencing is therefore unable either to precisely characterize individual ligands or to effectively identify overlaps in ligand presentation.

Shortly after the debut of examining class I ligands by pooled Edman sequencing, the first reports of ligand analyses via mass spectrometric sequencing of organic ions from similarly-prepared class I ligand extracts began to emerge (Henderson et al. 1992; Hunt et al. 1992; Huczko et al. 1993; Appella et al. 1995). The utilization of MS/MS on a triple quadrupole mass spectrometer provided for the precise characterization of individual constituents at sub-picomolar levels from pools of class I peptides, in contrast to the picomolar detection limits of Edman analysis. Furthermore, LC/MS prior to this step allowed for complexity estimates to be made. For example, based upon the quantitation of 200 different peptides present within HLA-A*0201 extracts, extrapolation with regard to the peptides detected versus their respective contributions to the mass spectrometric TIC obtained indicated that at least 1,000 and perhaps as many as 10,000 unique peptides are bound by this class I allotype (Hunt et al. 1992). The ability to characterize ligands as such additionally assisted in being able to identify and sequence single specific

epitopes from RP-HPLC fractions demonstrating biological activity via CTL assays (Rötzschke et al. 1990; Henderson et al. 1993; Kawakami et al. 1994; Skipper et al. 1996; Simmons et al. 1997; Wang et al. 1997; Hogan et al. 1998; Paradela et al. 1998). Examination of ligands by mass spectrometry was therefore an effective development in both starting to fill the gaps often present in pooled motifs and expediting the classification of ligands potentially bearing immunological significance.

However, the routine application of mass spectrometric techniques to class I ligand examination has remained relatively isolated; it is practiced in only a handful of laboratories (for example, Woods et al. 1995; Simmons et al. 1997; Flad et al. 1998; van der Heeft et al. 1998; Yagüe et al. 1998). This appears largely due to the inherent difficulties imposed in handling the small quantities of peptides extracted for study (Henderson et al. 1993; Appella et al. 1995; van der Heeft et al. 1998) as well as the notably tedious nature of the subsequent data processing (Papayannopoulos 1995; van der Heeft et al. 1998). Other issues concern the specific instrumentation and its mode of operation, since: (i) the ability to consistently identify peptides from particular extracts relies upon dependable RP-HPLC gradients for separation; and (ii) the possible ionization and/or detection interfaces function in distinct manners that can influence the data ultimately acquired (Chapman 1996; Watson 1997). In summary, understanding the impact of polymorphism upon the binding of

endogenous peptides has historically been limited by small amounts of ligands available for analyses.

What has not been accomplished yet by mass spectrometry is the systematic definition of peptides across diverse class I allotypes. Various *in vitro* assays have been developed to complement mass spectrometric approaches. The assays are performed using a number of different protocols with the common theme of assessing the relative abilities *in vitro* of synthetically defined peptides to associate with specific class I α -chains and β_2m . In the case of binding assays, synthetic peptides as well as a peptide standard are incubated with purified class I complexes and tested in their capacities to competitively displace the ligands already bound (Chen and Parham 1989; Ruppert et al. 1993; Sette et al. 1994). This has likewise been performed by stripping ligands from class I complexes expressed on the cell surface by acid treatment and then incubating the cells with the synthetic peptides (Drijfhout et al. 1995). Another common approach to competitive binding assays involves FACS analysis with class I-specific MAbs after incubation of synthetic peptides with various cell lines including RMA-S (Townsend et al. 1989), T2 (Salter et al. 1985), Hmy2.C1R (Storkus et al. 1989), or 721.221 (Kavathas et al. 1980) transfected to express HLA α -chains (Huczko et al. 1993; Takamiya et al. 1994; Zeh III et al. 1994; Boisgerault et al. 1996; Konya et al. 1997). Alternatively, *de novo* reconstitution assays can be performed by

incubating synthetic peptides with free α -chains and β_2m and comparing the resultant quantities of free versus complexed α -chains (Tanigaki 1992; Fruci et al. 1993). Other methods of reconstitution have also been applied (Silver et al. 1991; Parker et al. 1994; Gnjjatic et al. 1995; Robbins et al. 1995; Tan et al. 1997).

While these *in vitro* assays with synthetic ligands have led to the development of extensive "supermotifs", or enhanced motifs that ascribe binding preferences to particular class I allotypes or groups of related allotypes (del Guercio et al. 1995; Sidney et al. 1996a; Sidney et al. 1996b), they present obvious limitations. A technical concern is that binding/reconstitution determinations are often based upon either "standard" probe peptides for each allotype examined or mean experimental values that can significantly differ from one laboratory or experiment to another. A specific illustration is based on the results generated by three different groups testing HLA-A*0201 with panels of synthetic peptides; the assignment of residues preferred or deleterious for binding at given positions varied widely (Ruppert et al. 1993; Parker et al. 1994; Drijfhout et al. 1995). In fact, only three residue similarities carried across all three studies.

Ruppert and colleagues and Drijfhout and colleagues each employed competitive binding assays; however, the former involved assessing the ability of synthetic peptides to inhibit binding of a radioiodinated standard peptide to

membrane-extracted class I complexes, while the latter involved stripping peptides from class I complexes on B-LCLs and determining by FACS analysis the ability of synthetic peptides to inhibit binding of a different fluorescence-labelled standard. On the other hand, Parker and colleagues employed a reconstitution assay which measured the incorporation of radiolodinated β_2m into complexes refolded using synthetic peptides and α -chains prepared from *Escherichia coli* inclusion bodies. Of perhaps greater significance, these types of assays fail to account for the processing/loading physiology of trimolecular complex formation within the cell (Hogan et al. 1988); indeed, differences have been documented in performing comparative examinations by pooled Edman sequencing and mass spectrometry upon naturally-extracted versus artificially bound peptides (Davenport et al. 1997). In one specific case, an immunodominant HIV peptide divulged through other mechanisms completely failed to demonstrate binding to its restricting allotype via an *in vitro* assay (Tsomides et al. 1991). Conclusions regarding ligand presentation that are drawn from binding/reconstitution assays thus cannot be applied generally without caveat.

Thus, the present invention(s) aim to provide a methodology for the production of soluble MHC Class I and II molecules from either gDNA or cDNA starting material such that the structural and functional impact of HLA class I polymorphism on peptide binding can be assessed and, in particular, to test

how natural ligand presentation overlaps exist in varying degrees across the polymorphisms of divergent class I binding grooves. Furthermore, the soluble MHC molecules can be used in the functional testing of CTL binding assays and vaccine development. Utilizing this specification, one of ordinary skill in the art will be able to: (i) generate ligands and hence ligand maps from the peptide pools extracted from series of distinct yet related class I HLA-B*15 allotypes; (ii) compare the different ligand maps to identify potentially shared elements; and (iii) characterize the elements identified to positively or negatively validate the occurrence of overlapping ligands. One of ordinary skill in the art, given the present specification, will also realize that the ability to produce soluble MHC molecules from either gDNA or cDNA starting material also will allow for other useful assay and vaccine development such as the functional testing of peptide ligands of interest to determine if, when, and how such peptide ligands provoke and/or stimulate an immune response. All of which is directed toward the goal of identifying candidate peptide ligands that may be used singly or together as a vaccine and/or immune system primer.

BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWINGS

FIG. 1 is a graphical representation of MHC class I location and heavy chain coding region. (A) Simple map of the human MHC region, specifically highlighting the B, C, and A loci that encode the class I heavy chains as simplified from (Janeway and Travers 1994). Genetic distances are estimated in kilobases. Other genes (not shown) including heavy chains and transporter/chaperone proteins (class II) and complement proteins and cytokines (class III) are encoded within the remaining MHC regions. (B) The basic exon structure of MHC class I transcripts. A total of seven exons encode the leader peptide (1), α_1 domain (2), α_2 domain (3), α_3 domain (4), transmembrane domain (5), and cytoplasmic domains (6-7). Specific exon regions are indicated in parentheses for *HLA-B*15011* (accession number U03859, Hildebrand et al. 1994), which encodes the HLA-B*1501 heavy chain.

FIG. 2 is a three-dimensional graphical representation of the configuration of the MHC class I extracellular domains. (A) The complete extracellular portion of the class I molecule HLA-B*3501, revealing the three domains (α_1 , α_2 , and α_3) of the heavy chain as well as the noncovalently associated light chain, β_2m . Top (B) and side (C) views of the basket-like antigen binding groove formed by α_1 and α_2 ; the α -helix and β -sheets for each domain are respectively indicated in (B). These three images were rendered from the HLA-B*3501 Protein Data Bank (Brookhaven National Laboratory) crystal structure coordinate file, 1A1N

(Smith et al. 1996a), using the Swiss-PdbViewer 3.01 software (Guex and Peitsch 1996). The CHO linked to α_1 residue 86 is not shown.

FIG. 3 is a graphical representation of the specificity determining pockets of the MHC class I ligand binding groove. The structural residues contributing to the six pockets (A through F) of the antigen binding groove formed by α_1 and α_2 are indicated in black and numbered from the N terminus of the mature class I α -chain. The residues are as collectively defined in the literature (Saper et al. 1991; Matsumura et al. 1992; Chelvanayagam 1996). In terms of a nonameric ligand, the pockets are assumed to accommodate the amino acids occurring at given ligand positions as follows: A-pocket, P1; B-pocket, P2; C-pocket, P6; D-pocket, P3; E-pocket, P7; and F-pocket, P9. The majority of residues that contribute to these binding pockets are oriented such that they are solvent-inaccessible in mature trimers.

FIG. 4 is a pictorial representation summarizing MHC class I trimer assembly and transport. The typical endogenous processing pathway shown is simplified from the description disclosed herein. Major proteins participating in the processes are specifically illustrated and labelled. Some alternative processing and/or transport mechanisms currently under investigation are designated in the diagram by question marks. Assembled trimers travel through the Golgi complex prior to vesicular delivery to the surface membrane of the cell (not shown).

FIG. 5 is a schematic representation of the deduced contact region between the MHC class I α_1/α_2 and TCR V_α/V_β interfaces. From the diagonal orientation of the estimated binding footprints of CDRs 1, 2, and 3 (numbered) for each of TCR- V_α and V_β with respect to the top of MHC- α_1 and α_2 , the general interaction between TCR and MHC appears to predominantly involve V_α/V_β contact with both the class I solvent-accessible helical residues and the central portions of the bound ligand (not shown).

FIG. 6 is a flow diagram of the overall strategy for comparatively mapping and characterizing peptide ligands presented by class I HLA. The approach taken to address the presence of overlapping ligands across diverse class I molecules consisted of three basic parts: (A) sHLA-producing transfectant establishment and culture; (B) extraction, purification, and separation of ligands; and (C) ion map generation/comparison and characterization of individual peptides. Though only two molecules are indicated for simplification, numerous additional molecules (as indicated in the specification) were simultaneously carried through the steps shown.

FIG. 7. Hypothesized evolutionary relationships of HLA-B15 allotypes according to expressed α -chain polymorphisms. Shown in this scheme are the 46 allotypes whose amino acid sequences are provided in herein. Solid lines indicate single mutagenic events separating given allotypes. Dashed lines indicate that greater than one mutagenic event separates given allotypes;

intermediaries are indicated by question marks unless more specifically suspected (as in the case of a probable B*1539/B17 recombinant). None of the line lengths are reflective of mutational rates.

FIG. 8 is a graphical representation of the localization of antigen binding groove substitutions distinguishing the B*1512, B*1508, B*1501, B*1503, B*1518, and B*1510 allotypes. The structural residues of the antigen binding groove formed by α_1 and α_2 which differ among the six alleles according to Table 1 are indicated on the ribbon diagram in black and numbered from the N terminus of the mature class I α -chain.

FIG. 9 is a graphical representation summarizing of the Unisyn Technologies CP-3000 basic flow path. The system was assembled and operated as described herein. Arrows indicate unidirectional media flow.

FIG. 10 is two graphs showing sHLA production during bioreactor runs with two different B*1501 transfectants. Constructs using PSR $_{\alpha-neo}$ and pcDNA3 for producing soluble B*1501 were separately transfected into B-LCL 721.221; subclones were then cultured in CP-3000 systems as described herein. Harvests samples drawn on the days indicated along the x-axis during each run were subjected to ELISA, yielding the representative sHLA production data shown.

FIG. 11 is a graph showing RP-HPLC fractionation of peptides extracted from B*1510. The UV trace obtained during separation as described herein of

approximately 400 µg of peptides extracted from B*1510 reveals the bulk of absorbance to occur along the gradient (black line) between 10-40% buffer B (indicated at the right). The three peaks associated with the control dye (methyl violet base B), as well as the primary region subjected to intensive mapping between B*1501, B*1503, B*1508, and B*1510 peptides (fractions 6-19), are indicated.

FIG. 12 is a schematic showing the generalized components of a triple quadrupole mass spectrometer. The basic constituents of the system described herein include: (A) an electrospray source/ionization interface for sample introduction; (B) three quadrupoles for ion manipulation, which includes mass filtration (Q1 or Q3), transmission (all quadrupoles), and/or collision (q2); and (C) a detector for amplifying transmitted ion signals so that they can be recorded and analyzed.

FIG. 13 includes two graphs (A and B) and chart C that show the identification and verification of a ligand overlap across divergent HLA-B15 molecules. NanoES-MS spectral ion maps obtained individually from RP-HPLC fraction 8 for each of B*1501, B*1503, B*1508, and B*1510 were aligned for comparison; an expanded view of the range 495-555 m/z, or amu, is shown in (A). The ion mass centered at 517.2 m/z matched across the spectra of B*1501, B*1503, and B*1508 (top three panels) but not B*1510 (bottom panel). This ion was subsequently selected for NanoES-MS/MS from fraction 8

of the first three class I MHC molecules. The homologous spectra resulting from fragmentation of this $[M+2H]^{2+}$ ion (B) classified the peptide as a positive match, or ligand overlap, occurring across B*1501, B*1503, and B*1508 and allowed for primary sequence derivation (C). N- and C-terminal peptide fragments present in all three NanoES-MS/MS spectra are labelled according to standard nomenclature (Roepstorff and Fohlman 1984) in the top panel of (B) and underlined in (C); immonium ions are indicated by their single-letter amino acid codes in (B), and the sequences of internal cleavage products are also specified.

FIG. 14 is a graphical representation showing the reproducibility of the class I HLA ligand mapping and characterization strategy disclosed herein. Approximately 400 μ g each of B*1508 ligands obtained from two separate bioreactor runs performed six months apart were fractionated by RP-HPLC as shown for B*1510 (FIG. 11). NanoES-MS ion mapping and comparison were then performed as described herein. As illustrated by the spectra of fraction 15 for each (A and B), the ion maps were consistent with one another; subjecting an ion (514.0) from each B*1508 fraction to NanoES-MS/MS further demonstrated reliability of the protocols employed.

FIG. 15 is a tabulation of the pooled Edman sequencing motifs for peptides extracted from B*1501 purified by two different MAbs. Data derived from the eluates of B*1501 complexes purified using either W6/32 (A) or BBM.1

(B) indicated amino acid residues that had been previously attributed to the corresponding positions in B*1501 motifs (solid characters), as well as a residue which was not included in any previous motif descriptions (Falk et al. 1995; Barber et al. 1996). Edman degradation was carried out as described hereinafter, and relative picomolar fold increases are indicated in parentheses to the right of each residue; amino acids demonstrating a 2.0 to 3.5-fold picomolar increase over the previous degradation cycle were grouped as "strong", while those demonstrating an increase of greater than 3.5-fold were grouped as "dominant". Dashes indicate that no distinct residues could be detected as either dominant or strong at the given positions.

FIG. 16 is a tabulation and corresponding pictorial representation of pooled Edman sequencing motifs for peptides extracted from B*1508, B*1512, B*1503, B*1518, and B*1510. Edman degradation was carried out as described herein, and the data were analyzed as described hereinbefore with respect to FIG. 15. Ribbon diagrams of the class I antigen binding groove to the right of each motif show the structural residues of the α_1/α_2 antigen binding groove among each of the alleles which differ with respect to B*1501 according to Table 1; they are indicated in black and numbered from the N terminus of the mature class I α -chain.

FIG. 17 is a graphical showing RP-HPLC separation of ligands from BBM.1-purified B*1501. The UV trace obtained during separation as described herein

of approximately 150 µg of peptides extracted from BBM.1-purified B*1501 reveals the bulk of absorbance to occur along the gradient (black line) between 10-30% buffer B (indicated at the right). The fractions collected are numbered; although all fractions were examined, only those subsequently selected for analysis and data presentation in FIGS. 18 and 19 are marked by dots. It is noted that the chosen fractions were distributed evenly across the entire region of interest and included a wide variety that were of high as well as low UV absorbance and/or resolution.

FIG. 18 is four graphs showing the percentages of RP-HPLC ligand fractions from BBM.1-purified B*1501 demonstrating particular amino acid occupancies by Edman degradation. Sequence complexity among the peptides was summarized by averaging the frequency of occurrence for amino acid residues at P2 through up to P12 for the fractions marked by dots in FIG. 17. Amino acids are grouped along the x-axis according to their physicochemical natures (charged, polar, or hydrophobic), as specifically designated in the chart for P10 and indicated within all four charts by differential shading. Residues observed at P2 and P9 in former B*1501 motif descriptions are indicated in bold italics. Since it was not derivatized, cysteine was undetectable and is therefore excluded.

FIG. 19 is four tabular examples of Edman sequence complexity among RP-HPLC ligand fractions from BBM.1-purified B*1501. While each of the

fractions marked by dots in FIG. 15 was subjected to Edman sequencing, the results obtained from fractions 10, 15, 28, and 31 are shown here to specifically illustrate some of the diverse trends observed among constituents of the ligand mixture. Dominant and strong assignments were made as described hereinbefore for FIG. 13; weak assignments were made for amino acids demonstrating a 1.5 to less than 2.0-fold picomolar increase over previous cycles. No assignments are shown for P1 due to lower confidence since comparison with former cycles was impossible. The positional assignments in fraction 15, which are identified in bold italics, appear to correspond to a hexamer, IAVGYV, derived from HLA class I α -chain₂₃₋₂₈ (Prilliman et al. 1998).

FIG. 20 is five graphs summarizing the length diversity among the 449 characterized HLA-B15 ligands. Graphed data from the ligands listed in Tables A-E summarizes length diversity among the peptides respectively characterized from B*1501, B*1503, B*1508, B*1510, and B*1512.

FIG. 21 is two graphs showing N- and C-regional diversity observed through alignments of B*1501 ligands. Frequency of occurrence for amino acids at the first (A) and final (B) four positions among the ligands characterized from B*1501 (Table A). The graphs were generated from separate N- and C-terminal data alignments.

FIG. 22 is two graphs showing N- and C-regional diversity observed through alignments of B*1503 ligands. Frequency of occurrence for amino

acids at the first (A) and final (B) four positions among the ligands characterized from B*1503 (Table B). The graphs were generated from separate N- and C-terminal data alignments.

FIG. 23 is two graphs showing N- and C-regional diversity observed through alignments of B*1508 ligands. Frequency of occurrence for amino acids at the first (A) and final (B) four positions among the ligands characterized from B*1508 (Table C). The graphs were generated from separate N- and C-terminal data alignments.

FIG. 24 is two graphs showing N- and C-regional diversity observed through alignments of B*1510 ligands. Frequency of occurrence for amino acids at the first (A) and final (B) four positions among the ligands characterized from B*1510 (Table D). The graphs were generated from separate N- and C-terminal data alignments.

FIG. 25 is two graphs showing N- and C-regional diversity observed through alignments of B*1512 ligands. Frequency of occurrence for amino acids at the first (A) and final (B) four positions among the ligands characterized from B*1512 (Table E). The graphs were generated from separate N- and C-terminal data alignments. The information shown here for B*1512 ligands is skewed in relation to that obtained for ligands from the other four allotypes since (i) a comparatively fewer B*1512 peptides were sequenced, and (ii) over half of the B*1512 peptides characterized were specifically

investigated as ion map differences with B*1501.

FIG. 26 is a graphical and tabular representation showing ligand overlaps identified by NanoES-MS mapping and characterized by NanoES-MS/MS during comparative analysis of B*1508, B*1501, B*1503, and B*1510 extracts. Ribbon diagrams of the class I antigen binding groove indicate residue substitutions (black, numbered from the first residue of the mature α -chain) between each B15 molecule with respect to B*1501. The peptides are categorized into three different groups from top to bottom as follows: ligands that overlap B*1508 and B*1501; ligands that overlap B*1508, B*1501, and B*1503; and ligands that overlap B*1501 and B*1503. Dashes represent positions at which amino acids could not be unambiguously assigned through the NanoES-MS/MS fragmentation patterns and/or Edman data obtained. Ligand residues that coincide with dominant and strong amino acids for the given motifs (FIGS. 15 and 16) are indicated in bold.

FIG. 27 is a pictorial representation of the proposed N-proximal and C-terminal anchoring of a nonamer overlapping B*1508, B*1501, and B*1503. The shared C-terminal anchoring preferences for Tyr in the NQZHGSAEY ligand among B*1508, B*1501, and B*1503 as defined by their respective motifs (FIGS. 15 and 16) are shaded black, while the varied N-proximal anchoring preferences likewise reflected in the motifs are shaded gray. Ligand residues are numbered sequentially from the N terminus.

FIG. 28 is a pictorial schematic summarization the summary of structure-function relationships among the HLA-B15 allotypes from which ligands were characterized. HLA-B15 allotypes for which a motif is known are indicated in bold, while allotypes with currently undefined motifs are indicated in italics. Arrows are representative of single mutagenic events (which are specifically defined for each arrow); bold arrows emphasize evolutionary pathways between structures for which motifs have been described either here or elsewhere. P2 and P9 designations are listed. Tentative designations have been made for the italicized allotypes by extrapolation from known motifs in light of the polymorphisms present. P2/P9 specificities for B*1519 are based upon B*1512. P2/P9 specificities for B*1539 are based upon what has been observed between B*4402 and B*4403 (Fleischhauer et al. 1994). P2/P9 specificities for B*1529 are based upon B*1508. P2/P9 specificities for B*1523 are based upon what has been observed between B*1502 and B*1513 (Table 7), as well as observations between B*0801 and B*0802 (Arnett et al. 1998). P2/P9 specificities for B*1547 are based upon the relationship previously discussed between B*1503 and B*4801 (Martinez-Naves et al. 1997). P2/P9 specificities for B*1537 are based upon B*1402 (DiBrino et al. 1994). The three sections (A, B, and C) into which the molecules have been divided are discussed in the text. For reference, a ribbon diagram of the class I antigen binding groove indicating residue substitutions (black, numbered from the first

residue of the mature α -chain) between the molecules with respect to B*1501 is provided.

FIG. 29 is a pictorial representation of the PCR strategy for the production of soluble HLA from gDNA according to the methods of the present invention.

FIG. 30 is a gel image of the primary PCR of 3A394.

FIG. 31 is a gel image of the secondary PCR of 3A394.

FIG. 32 is a gel image showing 3A394 and pcDNA3.1 digested with EcoR I and Xba I.

FIG. 33 is a gel image showing the restriction digests of 3A394 clones.

FIG. 34 is a graph showing the comparative binding of three monoclonal antibodies to four soluble HLA molecules.

FIG. 35 is a pictorial representation of the MHC.

FIG. 36 is a pictorial representation of an HLA molecule.

FIG. 37 is a pictorial representation of the HLA binding groove that binds antigenic peptides.

FIG. 38 is a pictorial representation of the antigen processing and assembly of the MHC class I/peptide complex.

FIG. 39 is a pictorial representation of HLA peptide loading and movement of the HLA molecule to the cell surface.

FIG. 40 is a pictorial representation of a tetramer biotinylated sHLA complex of the present invention.

FIG. 41 is a pictorial representation of a bacterial expression vector encoding a biotinylation substrate peptide sequence.

FIG. 42 is a pictorial representation of the biotinylation of sHLA.

FIG.43 is a pictorial representation of the conjugate used to confirm biotinylation of the sHLA molecule.

FIG. 44 is a graph showing production of sHLA-B* 0702 by T₂ transfectants after peptide pulsing.

FIG. 45 is a graph showing the elution curve of the Elisa assay used to confirm sHLA production.

FIG.46 is a graph showing that an increase in time results in greater biotinylation.

FIG. 47 is a graph showing separation of biotinylated class I from free biotin.

BRIEF DESCRIPTION OF THE TABLES

Tables 1-24 and Tables A-E are set forth below on Pages 156-192.

Table 1. Amino acid differences among the B*1512, B*1508, B*1501, B*1503, B*1518, and B*1510 allotypes. Residues, noted across the top of the table, are numbered from the N terminus of the mature class I heavy chain for each; the 24 amino acid leader sequence is therefore not included. The residue positions are differentially highlighted in black (α_1) or gray (α_2). Positions of identity with the consensus sequence (italicized) drawn from B*1501 are indicated by dashes.

Table 2. PCR and sequencing primers for creating and verifying sHLA

constructs. Designated restriction enzyme cut sites are underlined on the 5' and 3' PCR primers, and the regions of the 3' PCR primers that inserted a stop codonn at position 300 are shown in bold italics. Sequencing primers were Cy5-labelled; the regions that they either sequenced through or hybridized with are indicated in parentheses.

Table 3. Additional P2, P9, and length variability revealed through Edman sequencing of RP-HPLC ligand fractions from W6/32-purified B*1501, B*1508, B*1503, and B*1510. The number of fractions for each which were subjected to Edman degradation as described herein and shown in FIG. 13 are shown; ligands present in the fractions from all four molecules demonstrated evidence of (i) expanded P2 and P9 occupancy, and (ii) sequence beyond the 9 residues described by their respective motifs (as shown in FIGS. 15 and 16).

Table 4. HLA-B15 ligands identically matching database source proteins, by category. The ligands are categorized according to source protein functions. Residues for the specific ligands are numbered from the initiating Met of their respective source proteins. Ligands from HLA-B15 molecules not studied here (B*1502, B*1509, and B*4601) are referenced herein.

Table 5. Ligands unique to B*1512, as compared with B*1501. Positively-charged P1 residues are indicated in bold. Dashes represent positions at which amino acids could not be unambiguously assigned through the NanoES-MS/MS fragmentation patterns and/or Edman data obtained.

Underlined residues designate tentative assignments.

Table 6. Ligand binding groove residues interacting with the B-pocket among HLA-B15 allotypes bearing known Edman-derived motifs. Residues, noted across the top of the table, are numbered from the N terminus of the mature class I heavy chain for each. The residue positions are differentially highlighted in black (α_1) or gray (α_2); vertical hatching immediately below further distinguishes those residues located on a helix. The molecules are grouped according to dominant/strong P2 motif similarities. Structural residues specifically discussed in the text are indicated in bold italics.

Table 7. Ligand binding groove residues interacting with the F-pocket among HLA-B15 allotypes bearing known Edman-derived motifs. Residues, noted across the top of the table, are numbered from the N terminus of the mature class I heavy chain for each. The residue positions are differentially highlighted in black (α_1) or gray (α_2); vertical hatching immediately below further distinguishes those residues located on a helix. The molecules are grouped according to dominant/strong P9 motif similarities. Structural residues specifically discussed herein are indicated in bold italics.

Table 8. B*1510 ligands exhibiting Pro with Ala and/or Val in various C-proximal combinations. C-proximal occurrences of Pro, Ala, and Val are indicated in bold. Dashes represent positions at which amino acids could not be unambiguously assigned through the NanoES-MS/MS fragmentation patterns.

and/or Edman data obtained. Underlined residues designate tentative assignments.

Table 9. B*1501 ligands: N- and C-regional occupancies observed at >10% among 126 ligands. The positional occupancy percentages for residues among the ligands from Table A greater than 10% are specifically shown; dashes indicate occupancy rates below 10% for the given side chains. N-regional positions and N_{sum} values are highlighted in black, while C-regional positions and C_{sum} values are highlighted in gray.

Table 10. B*1503 ligands: N- and C-regional occupancies observed at >10% among 74 ligands. The positional occupancy percentages for residues among the ligands from Table B greater than 10% are specifically shown; dashes indicate occupancy rates below 10% for the given side chains. N-regional positions and N_{sum} values are highlighted in black, while C-regional positions and C_{sum} values are highlighted in gray.

Table 11. B*1508 ligands: N- and C-regional occupancies observed at >10% among 96 ligands. The positional occupancy percentages for residues among the ligands from Table C greater than 10% are specifically shown; dashes indicate occupancy rates below 10% for the given side chains. N-regional positions and N_{sum} values are highlighted in black, while C-regional positions and C_{sum} values are highlighted in gray.

Table 12. B*1510 ligands: N- and C-regional occupancies observed at

>10% among 123 ligands. The positional occupancy percentages for residues among the ligands from Table D greater than 10% are specifically shown; dashes indicate occupancy rates below 10% for the given side chains. N-regional positions and N_{sum} values are highlighted in black, while C-regional positions and C_{sum} values are highlighted in gray.

Table 13. B*1512 ligands: N- and C-regional occupancies observed at >10% among 30 ligands. The positional occupancy percentages for residues among the ligands from Table E greater than 10% are specifically shown; dashes indicate occupancy rates below 10% for the given side chains. N-regional positions and N_{sum} values are highlighted in black, while C-regional positions and C_{sum} values are highlighted in gray.

Table 14. Overlaps identified through RP-HPLC/NanoES-MS ligand mass mapping and characterized by NanoES-MS/MS. Dashes represent positions at which amino acids could not be unambiguously assigned through the NanoES-MS/MS fragmentation patterns and/or Edman data obtained. Underlined residues designate tentative assignments.

Table 15. Overlap frequencies observed between B*1501 and B*1512, B*1508, B*1503, or B*1510. The frequencies of ligand presentation overlap for the molecules listed with respect to B*1501 were determined from the number of total matching ion masses subjected to NanoES-MS/MS and the number of those ions collided analyzed by NanoES-MS/MS confirmed as actual

overlaps. The resulting values are conservative since not all ions selected for examination successfully yielded fragments that could be used for evaluating the sequence positivity of mass matches.

Table 16. Potential B*1501 epitopes selected from the EBV gp85 structural antigen. The EBV gp85 protein (accession number 1334905, Arrand et al. 1981) was manually scanned for epitopes with consideration to B*1501. In the first column, epitope candidates matching the length and sequence constraints of the B*1501 pooled motif (FIG. 15) are listed. In the second column, epitope candidates matching the B*1501 motif-prescribed P2 and C-terminal occupancies but demonstrating relaxed length constraints (7 to 11 residues, according to the B*1501 panel in FIG. 20) are listed. In the third column, epitope candidates matching the B*1501 motif-prescribed nonameric length but demonstrating P2 flexibility (according to FIG. 21A) are listed.

Table 17 PCR of sHLA from gDNA primers.

Table 18 clone sequencing 5'CYS primers.

Table 19 optical density readings and concentration of DNA extracted from sample 3A394.

Table 20 optical density readings of positive clones.

Table 21 optical density readings of AF/102 plasmid extracted.

Table 22 viability of cells after two (2) days.

Table 23 Elisa results for positive transfectants exhibiting G418

resistance.

Table 24 HLA types and sHLA molecules produced.

Table A derived peptide ligands from B*1501.

Table B derived peptide ligands from B*1503.

Table C derived peptide ligands from B*1508.

Table D derived peptide ligands from B*1510.

Table E derived peptide ligands from B*1512.

DETAILED DESCRIPTION OF THE INVENTION

Before explaining at least one embodiment of the invention in detail by way of exemplary drawings, experimentation, results, and laboratory procedures, it is to be understood that the invention is not limited in its application to the details of construction and the arrangement of the components set forth in the following description or illustrated in the drawings, experimentation and/or results. The invention is capable of other embodiments or of being practiced or carried out in various ways. Also, it is to be understood that the phraseology and terminology employed herein is for the purpose of description and should not be regarded as limiting.

As described hereinabove, characterizing naturally processed HLA class I and class II ligands is a key element behind the basic understanding of how polymorphism impacts ligand presentation. However, technical and scientific challenges including both extreme sample heterogeneity and limited sample sizes complicate such examinations. Thousands of distinct peptides are present within a ligand extract prepared from a single type of class I molecule, and the immunoprecipitation/extraction protocols typically employed to recover peptide ligands yield sparse quantities on the order of ~20 μg (Hunt et al. 1992; Henderson et al. 1993). These factors often require specialized biochemical expertise not necessarily available in either the common laboratory or core facility.

Class I major histocompatibility complex (MHC) molecules, designated HLA class I in humans, bind and display peptide antigens upon the cell surface. The peptides they present are derived from either normal endogenous proteins ("self") or foreign proteins ("nonself"), such as products of malignant transformation or intracellular pathogens such as viruses. In this manner, class I molecules convey information regarding the internal fitness of a cell to immune effector cells including but not limited to CD8⁺ cytotoxic T lymphocytes (CTLs), which are activated upon interaction with "nonself" peptides and which lyse or kill the cell presenting such "nonself" peptides.

Class II MHC molecules, designated HLA class II in humans, also bind and display peptide antigens upon the cell surface. However, unlike class I MHC molecules which are expressed on virtually all nucleated cells, class II MHC molecules are normally confined to specialized cells, such as B lymphocytes, macrophages, dendritic cells, and other antigen presenting cells which take up foreign antigens from the extracellular fluid via an endocytic pathway. Therefore, the peptides they bind and present are derived from extracellular foreign antigens, such as products of bacteria that multiply outside of cells, wherein such products include protein toxins secreted by the bacteria that have deleterious and even lethal effects on the host. In this manner, class II molecules convey information regarding the fitness of the extracellular space in the vicinity of the cell displaying the class II molecule to immune effector

cells including but not limited to CD4⁺ helper T cells, which help eliminate such pathogens both by helping B cells make antibodies against microbes as well as toxins produced by such microbes and by activating macrophages to destroy ingested microbes.

Class I and class II HLA molecules exhibit extensive polymorphism, which is generated by systematic recombinatorial and point mutation events; as such, hundreds of different HLA types exist throughout the world's population, resulting in a large immunological diversity among the population. Such extensive HLA diversity in the population results in tissue or organ transplant rejection between individuals as well as differing susceptibilities and/or resistances to infectious diseases. HLA molecules also contribute significantly to autoimmunity and cancer. Because HLA molecules mediate most, if not all, adaptive immune responses, HLA proteins are needed to study transplantation, autoimmunity, and for developing vaccines.

There are several applications in which purified, individual class I and class II MHC proteins would be highly useful. Such applications include using MHC-peptide multimers as immunodiagnostic reagents for disease resistance/autoimmunity; assessing the binding of potentially therapeutic peptides; elution of peptides from MHC molecules to identify vaccine candidates; screening transplant patients for preformed MHC specific antibodies; and removal of anti-HLA antibodies from a patient. Since every

individual has different MHC molecules, the testing of numerous individual MHC molecules is a prerequisite for understanding differences in disease susceptibility between individuals. Therefore, purified MHC molecules representative of the hundreds of different HLA types existing throughout the world's population are highly desirable for unraveling disease susceptibilities and resistances and for designing therapeutics.

Currently there is no readily available source of individual HLA molecules. Until now, the quantities of HLA protein available were small and typically consist of a mixture of different HLA molecules. Production of HLA molecules traditionally involves growth and lysis of cells expressing multiple HLA molecules. Ninety percent of the population is heterozygous at each of the HLA loci; codominant expression results in multiple HLA proteins expressed at each HLA locus. To purify native class I or class II molecules from mammalian cells requires time-consuming and cumbersome purification methods, and since each cell typically expresses multiple surface-bound HLA class I or class II molecules, HLA purification results in a mixture of many different HLA class I or class II molecules. When performing experiments using such a mixture of HLA molecules or performing experiments using a cell having multiple surface-bound HLA molecules, interpretation of results cannot *directly* distinguish between the different HLA molecules, and one cannot be certain that any particular HLA molecule is responsible for a given result. Therefore, a need exists in the art

for a method of producing substantial quantities of individual HLA class I or class II molecules so that they can be readily purified and isolated independent of other HLA class I or class II molecules. Such individual HLA molecules, when provided in sufficient quantity and purity, would provide a powerful tool for studying and measuring immune responses.

The present invention envisions a method of producing MHC molecules which are secreted from mammalian cells in a bioreactor unit. Substantial quantities of individual MHC molecules are obtained by modifying class I or class II molecules so they are secreted. Secretion of soluble MHC molecules overcomes the disadvantages and defects of the prior art in relation to the quantity and purity of MHC molecules produced. Problems of quantity are overcome because the cells producing the MHC do not need to be detergent lysed or killed in order to obtain the MHC molecule. In this way the cells producing secreted MHC remain alive and therefore continue to produce MHC. Problems of purity are overcome because the only MHC molecule secreted from the cell is the one that has specifically been constructed to be secreted. Thus, transfection of vectors encoding such secreted MHC molecules into cells which may express endogenous, surface bound MHC provides a method of obtaining a highly concentrated form of the transfected MHC molecule as it is secreted from the cells. Greater purity can be assured by transfecting the secreted MHC molecule into MHC deficient cell lines.

Production of the MHC molecules in a hollow fiber bioreactor unit allows cells to be cultured at a density substantially greater than conventional liquid phase tissue culture permits. Dense culturing of cells secreting MHC molecules further amplifies the ability to continuously harvest the transfected MHC molecules. Dense bioreactor cultures of MHC secreting cell lines allow for high concentrations of individual MHC proteins to be obtained. Highly concentrated individual MHC proteins provide an advantage in that most downstream protein purification strategies perform better as the concentration of the protein to be purified increases. Thus, the culturing of MHC secreting cells in bioreactors allows for a continuous production of individual MHC proteins in a concentrated form.

Although class I and class II ligands were first examined by Edman sequencing, the primary characterization of individual ligands has been significantly improved upon through MS/MS applications. This is frequently performed via LC/MS using a microcapillary (<1 mm i.d) HPLC column directly interfaced with a triple quadrupole mass spectrometer (Hunt et al. 1992). The rationale behind using columns of small i.d. is that a lower solvent flow rate is permitted that ultimately increases the sensitivity of mass spectrometric detection (Tomer et al. 1994).

Unfortunately, since sample load capacity decreases proportionately to the column diameter, such columns and LC/MS methods can be technically

difficult or inconsistent for laboratories not routinely employing them to operate, more robust protocols for producing and studying class I-derived peptides have been desired. The methodology of the present invention, as described herein, solves and/or meets this need in the art by a methodology which increases the quantities of ligands extractable by producing recombinant soluble class I and class II molecules. The sample amounts subsequently available offset handling losses at the bench, which have been estimated at 50% (Veronese et al. 1996; Hogan et al. 1998; and Dr. Peter Parham, unpublished observations), and obviate the need for microcapillary LC/MS prior to MS/MS analysis. This is because significantly larger (20-fold) ligand samples are instead separated by standard offline RP-HPLC followed by NanoES-MS mapping and NanoES-MS/MS sequencing. The methodology of the present invention has proven consistent for comparatively examining peptides extracted from different class I molecules. As shown in FIG. 6, utilizing the production of soluble class I and class II molecules of the present invention, allows one of ordinary skill in the art to locate and characterize overlapping ligands among distinct allotypes (FIG. 6).

The initial HLA molecules selected for examination and production were from the HLA-B15 family, a hypothetical schematic of which is presented in FIG. 7. The HLA-B15 family represents a broad and diverse group of molecules comprised of nearly 50 evolutionarily related allotypes differing almost

sequentially by 1-15 peptide binding groove residues, and they are observed throughout numerous ethnic populations (Hildebrand et al. 1994); serological and DNA-based typing thus far confirm distribution of B15 alleles among Caucasians, Amerindians (North and South), Mexicans, Blacks (African and American), Indians, Iranians, Pakistanis, Chinese, Japanese, Koreans, and Thais. The majority of HLA B-locus polymorphisms known to exist are represented among the members of this allelic family. HLA-B*1501 appears to be the "ancestral allele."

The specific B15 allotypes initially selected for review and use with the present invention were B*1501 (Pohla et al. 1989; Choo et al. 1993; Hildebrand et al. 1994; Lin et al. 1996), B*1503 (Domena et al. 1993), B*1508 (Hildebrand et al. 1994), B*1510 (Domena et al. 1993; Rodriguez et al. 1993; Rodriguez et al. 1996), B*1512 (Hildebrand et al. 1994), and B*1518 (formerly B*7901, Choo et al. 1991; Lin et al. 1996; Rodriguez et al. 1996) (Table 1 and FIG. 8). B*1508 differs from B*1501 by a single mutagenic event in the α_1 helix, while B*1512 differs by a single mutagenic event in the α_2 helix; the remaining three alleles demonstrate a progressive series of polymorphisms throughout their binding grooves imposed by sequential mutagenic events during their divergent evolution from B*1501.

Using the primer sets of HLA5UT (Domena et al. 1993) and sHLA3TM (Prilliman et al. 1997) or 5PXI and 3PEI (Table 2) and template DNA from

reliable full-length cDNA clones of HLA-B15 molecules B*1501, B*1503, B*1508, B*1510, B*1512, and B*1518, truncating PCR was performed for each on a Robocycler (Stratagene) for 30 cycles as previously described (Prilliman et al. 1997). The resultant PCR products contained the leader peptide, α_1 , α_2 , and α_3 coding domains of the HLA heavy chain.

The PCR products were introduced into mammalian expression vectors. Initial constructs (truncated B*1501, B*1503, and B*1508) were prepared with the PSR α -neo vector (Lin et al. 1990), which has formerly been used to express non-truncated HLA molecules (Barber et al. 1997; Martinez-Naves et al. 1997), while other constructs (truncated B*1501, B*1503, B*1508, B*1510, B*1512, and B*1518) were additionally prepared with either pcDNA3 or pcDNA3.1(-) (Invitrogen). Constructs using the PSR α -neo vector were made from PCR products of the HLA5UT and sHLA3TM primers; the PCR products were subcloned into M13 (mp18 or mp19) according to standard protocols (Domena et al. 1993) so that confirmatory single-stranded DNA sequencing could be performed with Cy5-labelled versions of the primers M13 universal, 4N, and 3N (mp 18) or M13 universal, 3S, and JD3S (mp19), listed in Table 2 and described previously (Ennis et al. 1990; Domena et al. 1993), using the AutoLoad sequencing kit and ALFexpress automated sequencer (both Amersham Pharmacia Biotech). The insert was then prepared and purified, and this was followed by subcloning into PSR α -neo. Constructs using the pcDNA3 vector

were made from PCR products of the 5PXI and 3PEI primers; these PCR products were subcloned into M13 and sequenced as above, following which the insert was subcloned into pcDNA3. Constructs using the pcDNA3.1(-) vector were made from products of the 5PXI and 3PEI primers; PCR products were directly subcloned into pcDNA3.1(-), following which double-stranded DNA sequence analysis was performed with Cy5-labelled versions of the primers 3S, 4N, T7 promoter, and pcDNA3.1/BGH (Table 2).

DNA from each of the construct clones was prepared using Qiagen Midi kits for transfection of the class I-negative B-LCL 721.221. Cells growing in log phase in RPMI-1640 + 2 mM L-glutamine + phenol red + 20% FCS were pelleted and electroporation was performed as described (Gumperz et al. 1995) prior to beginning selection with 1.5 mg/mL G418. Upon establishment of confluent growth after approximately 3 weeks, putative transfectant wells were screened for sHLA production using a sandwich ELISA (Prilliman et al. 1997). Transfectant wells positive for sHLA production were then subcloned by limiting dilution to establish cell lines optimally secreting greater than 1 µg/mL of class I molecules in static culture over 48 h. Satisfactorily subcloned transfectants were then expanded, frozen in RPMI-1640 + 20% FCS + 10% DMSO, and stored at -135°C.

Since hollow-fiber bioreactors have been applied in place of *in vivo* hybridoma culture and MAb harvest from ascites (Evans et al. 1996) in order

to continuously produce large quantities of pure immunoglobulins, they were utilized to produce and harvest the sHLA of the present invention. The Unisyn Technologies CP-3000, the standard flow path and primary components of which are diagrammatically simplified in FIG. 9, was selected for hollow-fiber bioreactor culture of successfully established transfectants. In this system, basal media is pumped into the fully-assembled system from a 200 L barrel; the media flows from the 4 L reservoir tank into the hollow-fiber networks of the four bioreactors, which provide 2.7 m² of surface area per cartridge, and then exits as waste. ECS media and sHLA harvest are tandemly pumped into and out of the 270 mL cartridges, respectively, with each bioreactor receiving/yielding equal media/harvest volumes as regulated by in-line solenoids.

The CP-3000 was set up according to the manufacturer's protocol. After the system was completely prepared, at least 1×10^9 viable cells of a transfectant were grown in roller bottles of RPMI-1640 + 2 mM L-glutamine + phenol red + 10% FCS. The cells were pelleted and inoculated into the ECS of the bioreactor cartridges. ECS feed and harvest bottles were then attached to their corresponding lines, and the basal and recirculation rates were initially set to 100 and 1000 mL/h, respectively; the ECS was usually not activated until 24-36 h following inoculation. The system was then monitored at least twice daily over 4-6 weeks, with adjustments made as necessary. This involved checking

the glucose concentration and pH from manually-drawn reservoir samples, checking DO readings, and regulating the basal and ECS rates accordingly (Prilliman et al. 1997; Prilliman et al. 1998). A fresh harvest sample was periodically extracted directly from the system to quantitate sHLA production by ELISA for production level monitoring (FIG. 10).

Cells were cultured in the bioreactor system during each run until the desired amount of sHLA had been produced and collected in harvests. Each of the bioreactor runs chronologically depicted in FIG. 10 was aborted once approximately 150 mg of sHLA was determined by ELISA to be contained in the respective harvests collected (Prilliman et al. 1997). The majority of runs were performed using 721.221 cells transfected with pcDNA3 or pcDNA3.1 (-) vector constructs, considering: (i) the significant differences in time frame between the runs performed using cell lines expressing soluble B*1501 from either the PSR α -neo or pcDNA3 vectors (3 months versus 1 month); and (ii) the fact that peptides extracted during each run produced identical motif results.

Upon completing a bioreactor run, sHLA complexes were purified from the harvests obtained (Prilliman et al. 1997). A 100 mL matrix of either the β_2 m-specific MAb BBM.1 (Brodsky et al. 1979) or W6/32 (Barnstable et al. 1978) coupled to CNBr-activated Sepharose 4B (Amersham Pharmacia Biotech) according to the manufacturer's instructions was equilibrated with wash buffer (20 mM sodium phosphate, pH 7.2 + 0.02% sodium azide), and harvests were

applied to the column using a GradiFrac LC system (Amersham Pharmacia Biotech); the load capacities for 100 mL matrices of the MAbS BBM.1 and W6/32 were approximated at 10 and 40 mg sHLA respectively, as monitored for saturation ELISA of screening pre- and post-column samples. The column was then washed, eluted with 0.2 N acetic acid, and neutralized with wash buffer. Both BBM.1 and W6/32 were used to affinity purify B*1501, the first molecule prepared. However, due to the differences in purification efficiency noted above between the two MAbS, W6/32 alone was employed to isolate the remaining molecules from bioreactor harvests. This MAb has been frequently used by others in purifying HLA (Falk et al. 1991; Barber et al. 1997).

The fractions collected during affinity column elution demonstrating UV absorbance at 280 nm were pooled, and glacial acetic acid was added to 10% volume to extract the peptides as described (Barber et al. 1995). Bound peptides were separated from heavy chains, β_2m , and BSA by passage through 3 kDa exclusion membrane filters (Amicon) (Prilliman et al. 1997). The ligand-containing eluate was then lyophilized.

To remove residual salts and free amino acids remaining from the extraction process, isolated peptides were purified of free amino acids and salts prior to fractionation. This was done on a 2.1 x 100 mm C18 column (Vydac) with a steep RP-HPLC gradient using a DYNAMAX HPLC system (Rainin). The gradient was generated by increasing to 100% buffer B (0.06% TFA in 100%

acetonitrile) in 1 min, holding for 10 min, and returning to buffer A (0.1% TFA in HPLC-grade water) in 1 min. The column was loaded with peptides reconstituted in the minimum volume of buffer A required for solubilization. During the run, the region corresponding to absorbance at 214 nm was manually collected. Typically 1/100th of the total purified ligand collection volume was removed and subjected to Edman sequencing for 14 cycles on a 492A pulsed liquid-phase protein sequencer (Perkin-Elmer Applied Biosystems Division) according to established protocols (Falk et al. 1991; Barber et al. 1995).

The purified ligands were next fractionated by RP-HPLC. For preliminary diversity assessment, approximately 150 µg of peptides, as calculated from the ELISA-based total mass of SHLA bound to the affinity column and an estimated 50% handling loss, were loaded in 10% acetic acid onto a 1.0 x 150 mm C18 column (Michrom Bioresources, Inc.) and separated using an initial gradient of 2-10% buffer B (0.085% TFA in 95% acetonitrile) in 0.02 min followed by a linear gradient of 10-60% buffer B in 60 min at 40 µL/min on a 2.1 x 150 mm C18 column (Michrom Bioresources, Inc.); buffer A was 0.1% TFA in 2% acetonitrile. Absorbance was monitored at 214 nm, and fractions were automatically collected every minute. For comparative analyses, approximately 400 µg of peptides were injected in 10% acetic acid containing 2 µg of the dye methyl violet base B to control for gradient consistency between runs (FIG. 11).

The gradient formation parameters consisted of 2-10% buffer B in 0.02 min and 10-60% buffer B in 60 min at 180 μ L/min. Absorbance was monitored at 214 nm, and fractions were automatically collected every minute. Edman sequencing of fractions, when performed, was conducted on 1/20th of each.

To map extracted peptides and obtain primary sequences, a triple quadrupole mass spectrometer with an ES ion source, as generically depicted in FIG. 12, was employed. By using a triple quadrupole instrument, not only are all of the ions present within a given fraction be summarized for a designated mass range (mass mapping), but ions may then be selectively fragmented in order to obtain information from which sequence information can be derived (characterization). This is due to the flexibility afforded by the quadrupole mass analyzers: Q1 and Q3 act as mass filters which can be set to generate alternating DC and RF voltage fields for selectively transmitting specific ions (Watson 1997). However, q2 is an enclosed transmission-only quadrupole; it can be pressurized with inert gas for the collisional dissociation of an ion transferred through Q1. The specific ionization interface, NanoES, chosen here as an ES source functions on the principles described by developers Wilm and Mann (Wilm and Mann 1996). To establish and validate the procedure, comprehensive peptide mapping and sequencing were first performed among fractions 6 through 19 (FIG. 11), which represented a region of relatively rich ligand concentration (data not shown), for B*1501, B*1503,

B*1508, and B*1510; once this was accomplished, a more focused, and therefore less extensive, comparison was subsequently made between B*1501 and B*1512.

Prior to NanoES-MS, RP-HPLC fractions were completely dried by speed vac; the peptides were then resuspended in 0.1% acetic acid in 1:1 methanol:water. Aliquots from each of the individually concentrated fractions were loaded into 5 cm. gold/palladium alloy-coated borosilicate pulled glass NanoES sample capillaries (Protana A/S). To begin sample flow and data collection, the loaded capillary tube was next carefully opened as described (Wilm and Mann 1996). The capillary was then positioned directly in front of the API III⁺ (PE SCIEX) triple quadrupole mass spectrometer's orifice, and 20-30 scans were collected as separate data files for the mass range 325-1400 m/z while operating the instrument at positive polarity. This procedure was performed sequentially to obtain constituent mass data for samples drawn from each RP-HPLC fraction.

Spectral "ion maps" were generated from the TICs acquired for each fraction. The maps obtained from corresponding fractions of peptides eluted from different HLA-B15 molecules were aligned (FIG. 13A), and ions of interest for NanoES-MS/MS were located. The ion maps were typically compared following baseline subtraction and smoothing. Putative ligand matches or, in the case of B*1512, mismatches among the ions were identified through a

combination of data centroiding and direct visual assessment. Preference was placed upon selecting doubly-charged, or $[M+2H]^{2+}$, or higher ion forms commonly resulting from electrospray ionization for subsequent NanoES-MS/MS since the resulting daughter ion spectra were richer than those obtained from the collision of singly-charged, or $[M+H]^+$, ions (data not shown).

NanoES-MS/MS was performed by loading into a NanoES capillary tip, as described above, the desired volume of a fraction for which data was to be acquired. The volume loaded depended upon the relative sample flow rate achieved after opening the capillary tip and how long data acquisition was intended to proceed. Typically 3-4 μ L were loaded at a time to collect MS/MS data for 20-25 mid- to low-intensity ions from a given fraction. Once loaded, the source head was positioned and the capillary opened as before. Separate data files were collected for each ion subjected to collisional dissociation.

Daughter ion spectra were generated from the TICs obtained in this manner for each ion chosen. The specific approach taken to interpret individual MS/MS spectra varied from ion to ion depending upon the quality of the data sets obtained but adhered to the general rules of MS/MS fragment interpretation (Roepstorff and Fohlman 1984). The Predict Sequence algorithm included as part of the BioMultiView software (BioToolBox package, PE SCIEX) was employed for the *de novo* sequence interpretation, and the sequences deduced were checked for identity with source proteins in various databases

using the PeptideSearch algorithm (European Molecular Biology Laboratory; Mann and Wilm 1994) and performing advanced BLAST searches (Altschul et al. 1997) against the National Center for Biotechnology Information (National Institutes of Health) databases.

Leu and Ile were indistinguishable unless suggested by Edman data and/or specific sequence matches, as were Gln and Lys since lysyl derivatization prior to fragmentation was not performed. NanoES-MS/MS data from ions of potentially overlapping peptides was aligned to confirm or refute the presence of shared ligands among different HLA-B15 molecules, as shown for one ion confirmed as an overlapping peptide across B*1501, B*1503, and B*1508 in FIG. 13, (B and C). Reproducibility of the protocol in its entirety is demonstrated in FIG. 14. To establish a numerical description ($N_{\text{sum}}/C_{\text{sum}}$) comparing ligand N- and C-regional occupancies for each allotype, N and C values for the four ligand positions at either terminus were determined by summing occurrence frequencies (using an arbitrarily-defined baseline of 10%); N_{sum} was subsequently calculated from the four N values, and C_{sum} was calculated from the four C values.

Peptides from HLA-B15 molecules were subjected to pooled Edman sequencing as well as more extensive examinations, including fractional Edman sequencing and mass spectrometric characterization of individual ligands. This was done to: (i) confirm the production/purification methods employed; and

(ii) evaluate the relative nature and complexity of the peptides contained in extracts of naturally presented ligands.

Upon extracting peptides from each of six different B15 molecules, pooled Edman sequencing was performed. This was done both to validate results from the extraction of sHLA ligands with the techniques previously employed by others (B*1501 and B*1508, Falk et al. 1995; Barber et al. 1997), and to obtain novel motifs from the molecules that had not been previously examined (B*1503, B*1510, B*1512, and B*1518) for providing "traditional" points of reference.

Overall, the B*1501 motif (FIG. 15) was in agreement with the dominant P2 and P9 anchors (Gln and Tyr/Phe, respectively) previously defined (Falk et al. 1995; Barber et al. 1996; Barber et al. 1997). This result demonstrates that the peptides extracted from sHLA-B*1501 complexes were identical to those extracted by others from natural membrane-bound molecules. However, differences in the whole pool sequencing data arising from sHLA purified by BBM.1 (FIG. 15B) versus W6/32 (FIG. 15A) indicated that a greater number of peptides than originally realized should actually contribute to the consensus B*1501 motif.

First, while Gln and other residues including Leu, Met, and Val have been previously reported at P2 using W6/32 to isolate complexes, the aliphatic side chain Pro was strongly detected at P2 in the BBM.1-purified B*1501 motif as

well. Though not employed by other groups pursuing similar studies, the β_2m -specific MAb BBM.1 was initially chosen here to avoid biases potentially imposed upon the class I heavy chain by bound peptides (Bluestone et al. 1992; Catipovic et al. 1992; Solheim et al. 1993). Of interest was that Pro has not been reported before as a strong or even weak P2 anchor in the B*1501 peptide motif. It has been suggested that B-locus allotypes that present peptides with Pro at P2 demonstrate a shallower B-pocket within their binding grooves than does B*1501, which exhibits a Ser at α -chain position 67 rather than a more constricting residue such as Phe (Barber et al. 1997).

This data suggests the possibility that Pro binds amicably within this deeper pocket but perhaps induces an altered heavy chain conformation that negatively biases purification of complexes by the W6/32 MAb typically used. The observation of a P2 Pro occupying a pocket with suboptimal physical complementarity is corroborated by a similar occurrence among peptides bound by the murine class I molecule L^d (Corr et al. 1992; Balendiran et al. 1997). Purification methodology serves, therefore, as a factor in allele-specific motif predictions, and the whole pool sequencing with peptides extracted from both BBM.1 and W6/32-purified B*1501 complexes demonstrated that a strong Pro anchor at P2 is antibody dependent.

The pooled Edman motifs obtained for W6/32-purified molecules divergent from B*1501 are shown in FIG. 16. Like B*1501, each of the motifs

reflected a nonameric consensus with distinct P2 and P9 anchors and internal auxiliary anchor preferences. The B*1508 motif, described by another group while its preparation was in progress during the development of this invention (Barber et al. 1997), was consistent between the two laboratories; it demonstrated a preference for the small side chains Pro and Ala at P2 and aromatic residues Tyr and Phe at P9. The B*1512 motif appeared nearly identical to that obtained from B*1501; by extension, considering that B*1519 differs from B*1512 in α_3 , which does not contribute to the peptide binding groove, it is predicted that B*1519 would bear the same motif as B*1501 and B*1512.

B*1503 diverges somewhat from the other three molecules presented above in showing a distinct preference for ligands with a neutral, polar Gln or positively-charged Lys as the P2 anchor; the aliphatic Met was evident here as well, though to a lesser degree than noted for the hydrophilic Gln and Lys residues (Prilliman et al. 1999). Like B*1501, B*1508, and B*1512 however, aromatic residues Tyr and Phe defined a hydrophobic P9 anchor. The only other class I molecules with motifs whose definitions thus far indicate a Lys at P2 are B*3902 (Falk et al. 1995) and B*4801 (Martinez-Naves et al. 1997), both of which structurally bear B-pockets identical to B*1503 except for a single L→T or L→E substitution, respectively, at the α_2 helical residue 163 (Chelvanayagam 1996). The B-pocket of B*1503 is indistinguishable from that of B*4802

(Chelvanayagam 1996), whose motif remains undetermined but is likely to follow suit with those of B*1503 and these other molecules at the second ligand position. An assortment of polar, charged, and hydrophobic residues is evident at P3 of the B*1503 motif.

The B*1510 motif demonstrated a strict preference for ligands bearing a basic, hydrophilic His as a P2 anchor. A hydrophobic P9 anchor was described by residues including Leu and Phe. The B*1510 motif strongly resembled that previously defined for B*1509, which exhibits nearly identical anchor preferences with His at P2 and Leu, Phe, and Met at P9 (Barber et al. 1997). B*1510 and B*1509 differ structurally only by a substitution of N→D in α_2 at the α -sheet floor position 114, which takes part in forming several specificity pockets within the peptide binding groove (FIG. 3).

By extrapolation from its structural neighbors, it was assumed that B*1518 would have for its motif a P2 anchor of His (as seen for B*1510 and B*1509) and a P9 of Tyr and Phe (as seen with B*1501, B*1503, B*1508, and B*1512). B*1518 differs from B*1510 solely at position 116; two other HLA-B molecules that differ exclusively at this position are B*3501 and B*3503: they differ by a S→F substitution here, which would sterically mimic the substitution between B*1518 and B*1510 and confer B*1510-like P9 preferences (Steinle et al. 1995; Kubo et al. 1998). Based upon this, and the fact that the P9 environments of B*1518/B*3501 and B*1510/B*3503 are similar

(Chelvanayagam 1996), it was first predicted, and then confirmed following pooled sequencing, that B*1518 would bear the "hybrid motif" described.

Pooled Edman sequencing data therefore demonstrates that (i) the peptides extracted from sHLA complexes produced according to the methodology of the present invention are consistent with those previously extracted from native, cell surface-expressed complexes, and (ii) nonameric ligand lengths with anchor residues at P2 and P9 characteristic to specific polymorphisms are preferred. As for functional implications, the major anchors would predict natural ligand overlaps with B*1501 by B*1503 and B*1512.

Since class I peptide pools consist of thousands of different ligands, an early investigation during development of the analysis strategy and methods disclosed herein was to next fractionate and then Edman sequence the peptides extracted from one of the molecules. BBM.1-purified B*1501 (Prilliman, et al. 1997), the first soluble molecule produced by a non-repeatable precursor methodology to the fully repeatable and characterized methodology of the present invention, was initially examined to explore the general diversity around a pooled motif (Prilliman et al. 1998).

Edman sequencing of peptide-containing fractions collected from the RP-HPLC gradient shown and described in FIG. 17 supported the existence of peptides up to 12 residues long and revealed significant positional diversity among the peptides isolated from B*1501. This positional diversity was

illustrated in 16 representative fractions (FIG. 18). Assessment of dominant, strong, or weak amino acid residues present at the various positions indicated that the dominant anchors previously defined by whole pool sequencing did not necessarily predominate in fractions of the peptide pool (FIG. 19). In fractions 15 and 31 the dominant P2 Gln was replaced by a dominant Ala and a dominant Lys respectively, while the pooled motif Tyr fell below residues such as His and Lys at P9 in the same fractions. While variations on the consensus motif were prevalent at P2, this was not solely restricted to the N termini of bound peptides as shown in FIG. 18.

Among the 16 fractions studied, 12 demonstrated weak sequence yields out to 12 cycles of degradation; in nine of these fractions, P12 was occupied by the charged or polar residues Glu, Arg, Lys, Ser, His, or Gln. Additionally, the presence of numerous decamers within the B*1501 peptide population was suggested in that 11 of the fractions sequenced exhibited a typical P9 residue, Tyr or Phe, at P10. For example, in FIG. 19 the strong Phe presence at P10 in fraction 28 suggests that P10 serves as an anchor for a decamer(s) present within the fraction. The shifting of a P9 anchor preference to P10 is consistent with the P10 occupancies of individual decamers formerly characterized from B*1501 peptide pools (Falk et al. 1995; Barber et al. 1997). Although residues representing P9 anchors were seen at P10 and P12, amino acids unique in such longer ligands were also detected, suggesting that the shifting of a previously-

reported P9 anchor is not the only means by which longer B*1501 peptides are bound within the peptide binding groove (Collins et al. 1994).

During the course of examining ligands from other sHLA molecules, which were purified alternatively with MAb W6/32, aliquots of RP-HPLC fractions from some were also subjected to Edman sequencing on occasion. The results from these random samplings of B*1501, B*1508, B*1503, and B*1510 fractions are summarized for major characteristics in Table 3. The characteristics recorded included preferences other than those seen in the pooled motifs at traditional anchor positions P2 and P9 and whether signal levels indicated the presence or not of residues beyond the nine cycles of degradation typically observed. Of additional interest, no Lys was observed among Edman sequenced fractions as an additional preference at P2 for B*1501 purified with W6/32, a finding contrary to the P2 preferences among fractions of BBM.1-purified material which lends further support to the argument for MAb bias, as discussed earlier, existing in complex purification.

In summary, the fractionation of B*1501 peptides prior to Edman analysis resulted in amino acid sequence data demonstrating that the components of a peptide pool can vary considerably from the overall motif (Prilliman et al. 1998). These data, as well as results obtained from other B15 molecules, suggested that peptides bound by the various molecules include: (i) species which are either longer or shorter than the nonameric size typically indicated

by pool sequencing alone; and (ii) species that exhibit primary sequences different from those predicted by pooled sequencing. The additional data provided by further exploration of fractionated extracts from several of the B15 molecules in this manner expands the molecules with the potential for presenting B*1501-overlapping ligands to include not only B*1503 and B*1512 but B*1508 and B*1510 as well. Such diversity among fractions indicated that characterization of individual ligands would provide information not available in motifs.

Individual peptides from B*1501, B*1503, B*1508, B*1510, and B*1512 were comparatively examined to investigate whether the added flexibility observed through Edman degradation of RP-HPLC fractions would allow for natural ligand overlaps to occur across their respective polymorphisms.

More than 400 individual ligands extracted from the five distinct HLA-B15 allotypes were characterized according to the methodology of the present invention. The ligands characterized here were from ion map masses found in multiple B15 allotypes as demonstrated in FIG. 13A. Selected ions were then dissociated by NanoES-MS/MS, and the resulting fragment information was compared and interpreted, as described hereinabove, to determine if the ions represented sequence-identical or merely mass-identical ligand matches.

Individual peptide ligands characterized from the five B15 allotypes are listed in Tables A-E. The number of ligands for which either complete or partial

sequences were obtained here was as follows: B*1501 = 126, B*1503 = 74, B*1508 = 96, B*1510 = 123, and B*1512 = 30. While the pooled motifs of peptides extracted respectively from the five molecules described nonamers with various P2 and P9 dominant anchors and P3 auxiliary anchor preferences (FIGS. 15 and 16), the single peptide sequences ranged from 7 to 12 amino acids in length and demonstrated (i) greater heterogeneity at their N-terminal/proximal regions than their C termini, and (ii) varying degrees of observed ligand overlap, both of which will be examined in the subsequent sections of this chapter.

In terms of length heterogeneity, the endogenous peptides eluted from B*1501, B*1503, B*1508, B*1510, and B*1512 varied in length from 7 to 12 amino acids as shown (FIG. 20). An overall length breakdown of the peptides listed in Tables A-E demonstrates that approximately 6% are heptamers, 21% are octamers, 50% are nonamers, 19% are decamers, 3% are undecamers, and 1% are dodecamers. Further emerging from the length characterization of individual ligands is the observation that peptides bound by each of the B15 molecules spanned ranges of 5 to 6 amino acids in length. For example, peptides eluted from B*1501, B*1510, and B*1512 were 7-11 amino acids in length, while those from B*1503 and B*1508 were 7-12 amino acids in length.

Coupling this length variability with the likewise varying degrees of regional sequence heterogeneity (which will be discussed) leaves only 23% of

the endogenously loaded peptides characterized in Tables A-E as "ideal nonamers" with both P2 and P9 anchors in concordance with the dominant or strong preferences of the pooled Edman motifs from their respective source molecules. This finding is of principal significance in that a majority (77%) of potential ligands for any of these HLA-B15 molecules would therefore be overlooked if the length and sequence constraints of their pooled motifs were utilized as the primary criteria in searching for potential epitopes specific to them.

Examples of ligands from this study with homology to stretches of known proteins are shown in Table 4. The peptides yielding 100% identical BLAST database hits were grouped into seven categories, which were defined here according to the common natures of their potential source proteins: HLA ligands, replication/transcription/translation ligands, biosynthetic/degradative modification ligands, signalling/modulatory ligands, transporter/chaperone ligands, structural/cytokinesis ligands, and unknown function ligands. Aside from the HLA heavy chain-derived ligands, most appear to be derived from cytoplasmic or nuclear proteins, which illustrates that the typical endogenous pathway is involved in generating the majority of the class I-loaded peptides characterized (York and Rock 1996).

Of the 44 peptide sequences listed in Table 4, it is noteworthy that overlaps across other HLA-B15 molecules are evident within our data collection.

The B*1510 tapasin₃₅₄₋₃₆₂ ligand HHSDGSVSL, as well as both THTQPGVQL from septin 2 homolog₇₀₋₇₈ and SHANSAVVL from β -adaptin₂₄₉₋₂₅₇, have also been sequenced from B*1509 extracts (Barber et al. 1997), and the B*1501/B*1508/B*1512 ubiquitin-protein ligase₈₃₋₉₁-derived ligand ILGPPGSVY was characterized from endogenously bound B*1502 peptides (Barber et al. 1997). The eIF3-p66₆₁₋₆₉ nonamer SQFGGGSQY (Falk et al. 1995; Barber et al. 1996) was found here within B*1501, B*1503, B*1508, and B*1512 extracts. The decamer YMIDPSGVSY, which is homologous to proteasome subunit C8₁₅₀₋₁₅₉, was also previously described as a ligand for B*1502 (Barber et al. 1997), B*1508 (Barber et al. 1997), and B*4601 (Barber et al. 1996); it was found here presented by B*1501, B*1508, and B*1512. Some of the specific overlapping ligands identified in this study therefore overlap in antigen presentation with the HLA-B15 allotypes characterized by others.

Given the length heterogeneity observed among the ligands collectively characterized from B*1501, B*1503, B*1508, B*1510, and B*1512, analysis of peptide ligand primary structures proceeded through two separate alignments (N- and C-terminal) for each HLA-B15 allotype. The frequencies with which specific side chains occurred at (i) the N terminus and first three residues internal from it, and (ii) the C terminus and the first three residues internal to it were tabulated. As graphically summarized throughout parts A and B, respectively, of FIGS. 21-25, the salient features of these ligand regions

and how they correlated with the structures of the molecules from which they were extracted can be examined.

The N-terminal/proximal regions for ligands from each of B*1501 (FIG. 21A), B*1503 (FIG. 22A), B*1508 (FIG. 23A), B*1510 (FIG. 24A), and B*1512 (FIG. 25A) clearly demonstrated acceptance of a variety of amino acid side chains, particularly at P3 and P4, by the portions of the binding groove assumed to interact with ligands at the designated positions. With the exception of B*1512 ligands, which were obtained from both a smaller and more biased collection of ions (see FIG. 25 legend), higher points in each of the graphs occur for certain side chains indicated along the P1 and, to a greater extent, P2 data lines, which represent the first and second positions, respectively, of the characterized ligands.

The results for P1 obtained from the HLA-B15 ligands characterized are of interest since the analysis of Edman sequencing data depends upon comparing relative increases between cycles and is therefore unreliable for making side chain determinations at this first position, especially when complex mixtures of peptides are examined (Stevanovic and Jung 1993). All five B15 allotypes demonstrated a number of side chains at P1, with preferences for residues including Ala, Leu/Ile, Gly, Ser, Thr, and Tyr observed in varying degrees among them (FIG.s 21-25, A). Overall, P1 appeared to be occupied in a majority of ligands by aliphatic amino acids.

Though the pooled motifs of B*1501 and B*1512 (FIGS. 15 and 16) as well as the spectral ion maps obtained from their RP-HPLC fractions (data not shown) were virtually identical, the substitution difference between the two molecules at A-pocket residue 167 suggested that ligands bound by these molecules might differ at P1. Performing NanoES-MS/MS upon a handful of ions, which appeared to be exclusive to B*1512, confirmed the presence of several ligands presented by B*1512 but not B*1501 (Table 5). Of the 16 sequenced, seven indicated His, two indicated Arg, and one indicated Lys at P1. In comparison, only a single B*1501 peptide each presented with His, Arg, or Lys at P1 out of 126 ligands characterized (Table A). An explanation for the existence of this subset of B*1512-restricted ligands with positively-charged N-termini could lie in the W→G substitution observed between B*1501 and B*1512 at α_2 position 167, which might sterically enhance the influence by the adjacent acidic residue at position 166 (Glu in B*1501 and Asp in B*1512) of B*1512 upon P1 in the A-pocket. Comparing individual ligands between B*1512 and B*1501 supports the notion that the polymorphism segregating them will confer distinct yet subtle effects upon peptide binding by other allotypes differing in this manner.

P2, which has been considered to act as a primary anchor for peptide ligands among most class I molecules described to date as based upon pooled Edman motifs, is classically accepted to associate with the B-pocket of the

peptide binding groove. In terms of the motifs derived here by pooled sequencing (FIGS. 15 and 16) and the motifs previously established for other B15 family allotypes (Falk et al. 1995; Barber et al. 1996; Barber et al. 1997), a Gln at P2 is common to B*1501, B*1502, B*1503, B*1512, and B*1513 (Table 6). Three alleles, B*1502, B*1513, and B*1508, have a Pro at P2, while the lack of a strong Pro at P2 in both B*1501 and B*1503 corresponds to polymorphism at heavy chain positions 63 and 67. For example, B*1501 appears to lose the propensity for Pro at P2 due to polymorphism at position 63, while B*1508 appears to lose a Gln at P2 resulting from polymorphism at 67. Thus, comparisons within the B15 family highlight how substitutions at positions 63 and 67 of the class I heavy chain α_1 helix appear to confer differential interaction with P2 of the peptide ligand.

While residue 63 modulates the size/conformation of P2, it can be seen that residues 24 and 45 influence the P2 charge nature propensities. A comparison of B*1501 and B*1503 illustrates how polymorphisms at positions 24 and 45 of the B-pocket influence P2 preferences in this manner; B*1503 is one of four B15 alleles with known motifs bearing a positively charged P2. Allotypes B*1509, B*1510, and B*1518 recognize a positively charged His at P2 and have the same residues at 24 and 45 as B*1503, but the differences at positions 63 and 67, which separate B*1503 from the other three molecules, again modulate the contour of P2 such that different positively charged P2

residues fit respectively into the B*1503 and B*1509/B*1510/B*1518 B-pocket categories.

It has previously been proposed that polymorphisms in the α_1 helix prompt major changes in the repertoires of peptides bound by allotypes differing in this region (Barber et al. 1997). That any region, helical or sheet, of α_1 would influence peptide P2 preferences more than α_2 is of little surprise though since 14 of the 18 residues forming the B-pocket belong to the α_1 domain (Table 6). A comparison of 12 known B15 motifs in the B-pocket suggests more refined rules for α_1 in general, whereby polymorphisms in the helices sculpt the conformation and size of the amino acids that can fit into the peptide binding groove's B-pocket. Further analysis of the B15 motifs at P2 suggests that polymorphisms lining the floor of the groove tend to regulate the hydrophobic and/or charged nature of the residues at P2 of bound ligands. Perhaps in this way the walls and floor of the binding groove work in concert: the α -helical residues sterically control which amino acids can fit, while the β -sheet residues act to attract or repel particular side chains based on chemical compatibility within the ligand binding groove.

The interactions described for P2 here are, however, more relaxed than previously thought. For a majority of the allotypes shown in Table 6, three or more different side chains are observed by pooled Edman sequencing as dominant or strong B-pocket residents and/or the B-pockets of the molecules

demonstrate abilities to naturally accommodate alternative P2 residues (Table 3 and FIGS. 21-25, A). Of specific interest, although Met appears in the B*1503 pooled motif as a strong P2 anchor residue (FIG. 16), only two of the peptides characterized for this allotype bear Met at P2 (FIG. 22A), while other residues including Gly, Pro, Ala, and Asn occur more often than Met at P2 among B*1503 ligands. The fact that Met appears in the pooled motif but fails to demonstrate a strong presence among the individual peptides indicates that disparate concentrations of ligands within extracts may skew pooled Edman sequencing results so as to be misleading.

Pro and Ala likewise appear with frequencies comparable to or exceeding those of the W6/32-purified pooled motif residues for B*1501 (FIG. 21A), and B*1508 ligands illustrate P2 inclinations for a rich array of side chains in addition to the motif-prescribed residues Pro and Ala which include Gly, Val, Met, Leu/Ile, Ser, Thr, and Gln/Lys. Similar variety is observed within the limited B*1512 ligand data set (FIG. 25A). In contrast, the B-pocket composition for B*1510 indicates His as the sole dominant/strong P2 occupant (Table 6), and among individual ligands characterized from B*1510 His is noted at a markedly higher degree than are alternative residues (FIG. 24A). However, amino acids including not only the positively charged Arg but to a greater extent Gly, Ala, Val, Leu/Ile, and Gln/Lys occur at P2 of some peptides are also characterized at P2 from this allotype. Thus the majority of HLA-B15

molecules demonstrate elastic N-proximal occupancies.

In comparison with the findings at the N-terminal/proximal regions, the C termini of ligands from each of B*1501 (FIG. 21B), B*1503 (FIG. 22B), B*1508 (FIG. 23B), B*1510 (FIG. 24B), and B*1512 (FIG. 25B) demonstrated a stricter acceptance of amino acid side chains. C-proximal ligand residues also revealed the existence of more distinct side chain tendencies.

For allotypes B*1501, B*1503, B*1508, and B*1512 a dominant C terminus was especially prominent among the ligands characterized from them, while B*1510 exhibited a P2 anchor nearly as strong as its primary C-terminal residue preference (FIG. 24, A and B). The aromatic residues Phe and, even more prominently, Tyr occupied the C-terminal positions of most peptides bound by the first four B15 molecules, which appeared to agree with P9 of their respective motifs (FIGS. 15 and 16). The bulk of B*1510 ligands demonstrated Leu/Ile at their C termini; other occupants at this position included Phe and Val, an interesting observation in that more B*1510 peptides presented with Val, which is not included in either the pooled (FIG. 16) or fractional (Table 3) Edman motifs examined from this allotype, than Phe, which is identified as a strong P9 occupant by pooled sequencing. Such is the likely result of disparate peptide concentrations affecting the pooled Edman sequencing results as mentioned previously. Another factor includes the diminishing picomole yields per successive cycle of Edman degradation; this leads to progressively higher

background signals and thus negatively affects sensitivity in examining the C-terminal/proximal regions of peptides (Stevanovic and Jung 1993).

The overwhelming conservation at the C termini of individual ligands (FIGS. 21-25, B) indicates that the C terminus acts as a dominant anchor for peptide ligands. P9 of pooled Edman motifs has classically been accepted to associate with the F-pocket of the peptide binding groove. C-terminal anchoring is observed here regardless of length heterogeneity (FIG. 20 and Tables A-E). For the majority (91%) of peptides greater than 9 residues long, this observation agrees with evidence that longer peptide ligands bulge centrally outward from the peptide binding groove. Sequencing individual ligands supports a concept that the C terminus of a ligand plays a dominant role as an anchor within the class I binding groove for the HLA-B15 allotypes examined.

With regard to pooled sequence motifs for HLA-B15 allotypes, all 12 molecules demonstrate chemical homogeneity at P9, with dominant/strong occupancies by hydrophobic residues (Table 7). The eight different F-pockets structurally represented among these allotypes show preferences for Tyr, Phe, Met, Leu, and Trp according to three functional group categories. This is in marked contrast with the B-pockets, for which eight different B-pockets among the same allotypes comprised seven distinct functional groups encompassing a mixture of both hydrophobic and hydrophilic side chains (Table 6).

It is interesting that, of these HLA-B15 molecules, nine have F-pocket functionality in the same category (B*1501, B*1502, B*1503, B*1508, B*1512, B*1516, B*1517, B*1518, and B*4601), with preferences for Tyr, Phe, and/or Met, despite the fact that they exhibit amino acid substitutions at nine different positions throughout the α_1 helix and β_2 sheets. This redundancy demonstrates that, contrary to what was seen among structural residues affecting the B-pocket, the α_1 helical polymorphism(s) shown for allotypes in the first category of Table 7 do not necessarily play a defined role in sculpting either the conformation or size preferences of ligands in this region of the peptide binding groove.

While the Edman-derived motifs for the B15 allotypes shown in FIGS. 15 and 16 clearly indicated P2 and P9 primary anchors and suggest an assortment of preferences at both P3 and P4, they fail to sufficiently capture the trends for auxiliary anchoring at the C-proximal regions of endogenously bound ligands which were perceptible throughout the individual peptide sequences. Additional preferences that likely serve as auxiliary anchors were evident at the C-proximal positions C⁻¹, C⁻², and C⁻³ in the cases of nonamers, octamers, and heptamers, as well as in ligands longer than nonamers. A review of the positional frequencies observed among the HLA-B15 peptides shows that amino acids such as Val, Leu/Ile, Ser, Thr, and Gln/Lys tended to predominate at nearly all three of the C-proximal positions of ligands presented by B*1501,

B*1503, B*1508, B*1510, and B*1512 (FIGS. 21-25, B). Of these residues, the hydrophilic, hydroxyl-containing Ser and Thr were especially frequent among these positions; nearly half (40%) of the ligands listed in Tables A-E, bear Ser and/or Thr at the designated C-proximal positions. In general, Glu occupied C⁻¹ and Gly occupied C⁻³ to some extent among all five allotypes.

Val (C⁻¹) and Pro (C⁻²) were especially prominent C-proximal residues observed among the B*1510 ligands; the overriding presence of Pro, which distinguished this region of B*1510-derived peptides from those of the other allotypes, can likely be attributed to steric influences imposed by the Tyr at α_2 position 116 in B*1510, which additionally interacts with the C- and E-pockets of the peptide binding groove (FIG. 3). Further distinguishing several B*1510 ligands from but rare occurrences among B*1501, B*1503, B*1508, and B*1512, Pro frequently appeared as well in various C-proximal sequence combinations with Ala or Val (Table 8).

The amino acid residues characterized from each of the five HLA-B15 allotypes with occupancy rates of at least 10% for the first four (N-terminal/proximal) and last four (C-terminal/proximal) positions among ligands, respectively, are condensed in Tables 9-13. Presenting the data already discussed in this manner effectively emphasizes C-terminal dominance and N-proximal flexibility. By comparison, the data illustrates the limitations of pooled Edman motifs in being able to adequately reflect a consensus of the individual

peptides contained within a given ligand extract. The $N_{\text{sum}}/C_{\text{sum}}$ quotients obtained as described for Tables 9-13 were less than 1.00 in the cases of all allotypes, thus providing a more fixed description to the C-terminal/proximal region (gray) as a whole with respect to the N-terminal/proximal region (black).

Among the N-regional position ligand residues occurring at >10%, nothing appears to prominently stand out at P3 and P4 although assignments were made to these positions via Edman sequencing. The occupancies that were observed were not necessarily captured by the motifs; a specific illustration of this is Ala (19.51%) at P3 of B*1510 (Table 12 and FIG. 16). This trend appeared likewise applicable at the P2 anchor, where with the exceptions of B*1508 and B*1510, occupants at this position among >10% of ligands from each of the remaining allotypes included additional side chains (for example, Ala at P2 in both B*1501 and B*1512) not accounted for by pooled sequencing. The C termini of each allotype are comprised of two amino acid specificities as shown by more than 80% of characterized peptides in all cases (Tables 9-13). In summary, comparing observed N- and C-regional occupancies among the characterized ligands underscores the flexibility of N-proximal versus the dominance of C-terminal preferences among the B*1501, B*1503, B*1508, B*1510, and B*1512 binding grooves.

A total of 40 specific ligands among the 449 characterized here (Tables A-E) overlapped across the peptide binding grooves of B*1501, B*1503,

B*1508, and/or B*1512 (Table 14); as previously discussed from the information in Table 4, some of the overlapping ligands likewise coincided with ligands characterized by others from additional HLA-B15 allotypes including B*1502 and B*4601. Length variations among the overlapping ligands identified tended to mimic those observed among the entire set of ligands characterized, formerly discussed and illustrated in FIG. 18. Only seven overlapping ligands were longer than 9 amino acids in length (4 decamers and 3 undecamers), while 16 fell short of 9 residues long (6 heptamers and 10 octamers); less than 50% of the successful overlaps were therefore nonameric.

Throughout the mapping and sequencing approach that was developed and executed as outlined earlier (FIG. 6), an extensive comparison was first conducted upon ions occupying RP-HPLC fractions 6-19 from separations as shown in FIG. 11 of ~400 µg of peptides from each of B*1501, B*1503, B*1508, and B*1510, the first four B15 molecules prepared in the course of this study. From this, 21 peptide overlaps across B*1508 and B*1501 were defined. Similarly, eight ligands overlapping B*1501 and B*1503 were identified, and four ligands were found to overlap across B*1508, B*1503, and B*1501. A conservative estimate, based upon past examination of B*1501 ligands, is that the ion maps for each of the B15 allotypes represented at least 2,000 individual peptides per molecule (Prilliman et al. 1997), yet B*1510 was not observed to share ligand overlaps with B*1508, B*1501, or B*1503 (Table

14).

The sequence data indicates that overlapping ligands bind across divergent B*1508, B*1501, and B*1503 binding grooves but not B*1510. This pattern likewise accentuates an apparently dominant role for C-terminal anchors in natural peptide binding as discussed previously. FIG. 26 depicts, in the context of the class I peptide binding cleft, the locations of polymorphisms that individuate B*1508, B*1501, B*1503, and B*1510 and highlights the anchoring residues for the peptide overlaps according to the N-proximal and C-terminal specificities of their respective presenting molecule's motif (FIGS. 15 and 16). Bolding the amino acids of these overlapping ligands, which are in agreement with the traditional pooled motifs, underscores the trend whereby a C-terminal anchor sequence is conserved within overlaps while the N-proximal anchor is considerably more flexible in its location and/or sequence. A lack of overlaps with B*1510 could potentially be explained by the S→Y substitution observed between this allotype and the other three at α_2 position 116. Thus, the conserved C-terminal anchors that facilitate the occurrence of B*1508, B*1501, and B*1503 overlaps fail to preferentially interact with the B*1510 C-terminal specificity pockets.

A further example provided here of how C-proximal auxiliary anchors might positively impact endogenous ligand binding is that eight of the peptides overlapping both the B*1508 and B*1501 antigen binding grooves bear Thr at

C¹, C², or C³, and in four cases the peptides that bind B*1508/B*1501 or B*1508/B*1501/B*1503 are heptamers with Thr occupying P7, their C-terminal positions (FIG. 26). The prominent role of Thr as a C-terminal/proximal auxiliary anchor is dramatically illustrated by the B*1508/B*1501/B*1503 overlapping heptamer CPLSCFT, where Thr provides a C-terminal anchor for this ligand not evident in the pooled motifs of the three allotypes.

Distilling the data from the overlapping ligands among B*1501, B*1503, and B*1508 suggests a model for endogenous ligand binding whereby peptides are first anchored or held in the class I binding groove by their C termini. In order for a given peptide to remain stably fastened in the groove for successful trimer assembly and subsequent export from the cell, it is observed that following rigid anchoring at the C terminus as described, a ligand must be subsequently tethered into the class I antigen binding cleft at a more variably defined N-proximal position. Such is the case for peptide ligand NQZHGSAEY, a nonamer that overlaps across B*1508, B*1501, and B*1503 (FIG. 24). According to this model (FIG. 27), a C-terminal Tyr securely anchors NQZHGSAEY into all three B15 allotypes, while a Gln at P2 anchors the peptide into B*1501 and B*1503 and a Gln/Lys (most likely a Lys based upon both motif assignments and fractional Edman sequencing data) at P3 provides additional anchoring for B*1501 and serves as the sole N-proximal anchor for B*1508. This model appears clearly applicable to at least 75% of the ligands

presented in FIG. 26; for those peptides to which it does not evidently apply, the possible anchoring modes remain open to further speculation at the level of individual ligands.

For example, the B*1501/B*1503 overlap AQFASGAGZ (FIG. 26) may instead be additively stabilized through the N-proximal anchors indicated at P2 and P3 as well as at the N-terminal position, since Ala demonstrated significant P1 occupancy among both B*1501 and B*1503 ligands, as previously shown (FIGS. 21 and 22, A). Additionally, the four heptameric overlaps that were observed across B*1508/B*1501/B*1503, which terminate in Thr, could lie within the peptide binding groove such that they are anchored N-terminally/proximally and their C termini interact with the C-proximal regions of the groove, which have demonstrated preferences for Thr; these ligands might therefore fail to extend into the F-pocket. As compared with C-terminal sequences, both length and N-proximal specificity characteristics of ligands generally play secondary roles in the natural binding of B15 peptide epitopes.

Further stemming from the data obtained by comparatively examining B*1501, B*1503, B*1508, and B*1510 ligands, differential interactions with the chaperone tapasin specifically influence the loading of peptides into HLA-B15 molecules. Tapasin, an MHC-encoded chaperone discussed hereinabove (Herberg et al. 1998), is a recently-discovered 48 kDa transmembrane glycoprotein resident to the ER that directly interacts with both calreticulin and

empty α -chain/ β_2m dimers to form a "loading complex" linked to TAP1/TAP2 (Sadasivan et al. 1996; Grandea III et al. 1997; Li et al. 1999). Tapasin is not a requirement for ligand loading via the typical endogenous processing pathway (Lewis et al. 1998; Peh et al. 1998), and aside from its proposed role in serving as a bridge between a class I dimer and the peptide transporter until release of mature trimers upon peptide binding, the exact role of tapasin during class I assembly is unknown (Pamer and Cresswell 1998). Interactions between nascent class I molecules and TAP1/TAP2 have, however, been shown to be influenced either directly or indirectly by α_3 and positions 116 and 156 of α_2 (Suh et al. 1999; Kulig et al. 1998; Neisig et al. 1996).

Of specific interest, past analysis of divergent HLA-B35 molecules has indicated that allotypes bearing an aromatic amino acid (Phe or Tyr) at position 116 interacted with TAP1/TAP2; allotypes bearing a Ser substitution at this site failed to demonstrate the interaction (Neisig et al. 1996). Likewise, data subsequently obtained for the sHLA transfectants utilized here according to established immunoprecipitation protocols (for example, Harris et al. 1998) indicates that although all four allotypes associate with calreticulin, B*1501, B*1503, and B*1508 do not associate with tapasin (and therefore not with TAP1/TAP2) whereas B*1510 does. Membrane-bound forms of B*1501 and B*1516 have previously been shown by others to not associate with TAP1/TAP2 (Neisig et al. 1996; de la Salle et al. 1997), demonstrating that results obtained

from the sHLA transfects are in concordance with those of native molecules.

Though functionally divergent according to its pooled motif (FIG. 16) and the majority of peptides that it binds, B*1510 is capable of accommodating ligands with the properties favored by the B*1501, B*1503, and B*1508 binding grooves. Data both from individual ligands (Table D) and fractional Edman sequencing (Table 3) indicate that Tyr can occupy the C-terminal position, and specific examples in Table 4, including the spleen mitotic checkpoint BUB3₅₃₋₆₀ octamer YQHTGAVL and the splicing factor U2AF large chain₁₇₉₋₁₈₇ nonamer TQAPGNPVL, attest to B-pocket flexibility. It is intriguing that among the peptides bound by B*1510 is the tapasin₃₅₄₋₃₆₂ nonamer HHSDGSVSL; the peptide appears to occupy ligand extracts in a high copy number, as qualitatively based upon relative mass spectrometric ion intensities. Given this, as well as considering potential models of loading complex interactions suggested by others (Neisig et al. 1996; Elliott 1997), it can be extrapolated that a portion of tapasin, analogous to class II-associated invariant chain-derived peptides (Riberdy et al. 1992; Sette et al. 1992), extends into and blocks a region of the empty class I binding groove until it is displaced by an optimally-fitting ligand and/or secondary chaperone; this could also account for the differences in overall P2 flexibility observed between B*1510 peptides and those of the other three allotypes. Participating in ligand selection by this mechanism would describe a distinct peptide editing role for tapasin and could

clarify the inability to detect overlaps between B*1510 and either B*1501, B*1503, or B*1508.

In addition to the initial search for overlaps across B*1501, B*1503, B*1508, and B*1510, a comparative analysis was performed between the ion maps of B*1501 and B*1512. As discussed previously hereinabove, such an examination is primarily important in revealing the presence of ligands bound by B*1512 but not B*1501 (Table 5). A number of overlapping ligands from B*1512, however, were additionally identified (Table 14). Conservative percentages of overlap subsequently observed among each of the four molecules from which ligands were characterized and the ancestral HLA-B15 allotype, B*1501, were determined as shown and described in Table 15.

As expected from an overview of their nearly identical ion maps, B*1501 and B*1512 demonstrated the highest overlap frequency between the allotypes at 70% among ions subjected to NanoES-MS/MS. After this, B*1503 and B*1508 respectively exhibited 14% and 9% overlap frequencies, while as shown earlier B*1510 completely failed to reveal overlaps with B*1501. The trend distinctly illustrates that the polymorphisms which distinguish the B*1503, B*1508, B*1510, and B*1512 peptide binding grooves from B*1501 are not functionally equivalent in terms of their impacts upon class I ligand association. However, it is also evident that they do not create concrete barriers to ligand binding. Because the repertoires of peptides bound by the

various molecules examined may differ from B*1501 at frequencies greater than 80% (B*1508 and B*1503, Table 15) does not mean that they are unable to bind similar or completely identical peptides, a concept which has been incompletely addressed and occasionally negated by other studies grounded more upon pooled Edman sequencing, analysis of prominent extract constituents, or binding/reconstitution assays.

In particular and based upon previous research, the overlaps between B*1508 and B*1501 defined here would specifically not have been predicted (Barber et al. 1997); it may be anticipated by extension that other molecules differing solely by the polymorphism separating B*1508 and B*1501, for example B*1503 and B*1529 (Appendix A), yield similar overlap frequencies. Likewise, based upon the relatively high overlap frequency observed between B*1512 and B*1501, the substitutions at positions 166 and 167 distinguishing them exhibit a similarly subtle effect between A*2902 and A*2903 (which only differ from one another by the identical substitutions studied here; Prokupek et al. 1997) via mapping and sequencing of individual ligands. Systematically attempting to define the limits of overlap existence as conducted, therefore, demonstrates a critical departure from standard approaches which enhance predicting the abilities of different class I molecules to present overlapping ligands.

Comparative analyses of closely related soluble MHC class I molecules

produced by the recombinant methods described herein, provide a means for assessing the functional impact of individual α -chain polymorphisms. The primary impetus for characterizing peptides extracted from class I molecules is to more precisely understand the influence of structural polymorphism upon the presentation of endogenous ligands. This is important since a fundamental realization of how naturally processed peptides bind to both individual and multiple class I allotypes can then be translated into protein and/or peptide-based therapies intended to elicit protective CTL responses. Therefore, an accurate interpretation of sequence data from such class I-bound peptides, either individual or pooled, should in turn further the selection of optimal viral and tumor-associated ligands to expedite the development of successful therapeutic applications.

The extensive examination of HLA-B15 ligands, as described herein, enhances understanding the rules that govern natural class I peptide presentation and is secondary evidence of the success and usefulness of the methodology for producing soluble MHC class I and II molecules described and claimed herein. By first building upon the traditional foundations provided by pooled Edman motifs (FIGS. 15 and 16), the data from over 400 individual peptides characterized from B*1501, B*1503, B*1508, B*1510, and B*1512 subsequently indicated that queries for potential epitopes specific to these allotypes would benefit from being optimized in three ways. First, although

nonamers represent half of the ligand population, the other 50% of peptide epitopes range down to 7 and up to 12 amino acids in length. Second, effective N-proximal anchor requirements need not be strictly imposed at P2. Third, searches for ligands should weigh C-terminal/proximal sequence matches even more heavily than those of the N-region. The third trend revealed represent, the most substantial of the revised search criteria, since both length variations and lower sensitivity due to the diminishing returns and increasing backgrounds inherent to successive Edman sequencing cycles can C-regional motif trends (Stevanovic and Jung 1993). As stressed earlier, examples of this bias were evident here in both: (i) the stronger preference by B*1510 for peptides terminating in Val (absent from the pooled motif) rather than Phe (present in the pooled motif) (FIGS. 16 and 24B); and (ii) the inability of motifs to effectively reflect C-proximal auxiliary anchors (FIGS. 15; 16; and 21-25, B).

To illustrate the potential consequences of applying the modified search parameters described, the EBV structural antigen gp85, which has recently been implicated using a murine model as a favorable target against which protective CTLs might be generated (Khanna et al. 1999), was examined in the context of B*1501 to identify: (i) nonameric epitopes with motif-prescribed P2 and P9 occupancies; (ii) length variant epitopes with motif-prescribed P2 and P9 occupancies; and (iii) nonameric epitopes with flexible P2 occupancy (Table 16). Since only these three categories of ligands were designated, the inquiry

was not exhaustive. However, the information extracted showed that, of the 98 possible epitopes identified, only the 22% within the first column would be placed under further experimental consideration if pooled motifs alone were applied in the search. This is not to imply that the data in this category is invalid but that it might be considerably incomplete for later applications. For example, if either of the AMTSKFLMGTY₁₇₂₋₁₈₂ (varying by length) or the SAPLEKQLF₁₂₃₋₁₃₁ (varying by P2 occupancy) peptides was demonstrated to elicit a more effective antigen-specific CTL response than any of the nonamers bearing standard motif P2/P9 assignments, this knowledge is pivotal to subsequent vaccine design; even if the two designated peptides evoked responses only equivalent to some of the nonamers, their non-motif length and/or sequences discrepancies could prove superior in conferring the ability to overlap multiple allotypes in addition to B*1501. This is advantageous since a vaccine consisting of a single or limited number of peptide specificities could theoretically be effective for protecting populations differing in HLA type (Loftus et al. 1995; Gulukota et al. 1996; Sidney et al. 1996c).

The majority of information collected from examining divergent HLA-B15 allotypes of sHLA molecules recombinantly produced according to the methodology of the present invention demonstrates that similar and occasionally identical peptide ligands are presented by the different B15 molecules so long as polymorphisms do not alter C-terminal anchoring pockets

and while an N-proximal ligand residue can be subsequently anchored within the binding groove. Supporting data furthermore indicates that these principles additionally extend beyond the HLA-B15 allotypes described in specificity herein. Specifically, unpublished results by Ghosh and Wiley (noted in Bouvier and Wiley 1994) indicate that an octamer has been observed to successfully bind a class I molecule by its C terminus despite being shown through x-ray crystallography to not even reach the N-terminal pocket of the binding groove. In addition, a recently-described HIV-gag₁₉₇₋₂₀₅ CTL epitope presented by murine class I K^d fails to show a motif-prescribed Tyr at P2 and instead associates stably through its conserved C terminus and an N-proximal preference for Gln at P3 (Mata et al. 1998). The rules established here through examination of hundreds of natural ligands from B*1501, B*1503, B*1508, B*1510, and B*1512 indicate that such occurrences may be more commonplace than exception, as both of these examples appear in agreement with the model in FIG. 27.

A step in developing therapies intended to elicit protective CTLs requires the selection of pathogen- and tumor-specific peptide ligands for presentation by MHC class I and class II molecules. Binding/reconstitution assays provide information that is biased due to their technical inconsistency and/or *in vitro* nature, while Edman sequencing of extracted class I peptide pools generates "motifs" that indicate that the optimal peptides are nonameric ligands bearing

conserved P2 and P9 anchors; motifs have frequently been used to provide the search parameters for selecting potentially immunogenic epitopes that might be successfully presented by particular allotypes (Pamer et al. 1991; DiBrino et al. 1994; Kast et al. 1994; Davenport et al. 1995; Walden 1996; Holland et al. 1997; Zhang et al. 1997; Yoon et al. 1998; Chang et al. 1999). Therefore, to test the hypothesis that natural presentation overlaps exist despite the presence of various polymorphisms within the class I binding groove and thus determine how well pooled motifs actually represent their endogenously-derived constituents, ligands were purified from different sHLA molecules produced in hollow-fiber bioreactors, mapped by RP-HPLC and NanoES-MS, and sequenced by NanoES-MS/MS, all according to the methodology of the present invention.

Production of sHLA provides an efficient means of extracting large quantities of endogenous peptide ligands for the subsequent analyses, and comparative ion mapping of peptides extracted from distinct class I allotypes is a reliable method for detecting potential ligand overlaps. NanoES-MS/MS analysis then allows for sequence characterization to identify the overlap status of individual ion matches. The strategy developed to address overlap identification is additionally pertinent beyond the uses described herein. For example, similar mapping studies would be performed, with the primary intent instead of characterizing differences between maps, such as between pathogenically infected versus uninfected cell lines; the data obtained could

contribute to identifying optimal vaccine epitopes. This concept is currently in the early stages of pursuit by several independent research teams (Veronese et al. 1996; van der Heeft et al. 1998). Successfully locating differences in a similar manner between B*1501 and B*1512 ion maps, as discussed herein, effectively supports this application.

FIG. 28 summarizes the α -chain substitutions and motif-derived P2 and P9 anchors for the HLA-B15 allotypes examined here, as well as additional allotypes indicated earlier in FIG. 5 that appear to serve as evolutionary intermediates between or extensions from B*1503, B*1510, B*1512, and B*1518. B*4601 is included since it differs from B*1501 by a single mutagenic event and overlaps with it were among peptides characterized in this study (Table 4). In section A, the allotypes B*1501, B*1503, B*1508, B*1512, and B*4601 have been shown to bind variously overlapping ligands; based upon the structurally-predicted motif anchors of the remaining molecules in this section including B*1519, B*1529, B*1539, and B*1547, it is suspected that ion mapping and characterization would reveal further overlaps according to the model shown in FIG. 27. Likewise, in section C the allotypes B*1509 and B*1510 have demonstrated overlapping ligands and will probably share some with B*1537.

Systematically mapping and characterizing 449 ligands from the related molecules B*1501, B*1503, B*1508, B*1510, and B*1512 demonstrates

overall that the peptides bound by these allotypes: (i) vary in length from 7 to 12 residues; and (ii) are more conserved at their C termini than at their N-proximal positions. Flexibility at P2 in particular appears to arise at least in part from the combined effects of distinct steric and charge biases imposed respectively by α -helical and β -sheet structural residues throughout α_1 and α_2 of the various HLA-B15 molecules, while it is postulated that C-terminal preferences are influenced by tapasin-moderated loading selection within the ER.

Although not predictable from the pooled Edman motifs, the comparative peptide mapping strategy succeeded in identifying endogenously processed ligands which bind variously across the allotypes B*1501, B*1503, B*1508, and B*1512, but not B*1510. Overlapping peptide ligands appeared to favorably bind the first four B15 molecules since these allotypes share identical C-terminal anchoring pockets, whereas B*1510 diverges in this region. Endogenous peptide loading into the HLA-B15 allotypes therefore requires that a conserved C terminus be firmly anchored in the appropriate specificity pocket while N-proximal residues act more flexibly in terms of both location and sequence specificity to anchor the ligand into this binding groove region. Subsequently, the choice of allele-specific and/or overlapping peptide epitopes for CTL recognition may thus be contingent upon performing queries strongly based upon conserved C-terminal anchors.

Another embodiment of the present invention, as previously described herein-above, is the use of genomic DNA (gDNA) as the starting material for the production of the sHLA molecules described hereinbefore.

This alternative method of the present invention begins by obtaining genomic DNA which encodes the desired MHC class I or class II molecule. Alleles at the locus which encode the desired MHC molecule are PCR amplified in a locus specific manner. These locus specific PCR products may include the entire coding region of the MHC molecule or a portion thereof. In some cases a nested or hemi-nested PCR is applied to produce a truncated form of the class I or class II gene so that it will be secreted rather than anchored to the cell surface. In other cases the PCR will directly truncate the MHC molecule.

Locus specific PCR products are cloned into a mammalian expression vector and screened with a variety of methods to identify a clone encoding the desired MHC molecule. The cloned MHC molecules are DNA sequenced to insure fidelity of the PCR. Faithful truncated (i.e. sHLA) clones of the desired MHC molecule are then transfected into a mammalian cell line. When such cell line is transfected with a vector encoding a recombinant class I molecule, such cell line may either lack endogenous class I expression or express endogenous class I. It is important to note that cells expressing endogenous class I may spontaneously release MHC into solution upon natural cell death. In cases where this small amount of spontaneously released MHC is a concern, the

transfected class I MHC molecule can be "tagged" such that it can be specifically purified away from spontaneously released endogenous class I molecules in cells that express class I molecules. For example, a DNA fragment encoding a His tail which will be attached to the protein may be added by the PCR reaction or may be encoded by the vector into which the gDNA fragment is cloned, and such His tail will further aid in purification of the class I molecules away from endogenous class I molecules. Tags beside a histidine tail have also been demonstrated to work and are logical to those skilled in the art of tagging proteins for downstream purification.

Cloned genomic DNA fragments contain both exons and introns as well as other non-translated regions at the 5' and 3' termini of the gene. Following transfection into a cell line which transcribes the genomic DNA (gDNA) into RNA, cloned genomic DNA results in a protein product thereby removing introns and splicing the RNA to form messenger RNA (mRNA), which is then translated into an MHC protein. Transfection of MHC molecules encoded by gDNA therefore facilitates reisolation of the gDNA, mRNA/cDNA, and protein.

Production of MHC molecules in non-mammalian cell lines such as insect and bacterial cells requires cDNA clones, as these lower cell types do not have the ability to splice introns out of RNA transcribed from a gDNA clone. In these instances the mammalian gDNA transfectants of the present invention provide a valuable source of RNA which can be reverse transcribed to form MHC cDNA.

The cDNA can then be cloned, transferred into cells, and then translated into protein. In addition to producing secreted MHC, such gDNA transfectants therefore provide a ready source of mRNA, and therefore cDNA clones, which can then be transfected into non-mammalian cells for production of MHC. Thus, the present invention which starts with MHC genomic DNA clones allows for the production of MHC in cells from various species.

A key advantage of starting from gDNA is that viable cells containing the MHC molecule of interest are not needed. Since all individuals in the population have a different MHC repertoire, one would need to search more than 500,000 individuals to find someone with the same MHC complement as a desired individual – this is observed when trying to find a match for bone marrow transplantation. Thus, if it is desired to produce a particular MHC molecule for use in an experiment or diagnostic, a person or cell expressing the MHC allele of interest would first need to be identified. Alternatively, in the method of the present invention, only a saliva sample, a hair root, an old freezer sample, or less than a milliliter (0.2 ml) of blood would be required to isolate the gDNA. Then, starting from gDNA, the MHC molecule of interest could be obtained via a gDNA clone as described herein, and following transfection of such clone into mammalian cells, the desired protein could be produced directly or in mammalian cells or from cDNA in several species of cells using the methods of the present invention described herein.

Current experiments to obtain an MHC allele for protein expression typically start from mRNA, which requires a fresh sample of mammalian cells that *express* the MHC molecule of interest. Working from gDNA does not require gene expression or a fresh biological sample. It is also important to note that RNA is inherently unstable and is not easily obtained as is gDNA. Therefore, if production of a particular MHC molecule starting from a cDNA clone is desired, a person or cell line that is expressing the allele of interest must traditionally first be identified in order to obtain RNA. Then a fresh sample of blood or cells must be obtained; experiments using the methodology of the present invention show that ≥ 5 milliliters of blood that is less than 3 days old is required to obtain sufficient RNA for MHC cDNA synthesis. Thus, by starting with gDNA, the breath of MHC molecules that can be readily produced is expanded. This is a key factor in a system as polymorphic as the MHC system; hundreds of MHC molecules exist, and not all MHC molecules are readily available from MRNA. This is especially true of MHC molecules unique to isolated populations or of MHC molecules unique to ethnic minorities. Starting class I or class II protein expression from the point of genomic DNA simplifies the isolation of the gene of interest and insures a more equitable means of producing MHC molecules for study; otherwise, one would be left to determine whose MHC molecules are chosen and not chosen for study, as well as to determine which ethnic population from which fresh samples cannot be

obtained should not have their MHC molecules included in a diagnostic assay.

While cDNA may be substituted for genomic DNA as the starting material, production of cDNA for each of the desired HLA class I types will require hundreds of different, HLA typed, viable cell lines, each expressing a different HLA class I type. Alternatively, fresh samples are required from individuals with the various desired MHC types. The use of genomic DNA as the starting material allows for the production of clones for many HLA molecules from a single genomic DNA sequence, as the amplification process can be manipulated to mimic recombinatorial and gene conversion events. Several mutagenesis strategies exist whereby a given class I gDNA clone could be modified at either the level of gDNA or at the cDNA resulting from this gDNA clone. The process of the present invention does not require viable cells, and therefore the degradation which plagues RNA is not a problem. Thus, from a given gDNA clone, any number of gDNA and cDNA MHC molecules can be produced.

Three useful products can be obtained from the mammalian cell line expressing HLA class I molecules from such a genomic DNA construct. The first product is the soluble class I MHC protein, which may be purified and utilized in various experimental strategies, including but not limited to epitope testing. Epitope testing is a method for determining how well discovered or putative peptide epitopes bind individual, specific class I or class II MHC proteins. Epitope testing with secreted individual MHC molecules has several advantages over the prior art, which utilized MHC from cells expressing multiple membrane-bound MHCs. While the prior art method could distinguish if a cell or cell lysate

would recognize an epitope, such method was unable to directly distinguish in which specific MHC molecule the peptide epitope was bound. Lengthy purification processes might be used to try and obtain a single MHC molecule, but doing so limits the quantity and usefulness of the protein obtained. The novelty of the current approach is that individual MHC specificities can be utilized in sufficient quantity through the use of recombinant, soluble MHC proteins. Because MHC molecules participate in numerous immune responses, studies of vaccines, transplantation, immune tolerance, and autoimmunity can all benefit from individual MHC molecules provided in sufficient quantity.

A second important product obtained from mammalian cells secreting individual MHC molecules is the peptide cargo carried by MHC molecules. Class I and class II MHC molecules are really a trimolecular complex consisting of an alpha chain, a beta chain, and the alpha/beta chain's peptide cargo to be reviewed by immune effector cells. Since it is the peptide cargo, and not the MHC alpha and beta chains, which marks a cell as infected, tumorigenic, or diseased, there is a great need to characterize the peptides bound by particular MHC molecules. For example, characterization of such peptides will greatly aid in determining how the peptides presented by a person with MHC-associated diabetes differ from the peptides presented by the MHC molecules associated with resistance to diabetes. As stated above, having a sufficient supply of an individual MHC molecule, and therefore that MHC molecules bound peptides, provides a means for studying such diseases. Because the method of the present invention provides quantities of MHC protein previously unobtainable,

unparalleled studies of MHC molecules and their important peptide cargo can now be facilitated.

The methodology for producing sHLA from gDNA, while similar to the methodology for producing sHLA from cDNA, is different and as such requires different and/or unique steps and/or processes for its completion. One exemplary detailed production methodology for use gDNA as the starting material for the production of MHC class I or II molecules is described herein below.

– Genomic DNA Extraction

Greater than or equal to 200ul of sample either blood, plasma, serum, buffy coat, body fluid or up to 5×10^6 lymphocytes in 200ul phosphate buffered saline was used to extract genomic DNA using the QIAamp DNA Blood Mini Kit blood and body fluid spin protocol. Genomic DNA quality and quantity was assessed using optical density readings at 260nm and 280nm

– PCR Strategy

Primers have been designed for HLA-A, -B and -C loci in order to produce a truncated amplicon of the human class I MHC using a two-stage PCR strategy. The first stage PCR uses a primer set that amplify from the 5' Untranslated region to Intron 4. This amplicon is used as a template for the second PCR which results in a truncated version of the MHC Class I gene by utilizing a 3' primer that sits down in exon 4 at codon 298 (including the leader peptide), the 5' primer remains the same as the 1st PCR. An overview of this PCR strategy can be seen in FIG. 29. The primers for each locus are listed in Table 17.

Different HLA-B locus alleles require primers with different restriction cut sites depending on the nucleotide sequence of the allele. Hence there are two 5' and two 3' truncating primers for the HLA-B locus.

1. Primary PCR Materials

An Eppendorf Gradient Mastercycler is used for all PCR. H₂O: Dionized ultra filtered water (DIUF) Fisher Scientific, W2-4,41. PCR nt mix (10mM each deoxyribonucleoside triphosphate [dNTP]), Boehringer Mannheim, #1814, 362. 10X *Pfx* Amplification buffer, pH 9.0, GibcoBRLR, part # 52806, formulation is proprietary information. 50mM MgSO₄, GibcoBRLR, part #52044. Platinum[®] *Pfx* DNA Polymerase (B Locus only), GibcoBRLR, 11708-013. *Pfu* DNA Polymerase (A and C Locus), Promega, M7741. *Pfu* DNA Polymerase 10x reaction Buffer with MgSO₄, 200mM Tris-HCL, pH 8.8, 100mM KCl, 100mM (NH₄)₂SO₄, 20mM MgSO₄, 1mg/ml nuclease free BSA, 1% TritonX-100. gDNA Template. Amplification primers (in ng/ul):

- a. A locus: 5' sense PP5UTA (300); 3' antisense PPI4A (300)
- b. B locus (Not B*39's): sense PP5UTB (300); antisense PPI4B (300)
- c. B locus (B*39's): sense 5UTB39 (300); antisense PPI4B (300)
- d. C Locus: sense 5PKCE (300); antisense PPI4C (300).

B. Secondary PCR (also used for colony PCR)

H₂O: Dionized ultra filtered water (DIUF) Fisher Scientific, W2-4,41. PCR nt mix (10mM each deoxyribonucleoside triphosphate [dNTP]), Boehringer Mannheim, #1814, 362. *Pfu* DNA Polymerase, Promega, M7741. *Pfu* DNA Polymerase 10x reaction Buffer with MgSO₄, 200mM Tris-HCL, pH 8.8, 100mM KCl, 100mM (NH₄)₂SO₄, 20mM MgSO₄, 1mg/ml nuclease free BSA, 1% TritonX-

100. Template 1:100 dilution of the primary PCR product. Amplification primers (in ng/ul):

- a. A-locus: 5' sense PP5UTA (300), 3' antisense PP3PEI (300)
- b. B-locus: sense PP5UTB (300), antisense PP3PEI (300)
- c. B-locus: sense PP5UT (300), antisense PP3PEIH (300)
- d. B-locus B39's: sense 5UTB39 (300), antisense PP3PEIH (300)
- e. C-locus: sense 5PKCE (300), antisense PP3PEI (300)
- f. C-locus Cw*7's: sense 5PKCE (300), antisense 3PEIHC7 (300)

C. Gel Purification of PCR products and vectors

Dark Reader Transilluminator Model DR-45M, Clare Chemical Research.
SYBR Green, Molecular Probes Inc. Quantum Prep Freeze 'N Squeeze DNA Gel
Extraction Spin Columns, Bio-Rad Laboratories, 732-6165.

D. Restriction digests, Ligation and Transformation

Restriction enzymes from New England Biolabs: EcoR I #R0101S; Hind
III #R0104S; Xba I #R0145S. T4 DNA Ligase, New England Biolabs,
#M0202S. pcDNA3.1(-), Invitrogen Corporation, V795-20. 10x Buffers from
New England Biolabs. EcoR I buffer, 500mM NaCl, 1000mM Tris-HCL, 10mM
MgCL₂, 0.25% Triton-X 100, pH 7.5. T4 DNA ligase buffer, 500mM Tris-HCL,
100mM MgCL₂, 100mM DTT, 10mM ATP, 250ug/ml BSA, pH 7.5. NEB buffer 2,
500mM NaCl, 100 mM Tris-HCl, 100mM MgCL₂, 10mM DDT, pH 7.9. 100x BSA,
New England Biolabs. Z-Competent *E. coli* Transformation Buffer Set, Zymo
Research, T3002. *E. coli* strain JM109. LB Plates with 100ug/ml ampicillin. LB
media with 100ug/ml ampicillin

E. Plasmid Extraction

Wizard Plus SV minipreps, Promega, #A1460

3. Sequencing of Clones

Thermo Sequenase Primer Cycle Sequencing Kit, Amersham Pharmacia Biotech, 25-2538-01. CY5 labeled primers (Table 18). Alfexpress automated DNA sequencer, Amersham Pharmacia Biotech.

4. Gel Casting

PagePlus 40% concentrate, Amresco, E562, 500ml. Urea, Amersham Pharmacia Biotech, 17-0889-01, 500g. N'N'N'N'-tetramethylethylenediamine (TEMED), APB. Ammonium persulphate (10% solution), APB. Boric acid, APB. EDTA-disodium salt, APB. Tris, APB. Bind-Saline, APB. Isopropanol, Sigma. Glacial Acetic Acid, Fisher Biotech. DIUF water, Fisher Scientific. Ethanol 200-proof.

5. Plasmid Preparation for Electroporation

Qiagen Plasmid Midi kit, Qiagen Inc., 12143.

A. Electroporation

Biorad Gene Pulser with capacitance extender, Bio-Rad Laboratories. Gene Pulser Cuvette, Bio-Rad Laboratories. Cytomix: 120mM KCl, 0.15mM CaCl_2 , 10mM $\text{K}_2\text{HPO}_4/\text{KH}_2\text{PO}_4$, pH 7.6, 25mM Hepes, pH 7.6, 2mM EGTA, pH 7.6, 5mM MgCl_2 , pH 7.6 with KOH. RPMI 1640+ 20% Foetal Calf Serum + Pen/strep. Haemocytometer. Light Microscope. CO_2 37° Incubator. Cells in log phase.

Methods used for the production of soluble human HLA Class I and II proteins in mammalian cells from gDNA.

1. Primary PCR

A. A-Locus and C-Locus

10x Pfu buffer	5ul
5' Primer (300ng/ul)	1ul
3' Primer (300ng/ul)	1ul
dNTP's (10mM each)	1ul
gDNA (»50ng/ul)	10ul
DIUF H ₂ O	31.4ul
Pfu DNA Polymerase	0.6ul

96°C 2 min.	
95°C 1 min	x35
58°C 1 min	x35
73°C 5 min	x35
73°C 10 min	

B. B-locus

10x Pfx buffer	5ul
5' Primer (300ng/ul)	1ul
3' Primer (300ng/ul)	1ul
dNTP's (10mM each)	1.5ul
MgSO ₄ (50mM)	1ul
gDNA (100ng/ul)	1ul
DIUF H ₂ O	40ul
Pfx DNA Polymerase	0.5ul
94°C 2 min.	
94°C 1 min	x35
60°C 1 min	x35
68°C 3.5 min	x35
68°C 5 min	

2. Gel Purification of PCR (all PCR and plasmids are gel purified)

Mix primary PCR with 5ul of 10x SYBR green and incubate at room

temperature for 15 minutes then load on a 2% agarose gel. Visualize on the Dark Reader and purify using the Quantum Prep Freeze and Squeeze extraction kit according to the manufacturers instructions.

3. Secondary PCR of A, B and C Loci

10x Pfu buffer	5ul
5' Primer (300ng/ul)	1.0ul
3' Primer (300ng/ul)	1.0ul
dNTP's (10mM each)	1ul
1:100 1° PCR	10ul
DIUF H ₂ O	31.4ul
Pfu DNA Polymerase	0.6ul
96°C 2 min.	
95°C 1 min	x35
56°C 1 min	x35
73°C 4 min	x35
73°C 7 min	

4. Restriction digests

2° PCR (gel purified)	30ul
Restriction enzyme 1	Xul
Restriction enzyme 2	Xul
10x buffer	5ul
100x BSA	0.5ul
DIUF H ₂ O	10.5ul

The cut sites incorporated into the PCR primers for each individual PCR will determine the enzymes used. The expression vector pcDNA3.1(-) will be cut in a similar manner.

5. Ligation

pcDNA3.1(-) cut with same enzymes as PCR	x ng
Cut PCR	y ng
10x T4 DNA ligase buffer	2ul
T4 DNA Ligase	1ul
DIUF H ₂ O	up to 20ul

The ratio of vector to insert will vary between samples, a good starting point is a ratio of 1:6

6. Transformation

Transform JM109 using competent cells made using Z-competent *E.coli* Transformation Kit and Buffer Set.

7. Colony PCR and restriction digests.

Either following the secondary PCR protocol or carrying out another restriction digest can be used to screen the vector for the appropriate insert.

8. Mini Preps of colonies with insert

Use the Wizard Plus SV minipreps and follow the manufacturers instructions. Make glycerol stocks before beginning extraction protocol.

9. Sequencing of positive clones and gel casting

Using the Thermo Sequenase Primer Cycle Sequencing Kit

A,C,G or T mix	3ul
CY5 Primer 1pm/ul	1ul
DNA template 100ng/ul	5ul
96°C 2 min	
96°C 30 sec	x25
61°C 30 sec	x25

Add 6 ul formamide loading buffer and load 10ul onto sequencing gel

Analyse sequence for good clones with no misincorporations.

10. Gel Casting

A. Prepare a 10X TBE stock solution for the sequencing gel mix:

	<u>500mL</u>
Tris	60.5g

EDTA	1.85g
Boric Acid	25.5g
Fisher DIUF H ₂ O	440mL

Filter using a 0.22m or 0.45m filter and store at 4° C until required.

11. Prepare a 10X TBE stock solution for the running buffer:

	<u>1L</u>
Tris	121.0g
EDTA	3.7g
Boric Acid	51.0g
Ultra pure H ₂ O	880mL

12. Prepare the sequencing gel mix:

	<u>1 Gel, 6% PagePlus</u>
Urea	19.8g
PagePlus 40% conc.	7.95mL
10X TBE	5.5mL
Fisher DIUF H ₂ O	25.3mL

Filter using a 0.22m or 0.45m filter.

Initiate polymerization of the sequencing gel by adding 330uL of a freshly made 10% APS solution and 33uL of TEMED. Cast a 0.5mm sequencing gel and allow it to polymerize for 5 hours.

13. Midi Preps

Prepare plasmid for electroporation using the Qiagen Plasmid Midi Kit according to the manufacturers instructions.

14. Electroporation

Electroporations are performed as described in "The Bw4 public epitope of HLA-B molecules confers reactivity with natural killer cell clones that express NKb1, a putative HLA receptor. Gumperz, J.E., V. Litwin, J.H. Phillips, L.L. Lanier and P. Parham. J. Exp. Med. 181:1133-1144, 1995," which is herein

expressly incorporated by reference in its entirety.

15. Screening for production of Soluble HLA

An ELISA is used to screen for the production of soluble HLA, see ELISA protocol.

ELISA Protocol

Solutions:

3 N H₂SO₄:

For 500 ml:	200 ml	H ₂ O
	300 ml	5 N H ₂ SO ₄

Store at room-temperature

10x PBS (pH 7.4):

26.8 mM	KCl
14.7 mM	KH ₂ PO ₄
1.37 M	NaCl
81 mM	Na ₂ HPO ₄

For 1000 ml:	2 g	KCl
	2 g	KH ₂ PO ₄ anhydrous
	80 g	NaCl
	11.5 g	Na ₂ HPO ₄

Add H₂O
Adjust pH to 7.4
Add up with H₂O to 1000 ml
Filter
Store at 4°C

TBS coating buffer (pH 8.5):

25 mM	Tris-HCl	pH 8.5
150 mM	NaCl	

For 1000 ml: 100 ml 10x TBS pH 8.5
900 ml H₂O

Store at 4° C

10 % BSA

For 250 ml: 25 g BSA

Add H₂O
Stir to dissolve
Add H₂O up to 225 ml
Filter
Store at 4° C
(Add 10x PBS prior to use)

ELISA WASH

2.68 mM KCl
1.47 mM KH₂PO₄
137 mM NaCl
8.1 mM Na₂HPO₄
0.05 % Tween-20

For 4000 ml: 400 ml 10x PBS
3598 ml H₂O
2 ml Tween-20

Store at 4°C

OPD Substrate Solution for HRP:

0.05 M Na₂HPO₄ pH 5.0
0.025 M Citrate
0.4 mg/ml OPD
0.012 % H₂O₂

For 10 ml: 10 ml 0.05 M Phosphate-citrate buffer pH 5.0
2 tablets OPD (2 mg ea) (Sigma; P-6787)
4 µl 30 % Hydrogen peroxide (H₂O₂) (Sigma; H1009)

Always prepare fresh
Use within 1 hour of preparation
Add fresh 30 % H_2O_2 immediately prior to use

0.05 M Phosphate-Citrate buffer:

0.05 M	Na_2HPO_4	pH 5.0
0.025 M	Citrate	

For 1000 ml:	7.10 g	Na_2HPO_4 anhydrous (Dibasic)
	5.25 g	Citric acid monohydrate

Add dd H_2O
Adjust pH to 5.0, if necessary
Add dd H_2O to a final volume of 1000 ml
Filter
Store at room temperature

The HLA ELISA procedure

For biochemical analysis, monomorphic monoclonal antibodies are particularly useful for identification of HLA locus products and their subtypes. W6/32 is one of the most common monoclonal antibodies (mAb) used to characterize human class I major histocompatibility complex (MHC) molecules. It is directed against monomorphic determinants on HLA-A, -B and -C heavy chains, which recognizes only mature complexed class I molecules and recognizes a conformational epitope on the intact MHC molecule containing both beta2-microglobulin (b2m) and the heavy chain (HC). W6/32 binds a compact epitope on the class I molecule that includes both residue 3 of beta2m and residue 121 of the heavy chain (Ladasky JJ, Shum BP, Canavez F, Seuanes HN, Parham P. Residue 3 of beta2-microglobulin affects binding of class I MHC

molecules by the W6/32 antibody. Immunogenetics 1999 Apr;49(4):312-20.). The constant portion of the molecule W6/32 binds to is recognized by CTLs and thus can inhibit cytotoxicity. The reactivity of W6/32 is sensitive to the amino terminus of human beta2-microglobulin (Shields MJ, Ribaudo RK. Mapping of the monoclonal antibody W6/32: sensitivity to the amino terminus of beta2-microglobulin. Tissue Antigens 1998 May;51(5):567-70). W6/32 is available biotinylated (Serotec MCA81B) offering additional variations in ELISA procedures.

Anti-human b2m (HRP) (DAKO P0174) recognizes denatured as well as complexed b2m. Although in principle anti-b2m reagents could be used for the purpose of identification of HLA molecules, they are less suitable when association of heavy chain and b2m is weak. The patterns of class I molecules precipitated with W6/32 and anti-b2m are usually indistinguishable [Vasilov, 1983 #10].

Rabbit anti-b2-microglobulin dissociates b2-microglobulin from heavy chain as a consequence of binding (Rogers, M.J., Appella, E., Pierotti, M. A., Invernizzi, G., and Parmiani, G. (1979) Proc Natl. Acad. Sci. U.S.A. 76, 1415-1419). It also has been reported that rabbit anti-human b2-microglobulin dissociates b2-microglobulin from HLA heavy chains upon binding (Nakamuro, K., Tanigaki, N., and Pressman, D. (1977) Immunology 32, 139-146.). This anti-human b2m antibody is also available unconjugated (DAKO A0072).

The W6/32-HLA sandwich ELISA

Sandwich assays can be used to study a number of aspects of protein

complexes. If antibodies are available to different components of a heteropolymer, a two-antibody assay can be designed to test for the presence of the complex. Using a variation of these assays, monoclonal antibodies can be used to test whether a given antigen is multimeric.

The W6/32 - anti-b2m antibody sandwich assay is one of the best techniques for determining the presence and quantity of sHLA. Two antibody sandwich assays are quick and accurate, and if a source of pure antigen is available, the assay can be used to determine the absolute amounts of antigen in unknown samples. The assay requires two antibodies that bind to non-overlapping epitopes on the antigen. This assay is particularly useful to study a number of aspects of protein complexes.

To detect the antigen (sHLA), the wells of microtiter plates are coated with the specific (capture) antibody W6/32 followed by the incubation with test solutions containing antigen. Unbound antigen is washed out and a different antigen-specific antibody (anti-b2m) conjugated to HRP is added, followed by another incubation. Unbound conjugate is washed out and substrate is added. After another incubation, the degree of substrate hydrolysis is measured. The amount of substrate hydrolyzed is proportional to the amount of antigen in the test solution.

The major advantages of this technique are that the antigen does not need to be purified prior to use and that the assays are very specific. The sensitivity of the assay depends on 4 factors:

- (1) The number of capture antibody
- (2) The avidity of the capture antibody for the antigen

- (3) The avidity of the second antibody for the antigen
- (4) The specific activity of the labeled second antibody

Use an ELISA protocol template and label a clear 96-well polystyrene assay plate. Polystyrene is normally used as a microtiter plate. (Because it is not translucent, enzyme assays that will be quantitated by a plate reader should be performed in polystyrene and not PVC plates).

Company	Plate	Specificity	Cat#
Nunc	Maxisorp StarWell Modules Framed 8-well strips	standard/un treated	441653

Coating of the W6/32 should be performed in Tris buffered saline (TBS); pH 8.5. Prepare a coating solution of 8.0 µg/ml of specific W6/32 antibody in TBS (pH 8.5). Although this is well above the capacity of a microtiter plate, the binding will occur more rapidly. Higher concentrations will speed the binding of antigen to the polystyrene but the capacity of the plastic is only about 100 ng/well (300 ng/cm²), so the extra protein will not bind. If using W6/32 of unknown composition or concentration, first titrate the amount of standard antibody solution needed to coat the plate versus a fixed, high concentration of labeled antigen. Plot the values and select the lowest level that will yield a strong signal. Do not include sodium azide in any solutions when horseradish peroxidase is used for detection.

Immediately coat the microtiter plate with 100 µl per well using a

multichannel pipet. Standard polystyrene will bind antibodies or antigens when the proteins are simply incubated with the plastic. The bonds that hold the proteins are non-covalent, but the exact types of interactions are not known. Shake the plate to ensure that the antigen solution is evenly distributed over the bottom of each well. Seal the plate with plate sealers (sealplate adhesive sealing film, nonsterile, 100 per unit; Phenix (1-800 767-0665); LMT-Seal-EX) or sealing tape to Nunc-Immuno™ Modules (# 236366). Incubate at 4°C overnight. Avoid detergents and extraneous proteins. Next day, remove the contents of the well by flicking the liquid into the sink or a suitable waste container. Remove last traces of solution by inverting the plate and blotting it against clean paper toweling. Complete removal of liquid at each step is essential for good performance. Wash the plate 10 times with Wash Buffer (PBS containing 0.05 % Tween-20) using a multi-channel ELISA washer.

After the last wash, remove any remaining Wash Buffer by inverting the plate and blotting it against clean paper toweling. After the W6/32 is bound, the remaining sites on the plate must be saturated by incubating with blocking buffer made of 3% BSA in PBS. Fill the wells with 200 µl blocking buffer. Cover the plates with an adhesive strip and incubate overnight at 4°C. Alternatively, incubate for at least 2 hours at room temperature which is, however, not the standard procedure. Blocked plates may be stored for at least 5 days at 4°C. Good pipetting practice is most important to produce reliable quantitative results. The tips are just as important a part of the system as the pipette itself. If they are of inferior quality or do not fit exactly, even the best pipette cannot

produce satisfactory results.

When maximum levels of accuracy are stipulated, prewetting should be used at all times. To do this, the required set volume is first drawn in one or two times using the same tip and then returned. Prewetting is absolutely necessary on the more difficult liquids such as 3% BSA. Do not prewet, if your intention is to mix your pipetted sample thoroughly with an already present solution. However, prewet only for volumes greater than 10 μ l. In the case of pipettes for volumes less than 10 μ l the residual liquid film is as a rule taken into account when designing and adjusting the instrument. The tips must be changed between each individual sample. With volumes <10 μ l special attention must also be paid to drawing in the liquid slowly, otherwise the sample will be significantly warmed up by the frictional heat generated. Then slowly withdraw the tip from the liquid, if necessary wiping off any drops clinging to the outside. To dispense the set volume hold the tip at a slight angle, press it down uniformly as far as the first stop.

In order to reduce the effects of surface tension, the tip should be in contact with the side of the container when the liquid is dispensed. After liquid has been discharged with the metering stroke, a short pause is made to enable the liquid running down the inside of the tip to collect at its lower end. Then press it down swiftly to the second stop, in order to blow out the tip with the extended stroke with which the residual liquid can be blown out. In cases that are not problematic (e.g. aqueous solutions) this brings about a rapid and virtually complete discharge of the set volume. In more difficult cases, a slower

discharge and a longer pause before actuating the extended stroke can help. To determine the absolute amount of antigen (sHLA), sample values are compared with those obtained using known amounts of pure unlabeled antigen in a standard curve.

For accurate quantitation, all samples have to be run in triplicate, and the standard antigen-dilution series should be included on each plate. Pipetting should be preformed without delay to minimize differences in time of incubation between samples. All dilutions should be done in blocking buffer. Thus, prepare a standard antigen-dilution series by successive dilutions of the homologous antigen stock in 3% BSA in PBS blocking buffer. In order to measure the amount of antigen in a test sample, the standard antigen-dilution series needs to span most of the dynamic range of binding. This range spans from 5 to 100 ng sHLA/ml.

A stock solution of 1 µg/ml should be prepared, aliquoted in volumes of 300 µl and stored at 4°C. Prepare a 50 ml batch of standard at the time. (New batches need to be compared to the old batch before used in quantitation). Use a tube of the standard stock solution to prepare successive dilutions according to the scheme below. While standard curves are necessary to accurately measure the amount of antigen in test samples, they are unnecessary for qualitative "yes/no" answers.

For accurate quantitation, the test solutions containing sHLA should be assayed over a number of at least 4 dilutions to assure to be within the range of the standard curve. Prepare serial dilutions of each antigen test solution in

blocking buffer (3% BSA in PBS). Standard dilutions for purified, crude or flow through samples are given below:

After mixing, prepare all dilutions in disposable U-bottom 96 well microtiter plates before adding them to the W6/32-coated plates with a multipipette. Add 150 μ l in each well. To further proceed, remove any remaining blocking buffer and wash the plate as described above. The plates are now ready for sample addition. Add 100 μ l of the sHLA containing test solutions and the standard antigen dilutions to the antibody-coated wells. Cover the plates with an adhesive strip and incubate for exactly 1 hour at room temperature. After incubation, remove the unbound antigen by washing the plate 10x with Wash Buffer (PBS containing 0.05 % Tween-20) as described.

Prepare the appropriate developing reagent to detect sHLA. Use the second specific antibody, anti-human b2m-HRP (DAKO P0174 / 0.4 mg/ml) conjugated to Horseradish Peroxidase (HRP). Dilute the anti-human b2m-HRP in a ratio of 1:1'000 in 3% BSA in PBS. (Do not include sodium azide in solutions when horseradish peroxidase is used for detection).

	No. of plates	Total Antibody	anti-b2m-HRP	3%BSA in PBS
Mix: 1		10 ml	10 μ l	10 ml
2		20 ml	20 μ l	20 ml
3		30 ml	30 μ l	30 ml
4		40 ml	40 μ l	40 ml
5		50 ml	50 μ l	50 ml

Add 100 μ l of the secondary antibody dilution to each well. All dilutions should be done in blocking buffer. Cover with a new adhesive strip and

incubate for 20 minutes at room temperature. Prepare the appropriate amount of substrate prior to the wash step. Bring the substrate to room temperature. OPD (o-Phenylenediamine) is a peroxidase substrate suitable for use in ELISA procedures. The substrate produces a soluble end product that is yellow in color. The OPD reaction is stopped with 3 N H_2SO_4 , producing an orange-brown product and read at 492 nm. Prepare OPD fresh from tablets (Sigma, P6787; 2 mg/tablet). The solid tablets are convenient to use when small quantities of the substrate are required.

After second antibody incubation, remove the unbound secondary reagent by washing the plate 10x with Wash Buffer (PBS containing 0.05% Tween-20). After the final wash, add 100 μl of the OPD substrate solution to each well and allow to develop at room temperature for 10 minutes. Reagents of the developing system are light-sensitive, thus, avoid placing the plate in direct light. Prepare the 3 N H_2SO_4 stop solution. After 10 minutes, add 100 μl of stop solution per 100 μl of reaction mixture to each well. Gently tap the plate to ensure thorough mixing. Read the ELISA plate at a wavelength of 490 nm within a time period of 15 minutes after stopping the reaction.

The background should be around 0.1. If the background is higher, the substrate may have been contaminated with a peroxidase. If the substrate background is low and the background in your assay is high, this may be due to insufficient blocking. Finally analyze your readings. Prepare a standard curve constructed from the data produced by serial dilutions of the standard antigen. To determine the absolute amount of antigen, compare these values

with those obtained from the standard curve.

After running the ELISA on various transfectants it is possible to determine which ones will produce the highest levels of soluble HLA molecules. These "good producers" are grown to the point where a cell pharm can be seeded. The harvest from the cell pharm is then used to extract the soluble HLA using the following purification protocol.

To start the chromatography procedure, prepare the ÄKTA™ prime system. The system can be used immediately but the spectrophotometers full ability will not be obtained until after 1 hour of lamp warm-up. To prepare the system for a run, check that the buffer inlet tubings are immersed in the correct buffer vessels and the waste tubings are put into a waste bottle. Only use degassed and filtered liquids to make sure that the liquid remains free from air bubbles. Degass by applying a vacuum to the solution. Prepare and hook up the buffers necessary for a sHLA purification:

Buffer Valve (A)

- | | |
|--------------------------------------|--|
| 1. PBS, pH 7.4 | (Wash buffer) |
| 2. 20 % Ethanol / 70 % Ethanol | (Cleaning solutions) |
| 3. 0.1 N NaOH | (MOK elution buffer) |
| 4. 50 mM Diethylamine (DEA), pH 11.3 | (MOK elution buffer) |
| 5. Protein sample | (The line is stored in PBS/0.05% Na Azide, pH 7.4) |
| 6. 0.2 N Acetic acid, pH ~2.7 | (Cleaning & MOK solution) |
| 7. 0.1 M Citric acid, pH 3.0 | (Protein A elution buffer) |
| 8. 0.1 M Glycine, pH 11.0 | (sHLA elution buffer) |

It is important to purge the lines after a new hook-up with about 50 ml of liquid to get the air out of the system. Purging can be done manually through

the inlets of the buffer valve (A1-A8), while carefully immersing the tubing in the respective liquid. To remove any trapped air bubbles in the flow path, purge the pump in the order PBS / 20 % ethanol / PBS / final buffer solution. Next, prepare the recorder to monitor the purification. Autozero the built-in UV spectrophotometer with PBS as reference. Equilibrate all material to the temperature at which the chromatography will be performed. For large scale purifications, attach the column entrance/exit to the system.

Equilibrate the column by passing 10 bed-volumes of PBS over the matrix. Before starting any column purification, the protein concentration in the sample solution should be determined using a quantitative ELISA procedure. The sample volume loaded will depend on the size and loading capacity of the column and the concentration of the sample. Calculate the volume of the sample solution maximally saturating the column according to the columns capacity to bind the antigen.

(A) Antigen concentration:	mg/ml antigen
(B) Antigen binding capacity:	mg antigen/ml gel
(C) Matrix volume:	ml gel
(D) Maximal amount of antigen: (B*C)	mg
(E) Sample volume: (D/A)	ml

Since the binding capacity of the column will realistically not be reached, a much lower volume of sample solution should be chosen. A value between 40 to 50% of the calculated volume is more accurate which also will not result in the waste of lots of unbound antibody within the flow-through. Prepare the antibody sample solution for purification. Spin crude harvest at 5'000 rpm for 25 minutes (JA10 rotor) to remove lipid and cell debris. The antigen solution

must be free of particulate matter. Pour the supernatant into a suitable container. Prevent air bubble formation.

Name of the crude harvest:

Volume used: ml

Amount of sample: mg

The simplest method to bind the antigen to the antibody/Sepharose 4B matrix is to apply the sample through the system pump and pass the protein solution down the column. Set appropriate parameters to record the loading conditions on the recorder.

	Chart Speed	Conductivity	Optical Density
Load	0.1 mm/min	0.5 V	1.0 V

Save a 1 ml probe from the starting material (LOAD) before the purification procedure for analysis purposes. Set the buffer valve to position 5 and the injection valve to position LOAD. Make sure the inlet tubings are purged with sample buffer without any air-bubbles present. To have a purged sample line, disconnect shortly the column before loading and circulate the sample within the system with higher flow rate. Pass the solution slowly through the column with a flow rate of approximately 1.0 ml/min or lower to give the protein time to bind more efficiently. Higher flow rates will decrease efficiency. A disruption in flow may cause a rapid rise in back-pressure. If this occurs, immediately shut off the pump and check the gel bed for compression.

Collect the flow-through in an appropriate container. Keep until you are

sure all material has bound to the column and negligible amounts are in the flow through. Take a sample at the end of the run (Ft) which should be analyzed. Wash the column with PBS at 10 ml/min until UV absorbance at 280 nm is zero. For a large column use 2000-3000 ml wash buffer (PBS). Save the wash in a container until after the purification.

	Chart Speed	Conductivity	Optical Density
Wash	0.5 mm/min	0.5 V	1.0 V

Prepare borosilicate collection tubes by adding 1.2 ml of 1 M Tris-HCl, pH 7.0 per 4.8 ml of fraction to be collected (1:4). Neutralization is a safety measure to preserve the activity of the eluted molecule. MHC class I (SHLA) molecules are best eluted from a W6/32 column by 0.1 M glycine, pH 11.0. Absorbance is used for generating a protein elution profile.

	Chart Speed	Conductivity	Optical Density
Elution	0.5 mm/sec	0.2 V	0.1 V

Place the collector arm over the first collection tube. Elute 4.8 ml per fraction at 10 ml/min. Immediately afterwards, mix each tube gently to bring

the pH back to neutral. As with all protein solutions, avoid bubbling or frothing as this denatures the proteins. If a very low amount of protein is expected, change the conductivity on the recorder to a lower value. Identify the antigen-containing fractions by absorbance at 280 nm on the chart and combine them during up-concentration. Up-concentrate immediately and buffer exchange into PBS using MACROSEP™ centrifugal concentrators (Pall Filtron; Northborough, MA; MACROSEP 10K; OD010C37). Keep the protein on ice at all times and centrifuge at 4°C. After the buffer exchange, prepare the sample for storage at 4°C. Filter the pure samples through a 0.2 µ filter and aliquot directly into sterile, screw cap tubes. Label appropriately.

Determine the absorbance at 280 nm as well as the protein concentration with the Micro BCA kit. Activity can be determined with a regular ELISA procedure. The purity of the eluted sHLA can be assessed by SDS-PAGE, Western blotting or performing a Superdex column analysis. After the elution, quickly re-equilibrate the column with PBS to avoid denaturation of the W6/32 antibody linked to it. For analytical work in which more than one allele will be purified on the same column, extreme care must be taken. To be able to re-use the column, start a maintenance procedure after the re-equilibration. Cleaning-in-place is a procedure, which removes contaminants such as lipids, precipitates or denatured proteins that may remain in the column after regeneration. Such contaminations are especially likely when working with crude materials. The procedure helps to maintain the capacity, flow properties and general performance.

Mock elute the column using buffers with alternating pH. Start running over 10 gel volumes of 0.2 N acetic acid followed by 10 gel volumes of 50 mM diethylamine, pH 11.3 at a speed of 10 ml/min. Repeat three times and always equilibrate with 10 gel volumes PBS between buffer changes.

	Chart Speed	Conductivity	Optical Density
Mok-elution	1.0 mm/min	0.2 V	0.1 V

After Mok-elution, store the column at 4°C in PBS/0.05% Na Azide. Sanitization inactivates microbial contaminants in the packed column and related equipment. One generally recommended procedure is to wash alternately with high and low pH buffers as performed in the coupling reaction. For sanitization, disassemble the column and wash the matrix alternately with low pH wash buffer (0.1 M sodium acetate containing 0.5 M NaCl, pH 4.0) and high pH wash buffer (0.1 M Tris-HCl containing 0.5 M NaCl, pH 8.0) for 3 times followed by re-equilibration with PBS. Re-assemble the cleaned and sterilized column and store it at 4°C in PBS containing 0.05% sodium azide.

After the column is removed, the ÄKTA™ prime system has to be cleaned carefully. Start with the cleaning of line 5, where the sample was hooked up. Rinse the system pump and include the fraction collector line. First clean the inlet tubings, by manually running the system pump and flushing with 0.2 N acetic acid at 30 ml/min followed by 0.1 N NaOH. Always equilibrate with PBS.

Don't forget to add a line between the injection valve and the UV detector as a bridge, as replacement of the column. Finally, rinse with 20% ethanol. If the column was sanitized because of bacterial contamination, rinse with 70% ethanol.

In order to test whether or not the sHLA molecules produced were conformational, an assay was developed using a variety of different antibodies:

1. w6/32, recognizes conformational intact trimer
2. hc10, recognizes denatured HLA molecules
3. anti b_2M , recognizes free and non-covalently associated Beta 2 microglobulin

The methodology for this assay is essentially the same as the standard Elisa described above, except that instead of coating the plate with w6/32, the soluble HLA molecules are coated directly to it. The three different antibodies are then utilized in three different ways to detect the bound HLA. W6/32 is biotin labeled, therefore Vectastain kit and OPD are used for detection, anti b_2M is conjugated to horse radish peroxidase and so can be directly detected using OPD and with hc10 a secondary anti-mouse IgG horse radish peroxidase conjugated antibody and OPD are used.

Results for sHLA Production Using gDNA as the Starting Material

Using sample 3A394

Genomic DNA extraction.

Using 200ul whole blood, a Qiagen DNA extraction was performed. Table

19 shows the optical density readings and concentration of this extraction.

Primary PCR of 3A394

10x pfu buffer	5ul
PP5UTA	1ul
3PPI4A	1ul
dNTP	1ul
gDNA	10ul
H ₂ O	31.4ul
Pfu	1ul

See FIG. 30 for the gel image of this PCR.

FIG 30. 2% agarose gel showing the primary PCR product of 3A394 (5th lane from the right) at approximately 2kb in size.

The PCR product was gel purified and this along with unpurified PCR was used in the secondary PCR at a 1:100 dilution.

Secondary PCR of 3A394

10x pfu buffer	5ul
dNTP	1ul
PP5UTA	1ul
PP3PEI	1ul
1:100 1 ^o PCR	10ul
H ₂ O	31.4ul
Pfu	0.6ul

The results of this can be seen in FIG. 31. Secondary PCR of 3A394. Gel purified and unpurified can be seen in lanes 2 and 3 of FIG. 31.

The PCR product derived from gel purified primary PCR was in turn gel purified and used in the restriction digest in order to ligate into pcDNA 3.1(-), FIG. 32.

FIG. 32 is a gel showing 3A394 and pcDNA3.1 digested with EcoR I and Xba I. Once the ligations, at three different insert to vector ratios, and transformation were completed colonies were picked, grown overnight and then the plasmid DNA was extracted. A restriction digest was performed to screen for insert as shown in FIG. 33 is a restriction digests of 3A394 clones. A vector to insert ratio of 1:6 is the most efficient.

Following identification of positive clones the concentration of the vector was established by optical density (Table 20) and then diluted to 500ng/ul.

Cycle sequencing was then performed on sample 3A394TPC1 and it was established that this was a good clone of HLA-A*1102 truncated at codon 298.

The clone 3A394PC1 was then grown up and the plasmid containing A*1102 Truncated (A*1102T) was prepared for electroporation by performing a large scale plasmid extraction. The concentration of this was determined using optical density which can be seen in Table 21.

30ug of this was then electroporated into the cell line LCL 721.221 with the decay times being recorded then, after 2 days the viability of the cells was determined, Table 22.

The cells were then placed under G418 selection to screen for positive transfectants, those exhibiting G418 resistance were screened for soluble HLA production using the Elisa, Table 23. Only two replicates were tested for this sample and all were done using undiluted supernatant.

Samples 3A394TPC1 wells 1 and 2 were grown further and eventually well 1 cells were seeded into a cell pharm. This protocol has been performed on

many HLA types as can be seen by the production of soluble HLA molecules in Table 24.

Due to the fact that the monoclonal antibody w6/32 only recognizes the folded trimer complex of heavy chain-light chain-peptide the results of the ELISA demonstrate that we are producing conformational molecules originally derived from genomic DNA.

FIG. 34 is a graph showing the comparative binding of three monoclonal antibodies to four different soluble HLA molecules. A*1102T is sample 3A394TPC1well 1 from genomic DNA the other three are produced from cDNA. Three different amounts of soluble HLA were coated to the plate for each allele.

The results of the comparative binding assay demonstrate several properties of the soluble HLA produced from genomic DNA. Coating a plate with more protein will not necessarily yield a higher signal to protein ratio. Soluble HLA from genomic DNA gives results comparable to that of cDNA constructs. The fact that all three antibodies bind this confirms the correct epitopes of the recombinant molecules are present.

Due to the polymorphic nature of the HLA system production of many alleles as soluble molecules is very difficult as a viable cell line is required in order to make cDNA and quite often this is not available. Thus a method that allows us to produce many different alleles from a readily available starting point is invaluable. Production of the sHLA from genomic DNA provides such a starting point. We have shown here that two simple PCR reactions allows us

to clone many, if not all, HLA Class I alleles from genomic DNA.

The Elisa data allows us to test test how functional these molecule are. By using w6/32 and anti b₂M to establish production levels we also provide information as to how much of the protein is in a trimeric form. The comparative Elisa data helps back this up as the ratio of w6/32:hc10 needs to be greater than 1.0 in order for there to be more conformational molecule than denatured, this is shown to be the case. In summary we have developed a technique that will allow the production of virtually any HLA Class I molecule in a soluble form on demand.

An exemplary useful product which can be obtained from the mammalian cell line expressing such a genomic DNA construct is a cDNA clone encoding the desired class I or class II molecule. The cDNA clone encoding the desired class I or class II molecule is formed from the mRNA molecule encoding the desired class I molecule isolated from such mammalian cell line. The cDNA clone may be utilized for functional testing, as described in more detail herein below. Thus, gDNA clones can be used as a mechanism to obtain cDNA clones of the desired class I or class II HLA molecule.

The cDNA clones may be transfected into a cell which is unable to splice introns and process the mRNA molecule and therefore would not express the MHC molecule encoded by the genomic DNA, such as insect cells or bacterial cells. In addition, these cell lines will also be deficient in peptide processing and loading, and therefore the soluble MHC molecules expressed from such cells will not contain peptides bound therein (referred to as free heavy chain HLA). Such

soluble, free heavy chain HLA can effectively be tested for epitope binding as well. That is, MHC made in cells which do not naturally load peptide can be experimentally loaded with the peptide of choice. The heavy chain, light chain, peptide trimer can be reassembled *in vitro* using a high affinity peptide to facilitate assembly. Alternatively, a cell deficient in peptide processing can be pulsed with peptide such that the trimolecular MHC complex forms. DNA encoding a peptide (also encoding an appropriate targeting signal) could also be co-transfected into the cell with the MHC so that the MHC molecule which emerges from the cell is loaded only with the desired peptide. In this way MHC molecules could be loaded with a single low affinity peptide so that replacement with test peptides in a binding assay are more controlled.

Note that an advantage of secreting individual MHC molecules from a cell that naturally loads peptide is that the MHC molecule of interest is naturally loaded with thousands of different peptides. When used in a peptide binding assay, a synthetic peptide can therefore be compared to thousands of naturally loaded peptides.

Another use for the sHLA produced functional testing, according to the methodology of the present invention is the peptide-MHC complex can be multimerized to form soluble peptide-MHC dimers or tetramers, or other multiple soluble peptide-MHC - mers, such as fivemers, sixmers, etc. which serve as ligands for CTLs. The tetramers can be mixed with CTLs *in vitro* or with CTLs from the blood of human subjects to identify antigenic peptides responsible for immune responses in humans. Altman et al (Science, 1996),

herein expressly incorporated by reference in its entirety, discloses a method of functional testing using tetramer technology; however, the method of Altman, however, only discloses one soluble MHC molecule which has been utilized in such a method, and Altman's method faces the same disadvantages and defects described above for the prior art, that is, the method envisions isolating individual mRNA/cDNA molecules from hundreds of different, typed cell lines, and then manipulating the cDNA molecules to produce the desired soluble MHC molecule. The methods of the present invention envision combining the tetramer technology with amplification of genomic DNA, cloning the genomic DNA fragment, and transfection of the resulting construct into a mammalian cell line followed by isolation of cDNA from such transfected cell line and transfection into a cell line deficient in peptide processing and loading, thereby removing the need to isolate hundreds of different, typed cell lines for obtaining the different cDNAs.

MHC/peptide tetramers are widely utilized in the phenotypic analysis of T cells and in the study of T cell responses to pathological conditions such as viral infections and cancer. Current methodology for tetramer production consists of expressing the MHC class I heavy chain in bacterial or insect cells and refolding the heavy chain in the presence of β -2-microglobulin and a specific peptide ligand in vitro.

The methodology of the present invention had two specific aims, although this should not be regarded as limiting: 1) to engineer a cDNA construct of a

class I heavy chain containing a BirA substrate peptide (bsp) sequence at its 3' end (C-terminus) which would enable its subsequent biotinylation and 2) to develop a novel means of tetramer production using a mammalian expression system. The mammalian system used was a B cell/T cell hybrid, the antigen-processing mutant cell line CEM x 721.174.T2 (T2). When pulsed with established HLA B*0702-presented peptides from HIV-infected CD4+ T cells, T2 cells transfected with the recombinant, truncated B*0702 HLA heavy chain secreted specific MHC/peptide complexes. Following enzymatic biotinylation, these complexes were combined with avidin to form B7 tetramers. A mammalian expression system affords several advantages over a prokaryotic system, such as allowing normal glycosylation of the class I heavy chain and eliminating the need to refold the MHC/peptide complex in vitro following expression. The MHC class I molecules are therefore naturally folded in the cell rather than artificially folded outside the cell. Indeed, such artificial folding of the MHC class I molecules outside of the cell results in a MHC class I molecule which differs from one found in a "natural" system (i.e. a human immune system) and as such, is not an appropriate basis upon which to conduct vaccine development.

Abbreviations: MHC, major histocompatibility complex; β_2m , β_2 -microglobulin; HLA, human leukocyte antigen; sHLA, soluble human leukocyte antigen; bsp, BirA substrate peptide or biotinylation substrate peptide; CTL,

cytotoxic T lymphocytes; PCR, polymerase chain reaction; ELISA, enzyme-linked immunosorbent assay; HRP, horseradish peroxidase; PE, phycoerythrin; PBS, phosphate buffered saline; TAP, Transporters associated with Antigen Processing; TCR, T cell receptor; ER, endoplasmic reticulum; kDa, kiloDaltons.

The structure and function of MHC class I. Unlike B lymphocytes which can interact with antigen in its intact form, T lymphocytes only recognize protein antigen broken down into peptide fragments and presented in association with specialized cell-surface molecules. These molecules are the gene products of the major histocompatibility complex (MHC) (FIGS. 1 and 35), a region on chromosome six known to encode proteins that are critical for immunologic specificity and transplantation histocompatibility. MHC molecules, also termed human leukocyte antigens (HLA), are cell-surface glycoproteins that function primarily in communicating to T cells the presence of intracellular or extracellular invaders. MHC/peptide complexes are recognized by the T cell receptor (TCR), an interaction enabling T cells to discriminate between MHC molecules bearing "foreign" antigen (viral/tumor/bacterial peptides) and MHC molecules displaying "self" peptides. An immune response can then be initiated against cells determined to harbor foreign antigen on the basis of the avidity of this binding interaction.

MHC molecules are of two types, designated class I and class II. In general, class II molecules are located on antigen presenting cells (APCs), such

as macrophages or dendritic cells, and present peptides from exogenously synthesized protein, usually the breakdown products from phagocytosed bacteria or protozoa. Thus, class II molecules primarily function to warn the immune system of extracellular invaders. The class II/peptide complex is recognized by and interacts with TCRs of CD4+ T cells (T helper or Th cells). Class I molecules are located on platelets and all nucleated cells of the body and bind peptides derived from endogenously synthesized proteins. Examples of endogenously synthesized proteins which may be broken up and presented by class I molecules include both "self" proteins, such as those normally produced in healthy cells, and "foreign" proteins, such as the viral proteins in virus-infected cells or the abnormal proteins produced in tumor cells. Thus, class I molecules primarily function to report to the immune system the health of the intracellular environment. The TCRs of CD8+ cytotoxic T lymphocytes (CTLs) interact with the MHC class I/peptide complex.

Structurally, class I molecules are heterodimers comprised of two noncovalently bound polypeptide chains, a larger "heavy" chain (α) and a smaller "light" chain (β -2-microglobulin, or β_2m). The polymorphic, polygenic heavy chain (45 kDa), encoded within the MHC on chromosome six, is subdivided into three extracellular domains (designated α_1 , α_2 , and α_3), one intracellular domain, and one transmembrane domain (FIG. 36). The two outermost extracellular domains, α_1 and α_2 , together form the groove that

binds antigenic peptide (FIG. 37). Thus, interaction with the TCR occurs at this region of the protein. The α_3 domain of the molecule contains the recognition site for the CD8 protein on the CTL; this interaction serves to stabilize the contact between the T cell and the APC.

The invariant light chain (12 kDa), encoded outside the MHC on chromosome 15, consists of a single, extracellular polypeptide. In the recombinant truncated HLA class I molecule (soluble HLA or sHLA, approximately 47 kDa), the C-terminal intracellular and transmembrane domains of the heavy chain are not amplified by PCR and thus deleted, resulting in a functional MHC/peptide complex that is secreted from the cell and still capable of interaction with the TCR.

Antigen processing and assembly of the MHC class I/peptide complex. The heavy chain of the class I heterodimer is cotranslationally inserted into the lumen of the ER (FIG. 38). The extracellular (intralumenal) domains of newly synthesized α chains are glycosylated and become immediately associated with the ER chaperone protein calnexin. Calnexin is a membrane-bound molecule that functions to temporarily keep the heavy chain in a partially folded state. The subsequent noncovalent interaction of free β_2m with the heavy chain causes the release of calnexin, and the heavy chain/ β_2m complex becomes sequentially associated with the chaperones calreticulin and tapasin. These chaperones position the class I heterodimer in a manner that enables it to be

loaded with the processed peptides once they are transported into the ER by the TAP (transporter associated with antigen processing) complex, located within the ER membrane.

In the cytosol, a multisubunit, multicatalytic protease complex called the proteasome functions to prepare the endogenously synthesized proteins for presentation by MHC class I molecules. Once proteins (viral or otherwise) are cleaved into peptides of various lengths by the proteasome, the TAP complex actively transports the peptides into the ER where the MHC heterodimer awaits. In the ER lumen, peptides are further trimmed to lengths of 8-11 amino acids. If possessing the requisite binding affinity to the particular class I allele, a peptide will be loaded into the binding cleft of the class I heavy chain to form the MHC heavy chain/light chain/peptide heterotrimeric complex, which is subsequently routed to the cell surface via the Golgi apparatus (FIG. 39).

Cells incapable of antigen processing, for reasons such as inhibition of the proteasome or mutation of TAP, will likewise be deficient in the cell-surface expression of the class I/peptide complex since the class I heterodimer cannot leave the ER until it binds peptide. A particular cell line called T2, which is TAP-deficient and therefore has no cell-surface class I expression, is widely utilized in antigen processing studies. Additionally, it has been shown that abundant exogenous peptide can be pinocytosed, enter an intracellular trafficking pathway, and be delivered to the ER compartment directly in a TAP-

independent way. High affinity peptides delivered to the ER in this manner can then bind to *de novo*-synthesized MHC class I molecules and be subsequently displayed at the cell surface. Thus, T2 cells, the system of mammalian expression utilized in the experiments herein, are capable of cell surface MHC class I expression when pulsed with exogenous peptide of sufficient affinity for the particular class I allele.

The interaction of the MHC/peptide complex with the TCR is the central event in the initiation of most antigen-specific immune responses. The TCR recognizes the antigen complex by forming intermolecular contacts with both the class I molecule and the antigenic peptide. The outcome of this interaction (i.e. whether or not an immune response is generated) is dependent upon the density and duration of the TCR-MHC/peptide binding. Thus, an immunogenic MHC/peptide complex will bind the TCR with greater avidity than a non-immunogenic MHC antigen complex. Furthermore, it is known that a monomeric MHC/peptide complex, even if immunogenic, dissociates rapidly from a TCR, indicating that multiple TCRs must interact with multiple MHC/peptide complexes in order to activate T cells. In light of this information, Altman et al. disclosed a suggested methodology to analyze antigen-specific T cell populations by multimerizing the MHC/peptide complex into tetramers. Essentially, a tetramer consists of four biotinylated MHC/peptide complexes non-covalently bound to an avidin molecule (FIG. 40). Compared to a

monomeric MHC/peptide complex, tetramers have slower rates of dissociation from CTLs since they are able to bind more than one TCR on that particular CTL. This unique characteristic makes tetramers very useful as immunological stains.

Until now, tetramers have incorporated MHC/peptide complexes that were made by expressing a recombinant class I heavy chain in bacteria and subsequently refolding the heavy chain in vitro in the presence of β_2m and a specific peptide ligand. The purpose of this project was to make tetramers using MHC/peptide complexes synthesized in mammalian cells, specifically cells of the antigen-processing mutant cell line T2 so that every MHC/peptide complex secreted would contain the same antigenic peptide. It is desirable to reflect as much as possible the actual *in vivo* interaction between human cells expressing cell-surface MHC/peptide complexes and cytotoxic T lymphocytes.

The three following oligonucleotide primers, designated A, B, and C, were purchased from commercially available sources (i.e. operon):

Primer A, a 5' primer:

5'___GGGCCTCGAGGGACTCAGAATCTCCCCAGACGCCGAG___3'

Primer B, a 3' primer:

5'___CCGCAAGCTTCCATCTCAGGGTGAGGGGCT___3'

Primer C, a 3' primer:

5'___CCGCGAATTCTTATTCGTGCCATTCGATTTTCTG___3'

Primers A and B were used in PCR #1 and the template cDNA was a recombinant sHLA-B*0702 truncated gene with a 6-histidine tail in pcDNA3.1(-), a mammalian expression vector. PCR #1 was designed to incorporate 5' *Xho*

I (CTC GAG) and 3' *Hind* III (AAG CTT) cut sites and to amplify the truncated B*0702 heavy chain lacking a 3' stop codon. All PCR reactions for this project were performed with a proofreading taq polymerase (PFU Polymerase, Promega) and were subjected to 25 cycles of 95°C for 1 minute (strand separation), 59°C for 1 minute (primer annealing), and 72°C for 2 minutes (DNA synthesis), followed by a final extension time of 7 minutes at 72°C. The product of PCR #1 was purified using a QIAquick PCR purification kit (QIAGEN) and was subsequently digested for 2 hr at 37°C with *Xho* I and *Hind* III. Concurrently, a bacterial expression vector (C-Terminal Biotin AviTag Vector, Avidity; catalog number pAC-6) (FIG. 41) encoding the biotinylation substrate peptide sequence GLNDIFEAQKIEWHE was digested with the same restriction enzymes under the same conditions. The digest products were gel purified using a Freeze N Squeeze kit (BioRad) and ligated together for 2.5 hours at room temperature. Competent cells of *E. coli* strain JM109 were transformed with the ligated DNA, plated on LB/ampicillin agar, and incubated for 16 hours at 37°C. Using colony PCR, colonies were screened for insert of the gene into the vector.

The B*0702t-no stop/AviTag vector DNA was isolated and prepared using a DNA Miniprep kit (Promega) and served as the template cDNA for PCR #2. Primers A and C were used in PCR #2, which was designed to maintain a 5' *Xho* I cut site, incorporate a 3' *EcoR* I cut site (GAA TTC) distal to the bsp, and

amplify the B*0702t gene with the bsp on its 3' end. The PCR product was purified with the QIAquick purification kit and, along with the mammalian expression vector pcDNA3.1(-), was digested with *Xho* I and *Eco*R I restriction enzymes. Digest products were gel purified (Freeze N Squeeze), ligated together, and transformed into competent JM109 *E. coli* cells, which were then incubated overnight at 37°C on LB/ampicillin agar. Colony PCR was performed to check selected colonies for insert of the B*0702t-bsp gene into pcDNA(-). DNA from clones with insert was prepared (Miniprep kit) and sequenced using cycle sequencing. Sequences were analyzed and a good clone was identified. The plasmid DNA of the good clone was isolated and prepared for transfection using a DNA Midiprep kit (QIAGEN).

Cells of the human cell line T2 were cultured in RPMI 1640 media + 20% fetal calf serum, 1% penicillin/streptomycin, and 0.25% phenol red. A total of 1.7×10^7 T2 cells were transfected with 30 µg of B*0702t-bsp/pcDNA3.1(-) DNA by electroporation using the Gene Pulser (BioRad) at 0.25 V and 960 µFD. Transfected cells were selected in a medium containing 40% RPMI 1640, 40% conditioned media, 20% fetal calf serum, 2% penicillin/streptomycin, 0.2% phenol red, and 1.5 mg/mL G418 neomycin (Cellgro). Surviving cells were pulsed with the synthetic HIV GAG peptide NH₂-S-P-R-T-L-N-A-W-V-COOH at 20 µg/mL and then incubated for 24 hours at 37°C. Transfectants were then screened by ELISA using W6/32 (8 µg/mL), which is directed against the entire

HLA heavy chain/light chain (β_2m)/peptide complex, as the primary (capture) antibody and anti- β_2m conjugated to HRP (diluted 1:1000) as the secondary (conjugate) antibody (Dako).

High-producing cells were then expanded and cultured (continuously being repulsed with peptide), and their supernatant was collected and centrifuged to remove cell debris. The specific MHC/peptide complexes were purified using an affinity chromatography system (AKTA™ prime). In brief, the supernatant was passed over a 38 mL bed volume XK 26 column of cyanogen bromide-activated Sepharose 4B (Amersham Pharmacia Biotech) coupled to W6/32. The protein was then eluted intact in basic buffer (0.1 M glycine-NaOH, pH 11.0). To neutralize the eluate, 1.0 M Tris was added in a 1 to 4 ratio of neutralization buffer:elution buffer, resulting in a final solution of 0.25 M tris/0.075 M glycine, pH ~8.0. Immediately, the eluted protein was concentrated and buffer-exchanged to PBS + 0.05% Na Azide using MACROSEP centrifugal concentrators (Pall Filtron, MACROSEP 10K). Purified protein was quantified using ELISA.

The purified sHLA-bsp/peptide complex was enzymatically biotinylated on a single lysine residue within the bsp by incubation with BirA (Avidity; other names for this enzyme include biotin protein ligase, biotin ligase, biotin operon repressor protein, biotin holoenzyme synthetase, and biotin-[acetyl-CoA carboxylase] synthetase) (FIG. 42).

Incubation time was varied under constant conditions to determine the time required for maximum biotinylation efficiency. Four identical reactions were incubated for either 1, 4, 8, or 16 hours at 37°C. Biotinylation mixture components were as follows: sHLA-bsp, 6 μ M; MgOAc (Biomix B, Avidity), 1.9 mM; adenosine triphosphate (Biomix B, Avidity), 1.9 mM; BirA, 0.8 μ M; Tris-HCl, pH 8.0, 9.4 mM; biotin (Biomix B, Avidity), 10 μ M; Pefabloc SC Plus (Roche), a protease inhibitor, 0.42 mM. Biotinylation was confirmed using a modified ELISA, with W6/32 (8 μ g/mL) as the capture antibody and ABC Vectastain (Vector Laboratories), a kit containing avidin and biotinylated HRP, as the conjugate (FIG. 43).

Biotinylated sHLA was separated from free biotin by applying the biotinylation reaction product to a Bio-Spin chromatography column (BioRad, Bio-Spin 30 Tris columns). Tetrameric sHLA/peptide complexes were produced by the stepwise addition (1/10th volume, waiting 10 minutes between each addition) of the conjugate UltraAvidin-R-phycoerythrin (UltraAvidin-R-PE; Leinco Technologies) to the biotinylated class I complex. The final mixture contained a 1:4 molar ratio of UltraAvidin-R-PE:biotinylated class I, which is the same as a 1:1 molar ratio UltraAvidin-R-PE biotin binding sites:biotinylated class I. The volumes of avidin and biotinylated class I were determined by calculating the molar concentrations of each (in moles/ μ L) and then making certain that the number of moles of biotin binding sites in the avidin equaled the number of

moles of biotinylated class I. (Because the biotinylation assay was qualitative rather than quantitative, the assumption that biotinylation efficiency was high (~90%) was made prior to making these calculations.) Tetramers were purified by gel filtration on a Superdex S-200 column (Pharmacia; Molecular Biology Resource Facility, University of Oklahoma Health Sciences Center).

The construct successfully created using the AviTag bsp vector contained a 7-residue sequence (W-K-L-P-A-G-G) between the truncated B7 heavy chain at its C-terminus and the bsp. Despite the fact that the peptide binding groove is at the N-terminus of the heavy chain, it was undetermined at the time of transfection whether this 7-residue linker would in any way affect MHC protein folding or peptide binding capability. Because class I molecules must be properly folded and loaded with peptide before being directed to the cell surface, only the specific MHC/peptide complexes should be in the supernatant. Additionally, the W6/32 monoclonal capture antibody of the ELISA positively identified the presence of MHC-bsp/peptide complexes secreted into the supernatant by producing transfectants (FIG. 44). Thus, the 7 residue linker did not alter protein folding or peptide loading.

With regard to the purification of the class I-bsp molecule via affinity column chromatography, the bsp addition to the molecule perhaps affected column loading, as only ~50% recovery of the purified protein was achieved when pre-column and post-column protein amounts were compared. A large

percentage of this "lost" protein was detected both in the wash, indicating non-specific binding to the column during loading, and in the flow-thru, possibly indicating that the additional 22 amino acids (15 amino acids in the bsp and 7 amino acids linking the bsp to the class I, discussed above) on the C-terminus of the molecule interfered with loading. If the latter occurred, then ELISA results quantifying sHLA-bsp production by T2's may be underestimated, since the same W6/32 antibody used as the capture in the ELISA is also coupled the column. The elution curve (FIG. 45), produced as a result of the spectrophotometric (OD280) measurements of the column system during elution, allowed determination of which eluted fractions contained protein. These fractions were then pooled and the purified protein was concentrated and biotinylated.

In a 1993 paper, Peter Shatz demonstrated that an array of BirA substrate peptide sequences, all having in common a lysine residue at position 9 or 10, existed and were capable of being biotinylated. However, some bsp sequences biotinylate more robustly than other sequences. It is important, therefore, to determine the reaction time required to optimally biotinylate a given protein with its specific bsp, so as to not waste the protein in future biotinylation reactions. FIG. 46 demonstrates that increasing the incubation time of the biotinylation reaction, with all other variables unchanged, allows for greater biotinylation.

Importantly, purifying the biotinylated class I molecules from any free biotin remaining in the reaction mixture was performed prior to making tetramers, as free biotin may assume one or more of the four biotin binding sites on the avidin molecule in place of a biotinylated class I molecule. FIG. 47 indicates that some biotinylated protein was lost in this step. However, this purification ideally eliminates any "contamination" from monomers, dimers, or trimers when making tetramers, optimizing tetramer yield.

Upon purification of the tetramers via gel filtration (Superdex-200), tetramers (MW 498.6 kDa) were separated on the basis of size from any other possibly-present molecule contained in the reaction mixture [free avidin-PE conjugate (300 kDa—avidin 60 kDa and Phycoerythrin 240 kDa); free biotinylated class I (49.65 kDa); unbiotinylated class I-bsp, since biotinylation efficiency was not likely 100% (49.4 kDa); free biotin, despite the purification step (243 Daltons); monomers (349.65 kDa); dimers (399.3 kDa); or trimers (448.94 kDa)].

The present use of tetramers as reagents to stain T cells specific to an epitope is widely known and utilized. Additionally, the use of tetramers in vaccine development and immune modulation is a reality in present biomedical research. Several potential applications of tetramers in modulating the human immune system include tolerance induction in autoimmune diseases and adoptive transfer of antigen-specific T cells in the clinic. If the specific epitope

causing the immune response in an autoimmune disease is known, a tetrameric complex specific to the patient's HLA type and containing the immunogenic epitope could be made and, in theory, given therapeutically to the patient in a dose large enough to induce tolerance. Liposomes, which are artificial antigen presenting cells that can be made and have incorporated in their membranes specific MHC/peptide complexes, are a cousin to the tetramer and would likely be used, rather than tetramers, for this type of immune modulation. Adoptive transfer of T cells specific to an antigen of a disease-causing agent (tumor or viral) could be achieved by using tetramers (specific to the patient and the antigen) to sort live T cells, culturing these T cells *in vitro* to increase their numbers, and then transferring the cells back into the patient in hopes of enhancing the patient's immune response.

Thus, as the potential therapeutic use of tetramers in the clinic becomes a reality, it is evident that a tetramer molecule containing MHC molecules expressed in mammalian cells would more so reflect the protein produced *in vivo*. Although it is not known for certain the significance of the role of MHC glycosylation in *in vivo* intra- and inter-molecular interactions, it is important both in research and clinical therapy to use a system that emulates the actual interaction as much as possible. Likewise, it is not known if the class I trimeric complex formed inside mammalian cells is structurally identical to that formed outside the cell, and for this reason we choose to load sHLA class I molecules

inside the cell. In summary, we show that class I molecules can be loaded with peptide inside a mammalian cell, secreted from the cell, biotinylated, and that these biotinylated class I molecules interact with streptavidin.

Alternatively, a cell line deficient in peptide processing but still efficient in peptide loading may be utilized for both epitope and functional testing, so that a putative epitope can be expressed or pulsed into a cell and loaded into the HLA molecule in the ER of such cell. The cDNA construct isolated as described above may be ligated into a mammalian expression vector which also contains a DNA fragment encoding a peptide of interest attached to a fragment encoding a signal peptide so that the peptide of interest will be retained in the ER of the cell for loading, and such construct transfected into the mammalian cell line deficient in peptide processing but which retains the ability to load peptide in the HLA molecules, such as the T2 cell line. In this manner, the peptide of interest is produced together with the HLA molecule. The soluble HLA molecule (with or without a His or biotinylation signal tail) can then be purified and utilized as a reagent that has been produced in mammalian cells (fully glycosylated, etc.) and is loaded with the single co-transfected peptide. Optionally, random oligomers could be made and cloned into such a mammalian expression vector, and the soluble HLA molecules could again be purified and used to characterize T cells or other immune effector cells. In a further alternative, rather than expressing the peptides with the HLA molecule, the cells

expressing the HLA molecule could be pulsed with a single synthetic peptide or multiple synthetic peptides and analyzed as described above to identify bound peptides. Any of the HLA molecule-peptide complexes could be multimerized to form dimers, tetramers, etc. and tested for their ability to serve as ligands for CTLs and induce immune responses in humans.

In summary, the method of the present invention involves production of MHC class I and class II molecules beginning from gDNA and/or cDNA. The gDNA clones encoding a given MHC molecule can be truncated to be secreted rather than bound at the cell surface. This truncated version of the MHC molecule can be produced in mammalian or insect/bacterial cells such that milligram or greater quantities of an individual class I or class II molecule can be obtained. The secreted MHC class I molecules can be naturally loaded with thousands of endogenous peptides in mammalian cells, while the secreted MHC class II molecules can be naturally loaded with thousands of endocytic peptides in mammalian cells. Alternatively, the secreted MHC proteins can be produced in cells that do not load the MHC molecule with peptide ligand. Production of MHC proteins in cells which do not load the MHC molecule with peptide facilitates the loading of the MHC molecule via co-transfection with constructs encoding a given peptide(s). Alternatively, the MHC peptide-loading deficient MHC transfectant can be pulsed with peptides or DNA encoding peptides. The resulting individual secreted MHC molecules are useful for studies of peptide

loading (i.e. in vaccine development), for characterizing human immune responses to a given MHC molecule loaded with a particular peptide(s), and to the development of diagnostics where one needs sufficient MHC protein in order to directly assess reactivity to different MHC proteins.

Another important component of the secreted MHC molecules described here is that naturally loaded peptides can be eluted from the MHC molecules and characterized. Substantial quantities of peptide can be obtained from individual MHC molecules, and the peptides can be selectively characterized. Unique information results from having a sufficient supply of eluted peptide, and this information is essential to databases and predictive algorithms which are essential to the vaccine architect.

Thus, in accordance with the present invention, there has been provided a methodology for producing and manipulating Class I and Class II MHC molecules from gDNA as well as the sHLA molecules and their uses that fully satisfies the objectives and advantages set forth herein above. Although the invention has been described in conjunction with the specific drawings, experimentation, results and language set forth herein above, it is evident that many alternatives, modifications, and variations will be apparent to those skilled in the art. Accordingly, it is intended to embrace all such alternatives, modifications and variations that fall within the spirit and broad scope of the invention.

All of the numerical and quantitative measurements set forth in this application (including in the examples and in the claims) are approximations.

The invention illustratively disclosed or claimed herein suitably may be practiced in the absence of any element which is not specifically disclosed or claimed herein. Thus, the invention may comprise, consist of, or consist essentially of the elements disclosed or claimed herein.

The following claims are entitled to the broadest possible scope consistent with this application. The claims shall not necessarily be limited to the preferred embodiments or to the embodiments shown in the examples.

	24	45	46	63	67	116	156	166	167	ethnicity
consensus	A	M	A	E	S	S	W	E	W	
B*1512	-	-	-	-	-	-	-	D	G	Thai
B*1508	-	-	-	N	F	-	-	-	-	Iranian/Indian/Amerindian
B*1501	-	-	-	-	-	-	-	-	-	Caucasian
B*1503	S	E	E	-	-	-	L	-	-	Black
B*1518	S	E	E	N	C	-	L	-	-	Indian
B*1510	S	E	E	N	C	Y	L	-	-	Black

TABLE 1

primer	type	sequence (5'→3')
HLA5UT	PCR (5'; inserts <i>Sal</i> I site)	GGGCGTCGACGGACTCAGAATCTCCCCAGACGCCGAG
sHLA3TM	PCR (3'; inserts stop codon and <i>Hind</i> III site)	CCGCAAGCTTTTCATCTCAGGGTGAG
5PXI	PCR (5'; inserts <i>Xba</i> I site)	GGGCTCTAGAGGACTCAGAATCTCCCCAGACGCCGAG
3PEI	PCR (3'; inserts stop codon and <i>Eco</i> RI site)	CCGCGAATTCTCATCTCAGGGTGAG
M13 universal	sequencing (mp18, end through α_3) (mp19, leader through α_2)	TGTAAACGACGGCCAGT
3S	sequencing (α_2 through α_3)	CGGCAAGGATTACATCGCCCTG
JD3S	sequencing (α_3 through end)	CCCCATCGTGGGCATCGTTG
3N	sequencing (α_2 through leader)	CAGGGCGATGTAAATCCTTGCCG
4N	sequencing (α_3 through α_2)	GCCAGGTCAGTGTGATCTCCGC
T7 promoter	sequencing (T7 promoter forward priming site)	TAAACGACTCACTATAGGG
pcDNA3.1/BGH	sequencing (BGH reverse priming site)	TAGAAGGCACAGTCGAGG

TABLE 2

allele	# fractions	P2 extras	P9 extras	>9 cycles?
B*1501	7	P	-	yes (14)
B*1508	8	QVKRS	IVMQ	yes (14)
B*1503	3	P	MNL	yes (14)
B*1510	3	PR	MIY	yes (14)

TABLE 3

ligand	source protein	allele(s) characterized from
<i>HLA ligands</i>		
VGYYDDDTQF	HLA-I α (49-57)	B*1501, 1508
IAGVYVDDTQF	HLA-I α (47-57)	B*1501, B*1512
IKADHVSTY	HLA-II DP α (32-40)	B*1503
GSHSMRYF	HLA-I α (25-32)	B*1503
<i>Replication/transcription/translation ligands</i>		
GQRKGAGSVF	60S ribosomal protein L8 (7-16)	B*1501, 1503
AQAESLRY	40S ribosomal protein S3 (100-107)	B*1501
GKVRTDITY	40S ribosomal protein S4 (73-81)	B*1503
SHAQTVVL	40S ribosomal protein S27 (48-55)	B*1510
SQFGGGSQY	eIF3-p66 (61-69)	B*1501, 1503, 1508, B*1512
VQGPVGTDf	zinc finger transcription factor (296-304)	B*1501
APPPPPPPP	transcription factor ZFM1 (581-589)	B*1501
YQHTGAVL	spleen mitotic checkpoint BUB3 (53-60)	B*1510
AHGRKMSKSL	valyl-tRNA synthetase (859-868)	B*1510
LPHQPLATY	Oct-binding factor 1 (52-60)	B*1508
AKYSTPATL	probable ATP-dependent RNA helicase DDX10 (280-288)	B*1503
AKAGITTTTL	DNA replication licensing factor MCM5 (470-478)	B*1503
TQAPGNPVL	splicing factor U2AF large chain (179-187)	B*1510
SHQRQLLL	Kin17 (49-56)	B*1510
NQFQALLQY	polypyrimidine tract-binding protein (220-228)	B*1512
<i>Biosynthetic/degradative modification ligands</i>		
FVSNHAY	aldolase A (358-364)	B*1501, 1508
ILGPPGSVY	ubiquitin-protein ligase (83-91)	B*1501, B*1502, 1508, B*1512
YMIDPSGVSY	proteasome subunit C8 (150-159)	B*1501, B*1502, 1508, B*4601, B*1512
NHAIVSTSV	26S protease (S4) regulatory subunit (741-749)	B*1510
IHTPENPVI	lanosterol 14- α demethylase (488-496)	B*1510

TABLE 4

AHSNLASVL	O-linked GlcNAc transferase (237-245)	B*1510
<i>Signalling/modulatory ligands</i>		
VVAPITTG Y	calcyclin binding protein (63-71)	B*1501, 1508
GHSPPTSSL	tyrosine-protein kinase JAK3 (491-499)	B*1510
LPPPPPPPP	Fas antigen ligand (54-62)	B*1503
NHANGTL	serine/threonine protein phosphatase PP2A (α and β) (229-236)	B*1510
<i>Transporter/chaperone ligands</i>		
EHVASSPAL	13S Golgi transport complex 90 kD subunit (741-749)	B*1510
HHSDGSVSL	tapasin (354-362)	B*1509, B*1510
QPGPQIVY	GABA/noradrenaline transporter (261-268)	B*1503
<i>Structural/cytokinesis ligands</i>		
NMNDLVSEY	tubulin β chain (414-422)	B*1508
THTQPGVQL	septin 2 homolog (70-78)	B*1509, B*1510
SHANSAVVL	β -adaptin (249-257)	B*1509, B*1510
<i>Unknown function ligands</i>		
GQYPTQPTY	KIAA0058 (5-13); like <i>Mus musculus</i> proline-rich protein	B*1503
VKVIQQESY	mammary tumor-associated protein INT6 (278-286)	B*1503
AKYPHVEDY	Ki nuclear autoantigen (207-215)	B*1503
AMNPTNTVF	heat shock cognate 71 kD protein (60-68)	B*1503
CPLSCFT	human HTGS database	B*1501, B*1503, B*1508
MPHSGYGF	human EST	B*1508
CHSAFAL	human HTGS database	B*1510
LHLLTLEA	human EST	B*1510
KNANLVQLY	human EST	B*1512

TABLE 4 cont'd.

fraction	ion for MS/MS	derived peptide sequence
7	504.1 (+2)	H M S G Z P T S Y
7	549.2 (+2)	H N Z A A H Z E Y
8	526.0 (+2)	H A A X Y S Z V Y
10	484.3 (+2)	Y Q S D H R Y
11	424.3 (+2)	H X S T Z D F
11	464.3 (+2)	H A P P T D P P
11	550.0 (+2)	H G P A N R D S V F
11	563.3 (+2)	F P Y P T D P Z Y
12	531.2 (+2)	Z N A N X V Z X Y
14	585.6 (+2)	R S F X X E N E Y
16	488.7 (+2)	H M Z N P T S Y
16	661.9 (+2)	Y V X F - - - V Y
17	577.6 (+2)	R S M X R C P E Y
18	523.0 (+2)	- - F V T A Z T Y
20	582.4 (+2)	M Y N C N E X D Y
25	562.8 (+2)	N Q F Q A L L Q Y

TABLE 5

ALLELE	7	9	24	25	26	34	35	36	45	62	63	66	67	70	99	159	163	167	P2 MOTIF (dominant/strong)
<i>consensus</i>	Y	Y	A	V	G	V	R	F	M	R	E	I	S	N	Y	Y	L	W	
B*1508	-	-	-	-	-	-	-	-	-	-	N	-	F	-	-	-	-	-	PA
B*1513	-	-	-	-	-	-	-	-	-	-	N	-	-	-	-	-	-	-	ILQVP
B*1502	-	-	-	-	-	-	-	-	-	-	N	-	-	-	-	-	-	-	LVQP
B*1501	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	QMLV
B*1512	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	G	QLM
B*1503	-	-	S	-	-	-	-	-	E	-	-	-	-	-	-	-	-	-	QKM
B*1518	-	-	S	-	-	-	-	-	E	-	N	-	C	-	-	-	-	-	H
B*1509	-	-	S	-	-	-	-	-	E	-	N	-	C	-	-	-	-	-	H
B*1510	-	-	S	-	-	-	-	-	E	-	N	-	C	-	-	-	-	-	H
B*1517	-	-	-	-	-	-	-	-	-	-	-	N	M	S	-	-	-	-	TS
B*1516	-	-	-	-	-	-	-	-	-	-	-	N	M	S	-	-	-	-	T
B*4601	-	-	-	-	-	-	-	-	-	-	-	K	Y	Q	-	-	-	-	MI

Table 6

fraction	ion for MS/MS	derived peptide sequence
9	490.3 (+2)	A G G Z P A T P P A X
9	513.1 (+2)	S H Z G C V Z P A V
10	433.8 (+2)	G H D P D S P A A
10	455.4 (+2)	E H V A S S P A L
10	482.6 (+2)	M C Z - G M P A X
10	482.8 (+2)	G H G A N N D P A X
10	495.7 (+2)	X H S Z P A G P A X
11	448.9 (+2)	M H A D N P V X
11	482.8 (+2)	G H C P R N P A X
11	495.7 (+2)	X H S G A P Z A P X
11	516.7 (+2)	X H D T E H A P X
12	448.4 (+2)	T Q A P G N P V L
12	460.3 (+2)	T Z A G C M V P X
13	464.8 (+2)	M V - - H P V X
14	456.7 (+2)	A H S V P S P A F
14	477.7 (+2)	M H T - - P A P V
14	482.8 (+2)	P G A A V V P S X
15	510.1 (+2)	I H T P E N P V I
16	456.7 (+2)	S H D G S V P T X
16	522.7 (+2)	- - - - - P V X
16	523.3 (+2)	M A H S - - P V F
17	523.2 (+2)	- H - - - - P V F
18	474.8 (+2)	M X G X S F P A X
18	491.2 (+2)	V H T C V N P V X
18	515.8 (+2)	E W H Y P V S X
19	496.6 (+2)	E T P E H A P V X

TABLE 8

side chain	P1	P2	P3	P4	C ⁻³	C ⁻²	C ⁻¹	C-term	side chain
G	-	-	11.90%	10.32%	11.90%	-	-	-	G
P	-	10.32%	-	-	-	-	-	-	P
C	-	-	-	-	-	-	-	-	C
A	18.25%	11.11%	11.90%	-	-	-	10.32%	-	A
V	11.90%	-	-	-	-	10.32%	-	-	V
F	-	-	-	-	-	-	-	26.19%	F
M	-	-	-	-	-	-	-	-	M
I/L	13.49%	13.49%	-	-	-	-	-	-	I/L
S	-	-	-	-	-	-	-	-	S
T	-	-	-	-	-	13.49%	-	-	T
Y	-	-	-	-	-	-	-	63.49%	Y
N	-	-	-	-	-	-	-	-	N
W	-	-	-	-	-	-	-	-	W
Q/K	-	24.60%	-	-	-	-	15.87%	-	Q/K
H	-	-	-	-	-	-	-	-	H
R	-	-	-	-	-	-	-	-	R
D	-	-	-	-	-	-	-	-	D
E	-	-	-	-	-	-	10.32%	-	E
N value	13.64	19.52	3.80	0.32	1.90	3.81	6.51	69.68	C value
N _{sum}	37.28				81.90				C _{sum}
B*1501 N _{sum} /C _{sum} = 0.46									

TABLE 9

side chain	P1	P2	P3	P4	C ⁻³	C ⁻²	C ⁻¹	C-term	side chain
G	12.16%	-	12.16%	13.51%	12.16%	-	-	-	G
P	-	10.81%	-	12.16%	10.81%	-	-	-	P
C	-	-	-	-	-	-	-	-	C
A	31.08%	-	-	-	-	-	-	-	A
V	-	-	-	-	-	-	-	-	V
F	-	-	-	-	-	-	-	20.27%	F
M	-	-	-	-	-	-	-	-	M
I/L	-	-	-	-	-	13.51%	-	-	I/L
S	-	-	-	-	-	10.81%	-	-	S
T	-	-	-	-	-	13.51%	17.57%	-	T
Y	-	-	-	-	-	-	-	68.92%	Y
N	-	-	-	-	-	-	-	-	N
W	-	-	-	-	-	-	-	-	W
Q/K	-	43.24%	-	-	13.51%	12.16%	14.86%	-	Q/K
H	-	-	-	-	-	-	-	-	H
R	-	-	-	-	-	-	-	-	R
D	-	-	-	-	-	-	-	-	D
E	-	-	-	-	-	-	-	-	E
N value	23.24	34.05	2.16	5.67	6.48	9.99	12.43	69.19	C value
N _{sum}	65.12				98.09				C _{sum}
					B*1503 N _{sum} /C _{sum} = 0.66				

TABLE 10

side chain	P1	P2	P3	P4	C ⁻³	C ⁻²	C ⁻¹	C-term	side chain
G	-	-	-	-	10.42%	-	-	-	G
P	-	27.08%	-	-	-	-	-	-	P
C	-	-	-	-	-	-	-	-	C
A	-	16.67%	13.54%	-	-	-	14.58%	-	A
V	-	-	-	-	-	11.46%	12.50%	-	V
F	-	-	-	-	-	-	-	23.96%	F
M	-	-	-	-	-	-	-	-	M
I/L	11.46%	-	-	-	-	-	-	-	I/L
S	-	-	-	-	-	11.46%	11.46%	-	S
T	10.42%	-	-	-	-	-	17.71%	-	T
Y	11.46%	-	-	-	-	-	-	66.67%	Y
N	-	-	-	-	-	-	-	-	N
W	-	-	-	-	-	-	-	-	W
Q/K	-	-	-	-	12.50%	-	-	-	Q/K
H	-	-	-	-	-	-	-	-	H
R	-	-	-	-	-	-	-	-	R
D	-	-	-	-	-	-	-	-	D
E	-	-	-	-	-	-	10.42%	-	E
N value	3.34	23.75	3.54	0.00	2.92	2.92	16.67	70.63	C value
N _{sum}	30.63				93.14				C _{sum}
B*1508 N _{sum} /C _{sum} = 0.33									

TABLE 11

side chain	P1	P2	P3	P4	C ⁻³	C ⁻²	C ⁻¹	C-term	side chain
G	10.57%	-	10.57%	10.57%	11.38%	-	-	-	G
P	-	-	-	12.20%	-	28.46%	-	-	P
C	-	-	-	-	-	-	-	-	C
A	-	-	19.51%	-	-	-	14.63%	-	A
V	-	-	-	-	-	-	22.76%	10.57%	V
F	-	-	-	-	-	-	-	-	F
M	11.38%	-	-	-	-	-	-	-	M
I/L	12.20%	-	-	-	-	-	12.20%	70.73%	I/L
S	11.38%	-	-	-	-	-	15.45%	-	S
T	-	-	-	-	-	-	-	-	T
Y	-	-	-	-	-	-	-	-	Y
N	-	-	-	-	-	-	-	-	N
W	-	-	-	-	-	-	-	-	W
Q/K	-	-	-	-	-	-	-	-	Q/K
H	-	58.54%	-	-	-	-	-	-	H
R	-	-	-	-	-	-	-	-	R
D	-	-	-	-	-	-	-	-	D
E	-	-	-	-	-	-	-	-	E
N value	5.53	48.54	10.08	2.77	1.38	18.46	25.04	61.3	C value
Nsum	66.92				106.18				Csum
B*1510 Nsum/Csum = 0.63									

TABLE 12

side chain	P1	P2	P3	P4	C ⁻³	C ⁻²	C ⁻¹	C-term	side chain
G	-	-	-	-	-	-	-	-	G
P	-	-	-	13.33%	-	-	-	-	P
C	-	-	-	-	-	-	-	-	C
A	-	16.67%	-	13.33%	-	-	-	-	A
V	-	-	-	-	-	-	16.67%	-	V
F	-	-	16.67%	-	-	-	-	16.67%	F
M	-	13.33%	-	-	-	-	-	-	M
I/L	-	13.33%	13.33%	16.67%	-	-	-	-	I/L
S	-	-	-	-	-	13.33%	-	-	S
T	-	-	-	-	-	13.33%	-	-	T
Y	13.33%	-	-	-	-	-	-	80.00%	Y
N	-	-	-	-	-	-	-	-	N
W	-	-	-	-	-	-	-	-	W
Q/K	-	16.67%	13.33%	-	-	20.00%	13.33%	-	Q/K
H	23.33%	-	-	-	-	-	-	-	H
R	-	-	-	-	-	-	-	-	R
D	-	-	-	13.33%	16.67%	-	-	-	D
E	-	-	-	-	-	-	16.67%	-	E
N value	16.66	20.00	13.33	16.66	6.67	16.66	16.67	76.67	C value
N _{sum}	66.65				116.67				C _{sum}
	B*1512 N _{sum} /C _{sum} = 0.57								

TABLE 13

fraction	ion for MS/MS	derived peptide sequence	alleles overlapping
6	398.2 (+3)	- - W D R H T X F	B*1501/B*1508
6	448.2 (+2)	- - - - - Y T	B*1501/B*1508
7	418.7 (+2)	A Q F A S G A G Z	B*1501/B*1503
8	402.2 (+2)	- G - - C D Y	B*1501/B*1503
8	418.7 (+2)	G S H F G V A Y	B*1501/B*1508
8	516.7 (+2)	N Q Z H G S A E Y	B*1501/B*1503/B*1508/B*1512
8	642.7 (+2)	P M N D W X M T Z T Y	B*1501/B*1512
9	331.4 (+3)	A P M A R G Z Y	B*1501/B*1503
9	418.7 (+2)	F V S N H A Y	B*1501/B*1508
9	433.2 (+2)	N P P A Z Z P N	B*1501/B*1503
9	437.0 (+2)	T G - - - A Y	B*1501/B*1508
9	441.2 (+2)	- Q - D P P P D M Z Y	B*1501/B*1503
9	446.6 (+2)	G Q Z Z A V D F	B*1501/B*1503
9/10	465.2 (+2)	S Q F G G G S Q Y	B*1501/B*1503/B*1508/B*1512
9	476.2 (+2)	S Q F D H V T Y	B*1501/B*1508
9	578.0 (+2)	T P X G E P Y Z S Y	B*1501/B*1503/B*1508
10	398.3 (+2)	X A N - - V T	B*1501/B*1508
10	456.8 (+2)	C P L S C F T	B*1501/B*1503/B*1508
10	509.0 (+2)	F L Z A M Z S T Y	B*1501/B*1508/B*1512
10	532.0 (+2)	T V X D S Z T H Y	B*1501/B*1508/B*1512
13	503.6 (+2)	G Q R K G A G S V F	B*1501/B*1503
14	460.7 (+2)	V V A P I T T G Y	B*1501/B*1508
14	475.1 (+2)	V V A C V - - - Y	B*1501/B*1508
14	525.3 (+2)	P L A - N - H T Y	B*1501/B*1508
15	514.2 (+2)	F Q A R X T E Y	B*1501/B*1508
16	522.0 (+2)	V G Y V D D T Q F	B*1501/B*1508
17	351.3 (+3)	A A F C G - - - X V	B*1501/B*1508
17	408.7 (+2)	Y L H - - E T	B*1501/B*1508
17/18	451.4 (+2)	I L G P P G S V Y	B*1501/B*1508/B*1512
17	462.4 (+2)	X L G D V N M Y	B*1501/B*1508
17	507.0 (+2)	- - - - X V E F	B*1501/B*1508
17	519.2 (+2)	T A R V X S V E Y	B*1501/B*1508
18	565.7 (+2)	A E F W A C Z X Y	B*1501/B*1503
18/19	566.2 (+2)	Y M I D P S G V S Y	B*1501/B*1508/B*1512
19/20	560.0 (+2)	X V E X T T D Y Y	B*1501/B*1512
20/21	448.2 (+2)	A A G X G P T F Y	B*1501/B*1512
20/21	614.0 (+2)	I A V G Y V D D T Q F	B*1501/B*1512
21/22	507.2 (+2)	V A F V X F V G Y	B*1501/B*1512
21/22	557.2 (+2)	Y N R W S X E F	B*1501/B*1512
22/23	510.8 (+2)	A L M P - - X N Y	B*1501/B*1512

TABLE 14

allele	ion overlaps collided	positive overlaps	overlap frequency
B*1512	20	14	70%
B*1508	286	25	9%
B*1503	88	12	14%
B*1510	26	0	0%

TABLE 15

motif P2/P9		+ length variation only		+ P2 variation only	
DLASMLNRY	(64-72)	MQLLCVF	(1-7)	DIEGHASHY	(28-36)
MLNRYKLIY	(68-76)	HLDIEGHASHY	(26-36)	SAPLEKQLF	(123-131)
PLEKQLFYY	(125-133)	MLSAPLEKQLF	(121-131)	APLEKQLFY	(124-132)
YQLRCHLSY	(149-157)	PLEKQLF	(125-131)	LPNTRPHSY	(138-146)
ALSINGDKF	(159-167)	PLEKQLFY	(125-132)	NTRPHSYVF	(140-148)
DLPLDLRGP	(203-211)	TMLPNTRPHSY	(136-146)	SINGDKFQY	(161-169)
FVPNLKDMF	(242-250)	MLPNTRPHSY	(137-146)	YTGAMTSKF	(169-177)
AVTMTAASY	(253-261)	QLRCHLSY	(150-157)	TSKFLMGTY	(174-182)
TMFEVSVAF	(290-298)	YVALSINGDKF	(157-167)	L TSAQSGDY	(216-224)
DLRWLAKSF	(314-322)	FQYTGAMTSKF	(167-177)	YSLVIVTTF	(224-232)
HLTTEKQY	(366-374)	AMTSKFLMGTY	(172-182)	VIVTTFVHY	(227-235)
ALRLATVGY	(375-383)	HVLSLVF	(192-198)	TTFVHYANF	(238-246)
ALGTESGLF	(467-475)	SLTSAQSGDY	(215-224)	MTAASYARY	(256-264)
AVSNAVDGF	(505-513)	SLVIVTTF	(225-232)	DTETLTTF	(284-292)
ALYEASTTY	(564-572)	LVIVTTF	(226-232)	ATVKGMQSY	(338-346)
RQIPKIQNF	(597-605)	IVTTFVHY	(228-235)	ATSVLLSAY	(396-404)
ILSSNYFDF	(643-651)	IVTTFVHYANF	(228-238)	SAYNRHPLF	(402-410)
TVMEIAGLY	(666-674)	FVHYANFHF	(232-241)	HTVMRETLE	(414-422)
HVVLAILLY	(679-687)	FVHYANFHFY	(232-242)	ESGLFSPCY	(471-479)
VVLAILLYF	(680-688)	TMTAASY	(255-261)	SPCYLSLRF	(476-484)
FLVHKIVMF	(696-704)	TMTAASYARY	(255-264)	IIPLINVTF	(544-552)
LVHKIVMFF	(697-705)	ELDTETLTTF	(282-292)	TTYLSSSLF	(570-578)
		TMFEVSVAFF	(290-399)	NSILSSNYF	(641-649)
		TVLKDIIGICY	(326-326)	AIILYFIAF	(683-691)
		VLKDIIGICY	(327-326)	FIAFALGIF	(688-696)
		TVKGMQSY	(339-346)		
		RLATVGY	(377-383)		
		TVGYPKAGVY	(380-389)		
		LLSAYNRHPLF	(400-410)		
		PLHTVMRETLE	(412-422)		
		VMRETLE	(416-422)		
		GLALGTESGLF	(465-475)		
		GLFSPCY	(473-479)		
		LMIIPLINVTF	(542-552)		
		PLINVTF	(546-552)		
		EVRGSALY	(559-566)		
		YLSSSLF	(572-578)		
		TQKSCIF	(608-614)		
		TQKSCIFCGF	(608-617)		
		GLETTTY	(627-633)		
		VQNSILSSNY	(639-648)		
		VQNSILSSNYF	(639-649)		
		ILSSNYF	(643-649)		
		VMEIAGLY	(667-674)		
		VVLAILLY	(680-687)		
		VVLAILLYF	(680-688)		
		VLAILLY	(681-687)		
		VLAILLYF	(681-688)		
		VLAILLYFIAF	(681-691)		
		ILYFIAF	(685-691)		
		FLVHKIVMFF	(696-705)		

TABLE 16

Primer name	Sequence 5'-3'	Locus	Cut site	Annealing site
PP5UTA	GCGCTCTAGACCCAGACGCCGAGGATGGCC	A	XbaI	5UT
3PI4A	GCCCTGACCCCTGCTAAAGGT	A		Intron 4
PP5UTB	GCGCTCTAGACCCACCCGGACTCAGAATCTCCT	B	XbaI	5UT
3PI4B	TGCTTTCCTGAGAAAGAT	B		Intron 4
5UTB39	AGCGGAATTCAGAGTCTCCTCAGACGCG	B*39	EcoRI	5UT B39
5PKCE	GGCGGAATTCCCGCCGCCACCATGCGGGTCATGGCGCC	C	EcoRI	5UT
3PI4C	TTCTGCTTTCCTGAGAAAGAC	C		Intron 4
PP5UT	GGCGGAATTCGGACTCAGAATCTCCCCAGACGCCGAG	B	EcoRI	5UT
PP3PEI	CCGCGAATTCTCATCTCAGGGTGAGGGGCT	A,B,C	EcoRI	Exon 4
PP3PEIH	CCGCAAGCTTTCATCTCAGGGTGAGGGGCT	A,B,C	HindIII	Exon 4
3PEIHC7	CCGCAAGCTTTCAGCTCAGGGTGAGGGGCT	Cw*07	HindIII	Exon 4

TABLE 17

Primer Name	Sequence 5'-3'
T7Prom	TAATACGACTCACTATAGGG
BGHrev	TAGAAGGCACAGTCGAGG
PPI2E2R	GTCGTGACCTGCGCCCC
PPI2E2F	TTTCATTTTCAGTTTAGGCCA
ABC13E4F	GGTGTCTCTGTCCATTCTCA

5'CY5 Sequencing Primers

TABLE 18

Sample	OD 260nm	OD 280nm	260nm/280nm	Dilution factor	Concentration ug/ml
3A394	0.0346	0.0202	1.7111	20	34.5821

TABLE 19

Sample	OD 260nm	OD 280nm	260nm/280nm	Dilution factor	Concentration ug/ml
3A394TPC 1	0.2821	0.1505	1.8739	20	282.0960

TABLE 20

Sample	OD 260nm	OD 280nm	260nm/280nm	Dilution factor	Concentration ug/ml
3A394TPC 1	0.6919	0.3625	1.9087	50	1729.8492

TABLE 21

Sample	Decay time milliseconds	# live cells/ml	# dead cells/ml	Viability %
3A394TPC 1	19.8	1.12×10^6	1.65×10^5	87.16

TABLE 22

Sample	Optical Density 492nm	Dilution	Concentration of soluble HLA ng/ml
3A394TP C1 well 1	1.278 1.388 (over range)	1.0	247.270
3A394TP C1 well 2	1.227 1.274	1.0	229.855
3A394TP C1 well 3	1.021 1.042	1.0	154.403
3A394TP C1 well 4	1.108 1.070	1.0	169.001

TABLE 23

Allele	Allele Allele AlleleConcentration by ELISA ug/ml	Concentration by ELISA ug/ml Concentration by ELISA ug/ml Concentration by ELISA ug/mlTotal amount made mg
Total amount made mg Total amount made mg Total amount made mgHLA- A*0301	545.4	3.47
HLA-A*1102	888.5	2.57
HLA-A*2902	476.8	2.58
HLA-A*3002	50.3	3.38
HLA-A*3201	1382.0	9.61
HLA-A*3301	40.0	0.8
HLA-B*0801	66.0	21.0
HLA-B*1302	55.0	9.0
HLA-B*1401	146.0	50.0
HLA-B*1801	587.6	0.4
HLA-B*3701	1831.0	119.0
HLA-B*3801	128.0	66.0
HLA-B*3905	1400.0	120.0
HLA-B*40012	59.0	10.0
HLA-B*4002	400.0	180.0
HLA-B*4101	288.4	8.8
HLA-B*4402	59.0	10.0

TABLE 24

fraction	ion for MS/MS	derived peptide sequence
6	398.2 (+3)	- - W D R H T X F
6	448.2 (+2)	- - - - - Y T
7	382.7 (+2)	V Q F E A A T
7	418.7 (+2)	A Q F A S G A G Z
7	455.2 (+2)	A L G A - - R G Y
7	489.1 (+2)	- - V - - G H X Y
7	506.8 (+2)	X S - - - C E Y
8	402.2 (+2)	- G - - C D Y
8	419.2 (+2)	G S H F G V A Y
8	433.8 (+2)	A P P P P P P P P
8	455.2 (+2)	- - - Z A R G Y
8	462.2 (+2)	D P <u>H</u> <u>A</u> <u>P</u> P Z Y
8	507.2 (+2)	A V P S X H X X Y
8	512.3 (+2)	X A Z V Z M T A Y
8	512.8 (+2)	A L N G <u>R</u> <u>V</u> <u>T</u> M Y
8	516.9 (+2)	N Q Z H G S A E Y
8	522.9 (+2)	F G <u>X</u> <u>A</u> <u>C</u> X A T S Y
8	642.7 (+2)	P M N D W X M T Z T Y
9	331.4 (+3)	A P M A R G Z Y
9	418.7 (+2)	F V S N H A Y
9	426.2 (+3)	- - - - - - - - S Y
9	433.3 (+2)	N P P A Z Z P N
9	437.0 (+2)	T G - - - - A Y
9	441.2 (+3)	- Q - D P P P D M Z Y
9	446.6 (+2)	G Q Z Z A V D F
9	453.6 (+2)	X Q - - A G G Z Y
9	465.2 (+2)	S Q F G G G S Q Y
9	476.2 (+2)	S Q F D H V T Y
9	481.0 (+2)	G Q H A S V X S Y
9	514.2 (+2)	- - A A H V P P G Y
9	550.2 (+2)	F M D V G A P T V Y
9	578.0 (+2)	T P X G E P Y Z S Y
10	398.3 (+2)	X A N - - V T
10	448.2 (+2)	A Q A A P F A G Y
10	448.4 (+2)	V V V F G V Z F
10	450.4 (+2)	A Q M - - S E Y
10	456.8 (+2)	C P L S C F T
10	464.7 (+2)	- - - - F G H Y
10	473.7 (+2)	<u>A</u> <u>L</u> W - - P Z F
10	486.4 (+2)	V P H Z N A Y
10	498.7 (+2)	- - - - - G H G G Y
10	509.0 (+2)	F L Z A M Z S T Y
10	527.7 (+2)	G Q Y V <u>V</u> <u>Z</u> P T Y
10	532.0 (+2)	T V X D S Z T H Y
10	540.2 (+2)	P M F D P P Z T F
11	469.2 (+2)	A Q A E S L R Y
11	480.6 (+2)	X A V G H S G G T Y
11	511.2 (+2)	- - - - - P T Y
11	516.7 (+2)	E S X P N N V P Y
12	383.0 (+3)	L <u>A</u> <u>H</u> <u>T</u> <u>E</u> <u>C</u> P R G Y
12	435.0 (+2)	- - - - - P S Y
12	473.2 (+2)	<u>V</u> <u>Q</u> <u>G</u> P V G V Q Y

TABLE A

12	475.0 (+2)	R G X G V A G T A F
12	505.0 (+2)	T G A P V S E E G Y
12	513.7 (+2)	V Q X Y Y G <u>S</u> <u>V</u> V
12	519.0 (+2)	E P A M V X Z C F
12	531.2 (+2)	G Q P G A P X G G Z Y
12	541.0 (+2)	G P <u>P</u> <u>H</u> <u>N</u> G X R A Y
12	542.2 (+2)	A A H W H V E A Y
12	553.7 (+2)	T P P T R R E S Y
12	577.2 (+2)	F P T D R R S Q F
13	363.0 (+3)	Y T G V S Y X H F
13	447.0 (+2)	A Q A S A P D A Y
13	465.0 (+2)	V Q Y Y X P F
13	503.6 (+2)	G Q R K G A G S V F
13	553.2 (+2)	X Q Z X - - D V Y
13	590.8 (+2)	A T <u>G</u> <u>T</u> <u>A</u> <u>Z</u> N X N Z Y
14	460.7 (+2)	V V A P I T T G Y
14	471.5 (+2)	V V A C V - - - Y
14	495.2 (+2)	X Q Y T V G Y F
14	525.3 (+2)	P L A - N - H T Y
14	541.3 (+2)	P L F G Q T A G Q Y
14	550.4 (+2)	A - - - - Q X E Y
14	577.2 (+2)	Z G Y G N P X N G A Y
15	459.8 (+2)	V Q G P V G T D F
15	470.9 (+2)	V A G G W - - - F
15	514.2 (+2)	F Q A R X T E Y
15	536.6 (+2)	X A G F F X X E Y
15	544.2 (+2)	X Q - - - - Z Y
15	564.2 (+2)	S G A X D R A Y Z F
16	467.1 (+2)	F Q - - - - T X
16	500.4 (+2)	T P - - - - A Z A F
16	501.0 (+2)	V V A T Z N Z Z X
16	503.6 (+2)	Y M V T - - - F
16	517.4 (+2)	A L G S Z A X M P F
16	521.3 (+2)	A P A V - - - V G Y
16	522.0 (+2)	V G Y V D D T Q F
16	525.6 (+2)	- - - - - T G F
16	536.0 (+2)	P V P N V R X N Y
16	544.4 (+2)	- - - - - T X S X
16	557.6 (+2)	T L E G W M S Z Y
16	561.5 (+2)	Y M V C N A E E Y
16	596.7 (+2)	- - - - - X R D X Y
16	596.9 (+2)	S L X - - - - - F
17	343.2 (+3)	A Q H P <u>S</u> <u>A</u> <u>X</u> <u>R</u> F
17	351.3 (+3)	A A F C G - - - X V
17	408.7 (+2)	Y L H - - E T
17	441.2 (+2)	- - - - - Z A Y
17	451.4 (+2)	I L G P P G S V Y
17	455.0 (+2)	G L G Z T S A E F
17	462.4 (+2)	X L G D V N M Y
17	483.8 (+2)	V M <u>G</u> <u>X</u> <u>T</u> <u>N</u> A N F
17	490.2 (+2)	N A X G R E S S F
17	497.2 (+2)	A M N P T N T V F
17	507.0 (+2)	- - - - X V E F
17	511.2 (+2)	X Q A P A X F V Y
17	519.2 (+2)	T A R V X S V E Y

TABLE A cont'd.

17	526.8 (+2)	A L F - - - F T Y
17	542.8 (+2)	X Q X N A Y X S Y
17	563.2 (+2)	G L A R <u>C</u> <u>S</u> Z V E Y
18	503.8 (+2)	S Q X <u>A</u> <u>A</u> <u>G</u> V D V F
18	511.7 (+2)	P Q G Z M A - - Y
18	519.6 (+2)	- V F V S H T T F
18	538.8 (+2)	<u>H</u> X T G <u>N</u> E A T S F
18	565.7 (+2)	A E <u>F</u> W A C Z X Y
18	566.2 (+2)	Y M <u>I</u> D P S G V S Y
18	581.2 (+2)	X Q G H H E M F Y
20	448.2 (+2)	A A G X G P T F Y
20	560.0 (+2)	X V E X T T D Y Y
20	614.0 (+2)	I A V G Y V D D T Q F
21	507.2 (+2)	V A <u>F</u> <u>V</u> <u>X</u> F V G Y
22	510.8 (+2)	<u>A</u> <u>L</u> M P - - X N Y
22	557.2 (+2)	Y <u>N</u> <u>R</u> <u>W</u> S X E F
24	546.3 (+2)	- - Z D R N V T F
25	546.3 (+2)	V V T M - - - Z Y

* Dashes represent positions at which amino acids could not be unambiguously assigned through NanoES-MS/MS fragmentation patterns and/or Edman data obtained. Underlined residues designate tentative assignments.

TABLE A cont'd.

fraction	ion for MS/MS	derived peptide sequence
6	471.8 (+2)	A Z V E C E T Y
7	418.7 (+2)	A Q F A S G A G Z
7	504.2 (+2)	Z G X G G G P A T S Y
8	402.2 (+2)	- G - - C D Y
8	441.2 (+2)	- - - - - Z S F
8	516.9 (+2)	N Q Z H G S A E Y
9	331.4 (+3)	A P M A R G Z Y
9	349.4 (+3)	- - - - - G F Y
9	418.7 (+2)	A Z V N S G - Y
9	426.2 (+3)	A A S S Z V - - P P Z Y
9	433.3 (+2)	N P P A Z Z P N
9	437.0 (+2)	A C G G C G Z D Y
9	441.2 (+3)	- Z - D P P P D M Z Y
9	446.6 (+2)	G Q Z Z A V D F
9	578.0 (+2)	T P X G E P Y Z S Y
10	426.5 (+2)	G P - - - P Z Y
10	443.2 (+2)	A P Z Y P P P P
10	448.3 (+2)	G Z V C T P G S F
10	456.8 (+2)	C P L S C F T
10	464.7 (+2)	S Q F G G G S Q Y
10	465.4 (+2)	A S G F N G S Z Y
10	503.8 (+2)	- Z - - Y T A Y
10	508.7 (+2)	G Z P P H N G F Y
10	517.0 (+2)	I K A D H V S T Y
10	527.7 (+2)	X Z A D H V X P Y
10	540.2 (+2)	- - - - P G Z V Y
10	549.2 (+2)	Z S V - - - Z T G Y
11	437.0 (+2)	H X G N Q A A Y
11	511.4 (+2)	Z A G T T V P V S Y
11	527.4 (+2)	G Q Y P T Q P T Y
11	581.4 (+2)	F A G S Z S N T S T Y
12	494.8 (+2)	S Z G G - - - T G Y
12	526.8 (+2)	Z G P P N Y X T Y
12	547.1 (+2)	V K V I Q Q E S Y
13	454.6 (+2)	L P P P P P P P P
13	476.0 (+2)	A K Y S T P A T L
13	503.6 (+2)	G Q R K G A G S V F
13	513.1 (+2)	R Z S A N H E A X
13	526.4 (+2)	G K V R T D I T Y
13	553.2 (+2)	V V X P A V R S T Y
13	561.0 (+2)	A K Y P H V E D Y
13	571.3 (+2)	A Z N X S A Y V X Y
13	601.2 (+2)	E V V G D T Z Y
14	438.2 (+2)	A K A G I T T T L
14	490.8 (+2)	V - - T Z A G S A F
14	517.2 (+2)	A Z A A A N V X X Y
14	531.5 (+2)	A N H S V R D T Y
14	535.3 (+2)	E - - - G X R Z Y
14	552.8 (+2)	X Z H N D Z S T Y
14	577.2 (+2)	A N E Z X G - - - Y

TABLE B

15	497.3 (+2)	A A G P T A Z E S Y
15	514.2 (+2)	V A G X V F M Z Y
15	527.0 (+2)	A Z Y Z A Z V V F
15	564.2 (+2)	A Z F - - - Z X Y
15	577.2 (+2)	Z G Y G N P X N Z Y
15	626.0 (+2)	- - - - - Z A P C H Y
16	521.6 (+2)	A H A V Q R V V Y
16	525.6 (+2)	T Z X T V V X N Y
17	446.8 (+2)	A Z Z A S G X A F
17	492.8 (+2)	G S H S M R Y F
17	503.8 (+2)	Y G Y G A T V E F
17	967.6 (+1)	V Z - - - T T F
18	451.4 (+2)	Q P G P Q I V Y
18	455.2 (+2)	N G Z X S N N Y
18	475.2 (+2)	A N X V Z X E Y
18	489.1 (+2)	G Z - - - Z G X X Y
18	497.8 (+2)	A M N P T N T V F
18	525.2 (+2)	Y N - - - Z X F
18	538.8 (+2)	- M - - S Y Z N F
18	565.7 (+2)	A E <u>F</u> <u>W</u> A C Z X Y
19	521.6 (+2)	S Z <u>F</u> <u>G</u> <u>C</u> <u>P</u> T R F
19	524.6 (+2)	X G A X S N - - E F
19	571.2 (+2)	R Z A A Y R X T Y
19	646.2 (+2)	T N X H D G D G A T Z Y

* Dashes represent positions at which amino acids could not be unambiguously assigned through NanoES-MS/MS fragmentation patterns and/or Edman data obtained. Underlined residues designate tentative assignments.

TABLE B cont'd

fraction	ion for MS/MS	derived peptide sequence
6	398.2 (+3)	- - W D R H T X F
6	448.2 (+2)	- - - - - Y T
8	419.2 (+2)	G S H F G V A Y
8	441.2 (+2)	V P <u>C</u> <u>G</u> <u>Z</u> <u>Z</u> S Y
8	473.2 (+2)	T A Z X H R G Y
8	512.8 (+2)	X A Z Y E H T Y
8	516.9 (+2)	N Q Z H G S A E Y
8	546.8 (+2)	N G <u>X</u> <u>A</u> <u>M</u> <u>H</u> W T Y
9	418.7 (+2)	F V S N H A Y
9	437.0 (+2)	T G - - - A Y
9	465.2 (+2)	S Q F G G G S Q Y
9	476.2 (+2)	S Q F D H V T Y
9	481.0 (+2)	- P - - G Z D E V
9	514.2 (+2)	N G Y D G P N A G Y
9	578.0 (+2)	T P X G E P Y Z S Y
10	398.3 (+2)	X A N - - V T
10	448.3 (+2)	M P H S <u>G</u> Y G F
10	450.4 (+2)	V D X - - - Y
10	456.8 (+2)	C P L S C F T
10	464.7 (+2)	- - - - - P G F Y
10	486.2 (+2)	- A - P H P M G Y
10	494.2 (+2)	A Q T V G Y G E Y
10	508.7 (+2)	- - - - - S V Y
10	509.0 (+2)	F L Z A M Z S T Y
10	532.0 (+2)	T V X D S Z T H Y
11	444.1 (+2)	T P - - A R A P T
11	469.2 (+2)	S E H D R M Y
11	480.6 (+2)	T G N C S G T G T Y
11	496.8 (+2)	A Q V N P S X T Y
11	532.3 (+2)	S P G A E T R A X Y
12	473.2 (+2)	Y L G - - - G A F
12	494.8 (+2)	X T S F M Z V Y
12	499.0 (+2)	- P - - - P S S G Y
12	505.0 (+2)	T P - - - G R M Y
12	513.7 (+2)	P M F D Z Z V Y
12	519.0 (+2)	Y L - - - R T F
12	531.2 (+2)	A Q E H G C A A Z F
12	542.2 (+2)	- <u>M</u> - - - <u>G</u> <u>V</u> H D Y
12	550.2 (+2)	Y V S - - R N Q Y
12	553.7 (+2)	A Q Y A A G E S F Y
12	564.0 (+2)	<u>T</u> <u>P</u> H T Z H D E Y
12	565.2 (+2)	Y M - - - <u>F</u> M Y
13	396.1 (+3)	D P H Y V S G H Z F
13	401.2 (+2)	M V G X X P A T
13	526.4 (+2)	Z A S P G E X T S Y
14	460.7 (+2)	V V A P I T T G Y
14	471.5 (+2)	V V A C V - - - Y
14	525.3 (+2)	P L A - N - H T Y
14	543.2 (+2)	X A <u>X</u> <u>Y</u> <u>R</u> <u>R</u> M Y
14	550.4 (+2)	P L A <u>M</u> <u>Z</u> X Y T Y

TABLE C

15	460.6 (+2)	- P - M P G X A Y
15	461.0 (+2)	H T T S Z N A Y
15	506.0 (+2)	M A A M V G V A V Y
15	508.4 (+2)	G P Z V M Z H G Y
15	514.2 (+2)	F Q A R X T E Y
15	520.0 (+2)	L P H Q P L A T Y
15	525.2 (+2)	A A A X V - - - V T Y
15	536.6 (+2)	X P E M G Z F S Y
15	544.2 (+2)	Y V - - V R - V F
15	564.2 (+2)	F V T X N X E E Y
16	489.0 (+2)	A A P V G A X E S Y
16	500.4 (+2)	G S - - - S Y T Y
16	522.0 (+2)	V G Y V D D T Q F
16	525.7 (+2)	Y V A - - - P A F
16	533.0 (+2)	V G Y - - A H P G F
16	535.7 (+2)	Z A T N S V T S T Y
16	537.0 (+2)	- - - - - S T Y
16	545.8 (+2)	Y A T A G E M M A F
16	547.0 (+2)	S P T Y T H A V A F
16	557.0 (+2)	M P A - - M V M A F
17	351.3 (+3)	A A F C G - - - X V
17	393.7 (+2)	S P N E D X M Z V F
17	403.2 (+2)	V A A T A G A V F
17	408.7 (+2)	Y L H - - E T
17	414.8 (+2)	T A F P F V F
17	451.4 (+2)	I L G P P G S V Y
17	462.4 (+2)	X L G D V N M Y
17	476.2 (+2)	Y G - - - V X S M
17	490.8 (+2)	X P H C S C S S F
17	504.0 (+2)	D P <u>P</u> <u>C</u> <u>W</u> <u>G</u> V S F
17	507.0 (+2)	- - - - X V E F
17	511.2 (+2)	- - - - A H D A Y
17	519.2 (+2)	T A R V X S V E Y
17	526.8 (+2)	X S D G R Z X T Y
17	542.8 (+2)	N M N D L V S E Y
17	557.2 (+2)	M P A A <u>D</u> <u>Y</u> <u>E</u> V A F
18	474.8 (+2)	A E I L Q V I Y
18	503.8 (+2)	A P - - - X V S Y
18	514.7 (+2)	M P A G Y N N V Y
18	519.6 (+2)	Y M S G <u>X</u> <u>Y</u> <u>G</u> T F
18	526.8 (+2)	- - - <u>A</u> <u>V</u> <u>V</u> <u>A</u> <u>Z</u> <u>S</u> <u>Y</u>
18	538.8 (+2)	X P V V P A A Z T Y
18	566.2 (+2)	Y M I D P S G V S Y
18	616.3 (+2)	F A N G V <u>Z</u> <u>G</u> <u>C</u> <u>A</u> <u>F</u> A F

* Dashes represent positions at which amino acids could not be unambiguously assigned through NanoES-MS/MS fragmentation patterns and/or Edman data obtained. Underlined residues designate tentative assignments.

TABLE C cont'd.

fraction	ion for MS/MS	derived peptide sequence
6	493.0 (+2)	N H A V G - - V S M
6	557.8 (+2)	H N V F Z P T S N A
7	434.8 (+2)	S V C E T E S X
7	481.3 (+2)	T H P S Z A C A F
7	489.1 (+2)	- H - - S P X X
8	420.1 (+2)	A N X E G P H T
8	441.7 (+2)	G H S P P T S S L
8	494.8 (+2)	C H S A F A L
8	511.6 (+2)	H H A F A Z V X V
8	519.4 (+2)	D H Y Y X A G S X
9	411.4 (+2)	E X A P H A A X
9	424.3 (+2)	A A A X R C E X
9	426.1 (+2)	G H Z A P A A S X
9	441.7 (+2)	V H N P Z S S X
9	444.2 (+2)	A G G P T X C R X
9	455.5 (+2)	L H L L T L E A
9	490.3 (+2)	A G G Z P A T P P A X
9	513.1 (+2)	S H Z G C V Z P A V
9	520.0 (+2)	X H R L C S P T X
10	404.2 (+2)	S V <u>S</u> <u>X</u> P H A P
10	417.1 (+2)	A P F T G G N G X
10	433.8 (+2)	G H D P D S P A A
10	446.2 (+2)	E H G X E N G H
10	455.4 (+2)	E H V A S S P A L
10	460.4 (+2)	H H A P C G V S X
10	464.0 (+2)	N H A I V S T S V
10	464.7 (+2)	G H Z N S V T S V
10	465.3 (+2)	S H Z A P C T <u>S</u> V
10	469.4 (+2)	F V A R F V S X
10	469.6 (+2)	H H S D G S V S L
10	473.7 (+2)	S H A G A P P P T X
10	482.6 (+2)	M C Z - G M P A X
10	482.8 (+2)	G H G A N N D P A X
10	495.7 (+2)	X H S Z P A G P A X
10	508.3 (+2)	X H V V S - - V X
10	511.2 (+2)	A V X D C C Z V A V
10	522.3 (+2)	E X G G N T N P Z X
10	522.7 (+2)	Y H G S Z N P E X
10	569.6 (+2)	- - - - - T Y S Y
10	574.3 (+2)	- - - - - - - M
11	405.7 (+2)	S H - - - Y F
11	425.8 (+2)	A H P D Z A X V
11	444.7 (+2)	G T A H Y Z V X
11	448.9 (+2)	M H A D N P V X
11	455.7 (+2)	S H <u>V</u> <u>D</u> <u>R</u> P S X
11	459.7 (+2)	T G A A F Z N P X
11	482.8 (+2)	G H <u>C</u> <u>P</u> R N P A X
11	495.7 (+2)	X H S G A P Z A P X
11	516.7 (+2)	X H D <u>T</u> <u>E</u> <u>H</u> <u>A</u> P X
11	562.3 (+2)	- - - Y Z A Y V Y

TABLE D

12	411.7 (+2)	G H G P T X A A V
12	428.8 (+2)	V P - - - - -
12	444.7 (+2)	Y Q H T G A V L
12	448.4 (+2)	T Q A P G N P V L
12	460.3 (+2)	T Z A <u>G</u> <u>C</u> <u>M</u> <u>V</u> P X
12	490.9 (+2)	T H T Q P G V Q L
12	507.4 (+2)	G H A G H V P P E X
12	511.6 (+2)	T H F R Y V S X
12	528.1 (+2)	E H R P D R V F
13	427.6 (+2)	S H A Q T V V L
13	449.2 (+2)	S H A N S A V V L
13	464.8 (+2)	M V - - H P V X
13	487.6 (+2)	Y H H G G V S A F
13	506.2 (+2)	- H - - G H T G Y X
14	420.1 (+2)	N H A N G L T L
14	438.7 (+2)	- - - - - P X X
14	456.7 (+2)	A H S V P S P A F
14	477.7 (+2)	M H T - - P A P V
14	482.8 (+2)	P G A A V V P S X
14	560.8 (+2)	G H A G M G C V F Z X
14	592.3 (+2)	M R - - - - G X E X
15	418.9 (+2)	S H G V P R A X
15	439.0 (+2)	E H H M P X X
15	454.3 (+2)	H H Z C A A G A X
15	492.1 (+2)	X V D <u>Z</u> A <u>E</u> P X V
15	510.1 (+2)	I H T P E N P V I
15	520.0 (+2)	M G X P V R H M V
15	524.2 (+2)	S H Y D W Z V X
15	532.9 (+2)	M P H S H P F V X
15	577.2 (+2)	Z C V R C Z N G V F
16	412.9 (+2)	S H A G A G X V X
16	418.3 (+2)	G H X E G P X X
16	424.3 (+2)	X H G G D H V X
16	448.6 (+2)	E Z A H S X V X
16	448.9 (+2)	Y H H D X V X
16	454.3 (+2)	M A G A W C R X
16	456.7 (+2)	S H <u>D</u> <u>G</u> <u>S</u> V P T X
16	464.2 (+2)	F H - - X X X
16	469.9 (+2)	E H - - - T V X
16	472.3 (+2)	M A X - - - V V
16	499.0 (+2)	G H A <u>X</u> <u>T</u> D G X T X
16	504.1 (+2)	P V <u>S</u> <u>H</u> <u>X</u> V N E L
16	507.7 (+2)	X X Y <u>T</u> <u>P</u> <u>G</u> H T X
16	522.7 (+2)	- - - - - P V X
16	523.3 (+2)	M A H S - - P V F
16	529.9 (+2)	X H Y D R N Q X
16	536.2 (+2)	E A - - C Z V T T Y
16	547.9 (+2)	- - - - - A X S V
16	552.4 (+2)	X Z <u>A</u> <u>P</u> <u>T</u> S V F Z X
17	367.7 (+3)	<u>F</u> <u>T</u> <u>M</u> <u>P</u> A H P S T X
17	490.8 (+2)	M T X G Y G E P X
17	557.3 (+2)	A H G R K M S K S L
17	340.7 (+3)	- <u>H</u> - - H A Z V X
17	367.7 (+3)	- - - - R X S H X
17	419.8 (+2)	- - - H A V G X X

TABLE D cont'd.

17	462.8 (+2)	M S S N E X X M
17	476.2 (+2)	G H - - - P C C
17	504.2 (+2)	X H V X A V N E X
17	523.2 (+2)	- H - - - - P V F
17	543.2 (+2)	X H E V Z P H X X
17	590.2 (+2)	A T E H <u>C</u> <u>F</u> <u>V</u> M E X
18	456.4 (+2)	A H S N L A S V L
18	463.3 (+2)	V X A P A N D X X
18	474.8 (+2)	M X <u>G</u> <u>X</u> S F P A X
18	491.2 (+2)	V H T C V N P V X
18	497.8 (+2)	S H Q R Q L L L
18	515.8 (+2)	E W <u>H</u> <u>Y</u> P V S X
18	519.7 (+2)	<u>F</u> <u>H</u> <u>M</u> <u>D</u> X Z T F
18	543.4 (+2)	X H E V <u>Z</u> P H X X
18	596.8 (+2)	<u>F</u> <u>H</u> <u>H</u> <u>T</u> Z S N P X X
19	434.6 (+2)	- H <u>G</u> <u>C</u> <u>P</u> G M P X
19	496.6 (+2)	<u>E</u> <u>T</u> <u>P</u> <u>E</u> H A P V X
19	539.6 (+2)	<u>M</u> <u>X</u> P G N S A X Y X

* Dashes represent positions at which amino acids could not be unambiguously assigned through NanoES-MS/MS fragmentation patterns and/or Edman data obtained. Underlined residues designate tentative assignments.

TABLE D cont'd.

fraction	ion for MS/MS	derived peptide sequence
7	504.1 (+2)	H M S G Z P T S Y
7	549.2 (+2)	H N Z A A H Z E Y
8	517.0 (+2)	N Q Z H G S A E Y
8	526.0 (+2)	H A A <u>X</u> <u>Y</u> <u>S</u> Z V Y
8	642.7 (+2)	P M N D W X M T Z T Y
10	465.3 (+2)	S Q F G G G S Q Y
10	484.3 (+2)	Y Q S <u>D</u> <u>H</u> R Y
10	509.0 (+2)	F L Z A M Z S T Y
10	532.0 (+2)	T V X D S Z T H Y
11	424.3 (+2)	H X S T Z D F
11	464.3 (+2)	H A P P T D P P P
11	550.0 (+2)	H G P A N <u>R</u> <u>D</u> <u>S</u> V F
11	563.3 (+2)	F P Y P T D P Z Y
12	531.2 (+2)	K N A N L V Q L Y
14	585.6 (+2)	R S F X X E N E Y
16	488.7 (+2)	H M Z N P T S Y
16	661.9 (+2)	Y V X F - - - V Y
17	577.6 (+2)	R S M X <u>R</u> <u>C</u> <u>P</u> E Y
18	451.1 (+2)	I L G P P G S V Y
18	523.0 (+2)	- - <u>F</u> <u>V</u> <u>T</u> A Z T Y
19	565.6 (+2)	Y M I D P S G V S Y
19	503.8 (+2)	S Q X A A G V D V F
20	560.0 (+2)	X V E X T T D Y Y
20	582.4 (+2)	M Y N <u>C</u> <u>N</u> E X D Y
21	448.2 (+2)	A A G X G P T F Y
21	614.0 (+2)	I A V G Y V D D T Q F
22	507.2 (+2)	V A <u>F</u> <u>V</u> <u>X</u> F V G Y
22	557.2 (+2)	Y N <u>R</u> <u>W</u> S X E F
23	510.8 (+2)	<u>A</u> <u>L</u> M P - - X N Y
25	562.8 (+2)	N Q F Q A L L Q Y

* Dashes represent positions at which amino acids could not be unambiguously assigned through NanoES-MS/MS fragmentation patterns and/or Edman data obtained. Underlined residues designate tentative assignments.

TABLE E

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What I claim is:

1. A method for the production of soluble HLA molecules in a cell pharm having an appropriate growth media therein, comprising the steps of:
 - isolating HLA allele mRNA from a source and reverse transcribing the mRNA to obtain allelic cDNA;
 - amplifying the allelic cDNA by PCR, wherein the amplification utilizes at least one locus-specific primer having a stop codon incorporated into a 3' primer thereby resulting in a truncated PCR product having the coding regions encoding cytoplasmic and transmembrane domains of the allelic cDNA removed such that the truncated PCR product has a coding region encoding a soluble HLA molecule;
 - inserting the truncated PCR product into a mammalian expression vector to form a plasmid containing the truncated PCR product having the coding region encoding a soluble HLA molecule;
 - electroporating the plasmid containing the truncated PCR product into at least one suitable host cell; and
 - inoculating the cell pharm with the at least one suitable host cell containing the plasmid containing the truncated PCR product such that the cell pharm produces soluble HLA molecules.

2. The method according to claim 1, further comprising the step of harvesting the soluble HLA molecules from the cell pharm.
3. The method according to claim 1 wherein the soluble HLA molecules are Class I HLA molecules or Class II HLA molecules.
4. The method according to claim 1 wherein, in the step of isolating HLA allele mRNA from a source, the source is selected from the group consisting of mammalian DNA and an immortalized cell line.
5. The method according to claim 1 wherein, in the step of inserting the truncated PCR product into a mammalian expression vector, the mammalian expression vector contains a promoter that facilitates increased expression of the truncated PCR product.
6. The method according to claim 1 wherein, in the step of electroporating the plasmid containing the truncated PCR product into at least one suitable host cell, the suitable host cell lacks expression of Class I HLA molecules.
7. A method for the production of soluble HLA molecules in a cell pharm

having an appropriate growth media therein, comprising the steps of:

- isolating HLA allele mRNA from a source and reverse transcribing the mRNA to obtain allelic cDNA;
- amplifying the allelic cDNA by PCR, wherein the amplification utilizes at least one locus-specific primer that truncates the allelic cDNA, thereby resulting in a truncated PCR product having the coding regions encoding cytoplasmic and transmembrane domains of the allelic cDNA removed such that the truncated PCR product has a coding region encoding a soluble HLA molecule;
- inserting the truncated PCR product into a mammalian expression vector to form a plasmid containing the truncated PCR product having the coding region encoding a soluble HLA molecule;
- electroporating the plasmid containing the truncated PCR product into at least one suitable host cell; and
- inoculating the cell pharm with the at least one suitable host cell containing the plasmid containing the truncated PCR product such that the cell pharm produces soluble HLA molecules.

8. The method according to claim 7, further comprising the step of harvesting the soluble HLA molecules from the cell pharm.

9. The method according to claim 7 wherein the soluble HLA molecules are Class I HLA molecules or Class II HLA molecules.
10. The method according to claim 7 wherein, in the step of isolating HLA allele mRNA from a source, the source is selected from the group consisting of mammalian DNA and an immortalized cell line.
11. The method according to claim 7 wherein, in the step of amplifying the allelic cDNA by PCR, the locus-specific primer includes a sequence encoding a tail such that the soluble HLA molecule encoded by the truncated PCR product contains a tail attached thereto that facilitates in purification of the soluble HLA molecules produced therefrom.
12. The method according to claim 7 wherein, in the step of inserting the truncated PCR product into a mammalian expression vector, the mammalian expression vector contains a promoter that facilitates increased expression of the truncated PCR product.
13. The method according to claim 7 wherein, in the step of electroporating the plasmid containing the truncated PCR product into at least one suitable host cell, the suitable host cell lacks expression of Class I HLA

molecules.

14. A method for the production of soluble HLA molecules in a cell pharm having an appropriate growth media therein, comprising the steps of:
- isolating HLA allele mRNA from a source and reverse transcribing the mRNA to obtain allelic cDNA;
 - amplifying the allelic cDNA by PCR, wherein the amplification utilizes at least one locus-specific primer that truncates the allelic cDNA, thereby resulting in a truncated PCR product having the coding regions encoding cytoplasmic and transmembrane domains of the allelic cDNA removed such that the truncated PCR product has a coding region encoding a soluble HLA molecule;
 - inserting the truncated PCR product into a mammalian expression vector to form a plasmid containing the truncated PCR product having the coding region encoding a soluble HLA molecule;
 - electroporating the plasmid containing the truncated PCR product into at least one suitable host cell; and
 - inoculating the cell pharm with the at least one suitable host cell containing the plasmid containing the truncated PCR product such that the cell pharm produces soluble HLA molecules, wherein the soluble HLA molecules are folded naturally and are trafficked

through the cell in such a way that they are identical in functional properties to an HLA molecule expressed from the HLA allele mRNA and thereby bind peptide ligands in an identical manner as full-length, cell-surface-expressed HLA molecules.

15. The method according to claim 14, further comprising the step of harvesting the soluble HLA molecules from the cell pharm.
16. The method according to claim 14 wherein the soluble HLA molecules are Class I HLA molecules or Class II HLA molecules.
17. The method according to claim 14 wherein, in the step of isolating HLA allele mRNA from a source, the source is selected from the group consisting of mammalian DNA and an immortalized cell line.
18. The method according to claim 14 wherein, in the step of amplifying the allelic cDNA by PCR, the at least one locus-specific primer is a 3' primer having a stop codon incorporated therein.
19. The method according to claim 14 wherein, in the step of amplifying the allelic cDNA by PCR, the locus-specific primer includes a sequence

encoding a tail such that the soluble HLA molecule encoded by the truncated PCR product contains a tail attached thereto that facilitates in purification of the soluble HLA molecules produced therefrom.

20. The method according to claim 14 wherein, in the step of inserting the truncated PCR product into a mammalian expression vector, the mammalian expression vector contains a promoter that facilitates increased expression of the truncated PCR product.
21. The method according to claim 14 wherein, in the step of electroporating the plasmid containing the truncated PCR product into at least one suitable host cell, the suitable host cell lacks expression of Class I HLA molecules.
22. A method for the production of soluble HLA molecules in a cell pharm having an appropriate growth media therein, comprising the steps of:
 - obtaining gDNA which encodes a HLA allele;
 - amplifying the allelic gDNA by PCR, wherein the amplification utilizes at least one locus-specific primer having a stop codon incorporated into a 3' primer thereby resulting in a truncated PCR product having the coding regions encoding cytoplasmic and

transmembrane domains of the allelic gDNA removed such that the truncated PCR product has a coding region encoding a soluble HLA molecule;

- inserting the truncated PCR product into a mammalian expression vector to form a plasmid containing the truncated PCR product having the coding region encoding the soluble HLA molecule;
- electroporating the plasmid containing the truncated PCR product into at least one suitable host cell; and
- inoculating the cell pharm with the at least one suitable host cell containing the plasmid containing the truncated PCR product such that the cell pharm produces soluble HLA molecules.

23. The method according to claim 22, further comprising the step of harvesting the soluble HLA molecules from the cell pharm.
24. The method according to claim 22, wherein the soluble HLA molecules are Class I HLA molecules or Class II HLA molecules.
25. The method according to claim 22, wherein, in the step of obtaining gDNA which encodes a HLA allele, the gDNA is obtained from blood, saliva, hair, semen, or sweat.

26. The method according to claim 22, wherein, in the step of inserting the truncated PCR product into a mammalian expression vector, the mammalian expression vector contains a promoter that facilitates increased expression of the truncated PCR product.
27. The method according to claim 22, wherein, in the step of electroporating the plasmid containing the truncated PCR product into at least one suitable host cell, the suitable host cell lacks expression of Class I HLA molecules.
28. A method for the production of soluble HLA molecules in a cell pharm having an appropriate growth media therein, comprising the steps of:
- obtaining gDNA encoding a HLA allele;
 - isolating HLA allele mRNA from gDNA and reverse transcribing the mRNA to obtain allelic cDNA;
 - amplifying the allelic cDNA by PCR, wherein the amplification utilizes at least one locus-specific primer that truncates the allelic cDNA, thereby resulting in a truncated PCR product having the coding regions encoding cytoplasmic and transmembrane domains of the allelic cDNA removed such that the truncated PCR product

- has a coding region encoding a soluble HLA molecule;
 - inserting the truncated PCR product into a mammalian expression vector to form a plasmid containing the truncated PCR product having the coding region encoding a soluble HLA molecule;
 - electroporating the plasmid containing the truncated PCR product into at least one suitable host cell; and
 - inoculating the cell pharm with the at least one suitable host cell containing the plasmid containing the truncated PCR product such that the cell pharm produces soluble HLA molecules.
29. The method according to claim 28, further comprising the step of harvesting the soluble HLA molecules from the cell pharm.
30. The method according to claim 28, wherein the soluble HLA molecules are Class I HLA molecules or Class II HLA molecules.
31. The method according to claim 28, wherein in the step of obtaining gDNA which encodes a HLA allele, the gDNA is obtained from blood, saliva, hair, semen, or sweat.
32. The method according to claim 28, wherein, in the step of amplifying the

allelic cDNA by PCR, the locus-specific primer includes a sequence encoding a tail such that the soluble HLA molecule encoded by the truncated PCR product contains a tail attached thereto that facilitates in purification of the soluble HLA molecules produced therefrom.

33. The method according to claim 28, wherein in the step of inserting the truncated PCR product into a mammalian expression vector, the mammalian expression vector contains a promoter that facilitates increased expression of the truncated PCR product.
34. The method according to claim 28, wherein in the step of electroporating the plasmid containing the truncated PCR product into at least one suitable host cell, the suitable host cell lacks expression of Class I HLA molecules.
35. A method for the production of soluble HLA molecules in a cell pharm having an appropriate growth media therein, comprising the steps of:
 - obtaining gDNA encoding a HLA allele;
 - isolating HLA allele mRNA from gDNA and reverse transcribing the mRNA to obtain allelic cDNA;
 - amplifying the allelic cDNA by PCR, wherein the amplification

- utilizes at least one locus-specific primer that truncates the allelic cDNA, thereby resulting in a truncated PCR product having the coding regions encoding cytoplasmic and transmembrane domains of the allelic cDNA removed such that the truncated PCR product has a coding region encoding a soluble HLA molecule;
- inserting the truncated PCR product into a mammalian expression vector to form a plasmid containing the truncated PCR product having the coding region encoding a soluble HLA molecule;
 - electroporating the plasmid containing the truncated PCR product into at least one suitable host cell; and
 - inoculating the cell pharm with the at least one suitable host cell containing the plasmid containing the truncated PCR product such that the cell pharm produces soluble HLA molecules, wherein the soluble HLA molecules are folded naturally and are trafficked through the cell in such a way that they are identical in functional properties to an HLA molecule expressed from the HLA allele mRNA and thereby bind peptide ligands in an identical manner as full-length, cell-surface-expressed HLA molecules.
36. The method according to claim 35, further comprising the step of harvesting the soluble HLA molecules from the cell pharm.

37. The method according to claim 35, wherein the soluble HLA molecules are Class I HLA molecules or Class II HLA molecules.
38. The method according to claim 35, wherein in the step of obtaining gDNA which encodes a HLA allele, the gDNA is obtained from blood, saliva, hair, semen, or sweat.
39. The method according to claim 35, wherein in the step of amplifying the allelic cDNA by PCR, the at least one locus-specific primer is a 3' primer having a stop codon incorporated therein.
40. The method according to claim 14 wherein, in the step of amplifying the allelic cDNA by PCR, the locus-specific primer includes a sequence encoding a tail such that the soluble HLA molecule encoded by the truncated PCR product contains a tail attached thereto that facilitates in purification of the soluble HLA molecules produced therefrom.
41. The method according to claim 14 wherein, in the step of inserting the truncated PCR product into a mammalian expression vector, the mammalian expression vector contains a promoter that facilitates

increased expression of the truncated PCR product.

42. The method according to claim 14 wherein, in the step of electroporating the plasmid containing the truncated PCR product into at least one suitable host cell, the suitable host cell lacks expression of Class I HLA molecules.
43. A multimeric HLA complex, comprising:
 - a substrate; and
 - at least two soluble HLA molecules attached to the substrate.
44. The multimeric HLA complex of claim 43, wherein the multimeric HLA complex is used to test functionality of peptide ligands bound by the at least two soluble HLA molecules.
45. A method for producing HLA molecules, comprising the steps of:
 - isolating HLA allele mRNA from a source and reverse transcribing the mRNA to obtain allelic cDNA;
 - amplifying the allelic cDNA, wherein the amplification utilizes at least one locus-specific primer having a stop codon incorporated into a 3' primer thereby resulting in a truncated product;

- inserting the truncated product into a vector to form a plasmid containing the truncated product;
- electroporating the plasmid containing the truncated product into at least one suitable host cell; and
- inoculating a cell pharm with the at least one suitable host cell containing the plasmid containing the truncated product such that the cell pharm produces HLA molecules.

46. A method for producing HLA molecules, comprising the steps of:

- isolating HLA allele mRNA from a source and reverse transcribing the mRNA to obtain allelic cDNA;
- amplifying the allelic cDNA, wherein the amplification utilizes at least one locus-specific primer that truncates the allelic cDNA, thereby resulting in a truncated product;
- inserting the truncated product into a vector to form a plasmid containing the truncated product;
- electroporating the plasmid containing the truncated product into at least one suitable host cell; and
- inoculating a cell pharm with the at least one suitable host cell containing the plasmid containing the truncated product such that the cell pharm produces HLA molecules.

47. A method for producing HLA molecules, comprising the steps of:
- isolating HLA allele mRNA from a source and reverse transcribing the mRNA to obtain allelic cDNA;
 - amplifying the allelic cDNA by PCR, wherein the amplification utilizes at least one locus-specific primer that truncates the allelic cDNA, thereby resulting in a truncated PCR product;
 - inserting the truncated PCR product into a mammalian expression vector to form a plasmid containing the truncated PCR product;
 - electroporating the plasmid containing the truncated PCR product into at least one suitable host cell; and
 - inoculating a cell pharm with the at least one suitable host cell containing the plasmid containing the truncated PCR product such that the cell pharm produces HLA molecules.
48. A method for the producing HLA molecules, comprising the steps of:
- obtaining gDNA which encodes a HLA allele;
 - amplifying the allelic gDNA, wherein the amplification utilizes at least one locus-specific primer having a stop codon incorporated into a 3' primer thereby resulting in a truncated product;
 - inserting the truncated product into a vector to form a plasmid

- containing the truncated product;
 - electroporating the plasmid containing the truncated product into at least one suitable host cell; and
 - inoculating the cell pharm with the at least one suitable host cell containing the plasmid containing the truncated product such that the cell pharm produces HLA molecules.
49. A method for the producing HLA molecules, comprising the steps of:
- obtaining gDNA encoding a HLA allele;
 - isolating HLA allele mRNA from gDNA and reverse transcribing the mRNA to obtain allelic cDNA;
 - amplifying the allelic cDNA, wherein the amplification utilizes at least one locus-specific primer that truncates the allelic cDNA, thereby resulting in a truncated product;
 - inserting the truncated product into a vector to form a plasmid containing the truncated product;
 - electroporating the plasmid containing the truncated product into at least one suitable host cell; and
 - inoculating the cell pharm with the at least one suitable host cell containing the plasmid containing the truncated product such that the cell pharm produces HLA molecules.

50. A method for the producing HLA molecules, comprising the steps of:
- obtaining gDNA encoding a HLA allele;
 - isolating HLA allele mRNA from gDNA and reverse transcribing the mRNA to obtain allelic cDNA;
 - amplifying the allelic cDNA by PCR, wherein the amplification utilizes at least one locus-specific primer that truncates the allelic cDNA, thereby resulting in a truncated PCR product;
 - inserting the truncated PCR product into a mammalian expression vector to form a plasmid containing the truncated PCR product;
 - electroporating the plasmid containing the truncated PCR product into at least one suitable host cell; and
 - inoculating a cell pharm with the at least one suitable host cell containing the plasmid containing the truncated PCR product such that the cell pharm produces HLA molecules.
51. An HLA complex, comprising:
- a substrate; and
 - at least one HLA molecule attached to the substrate.

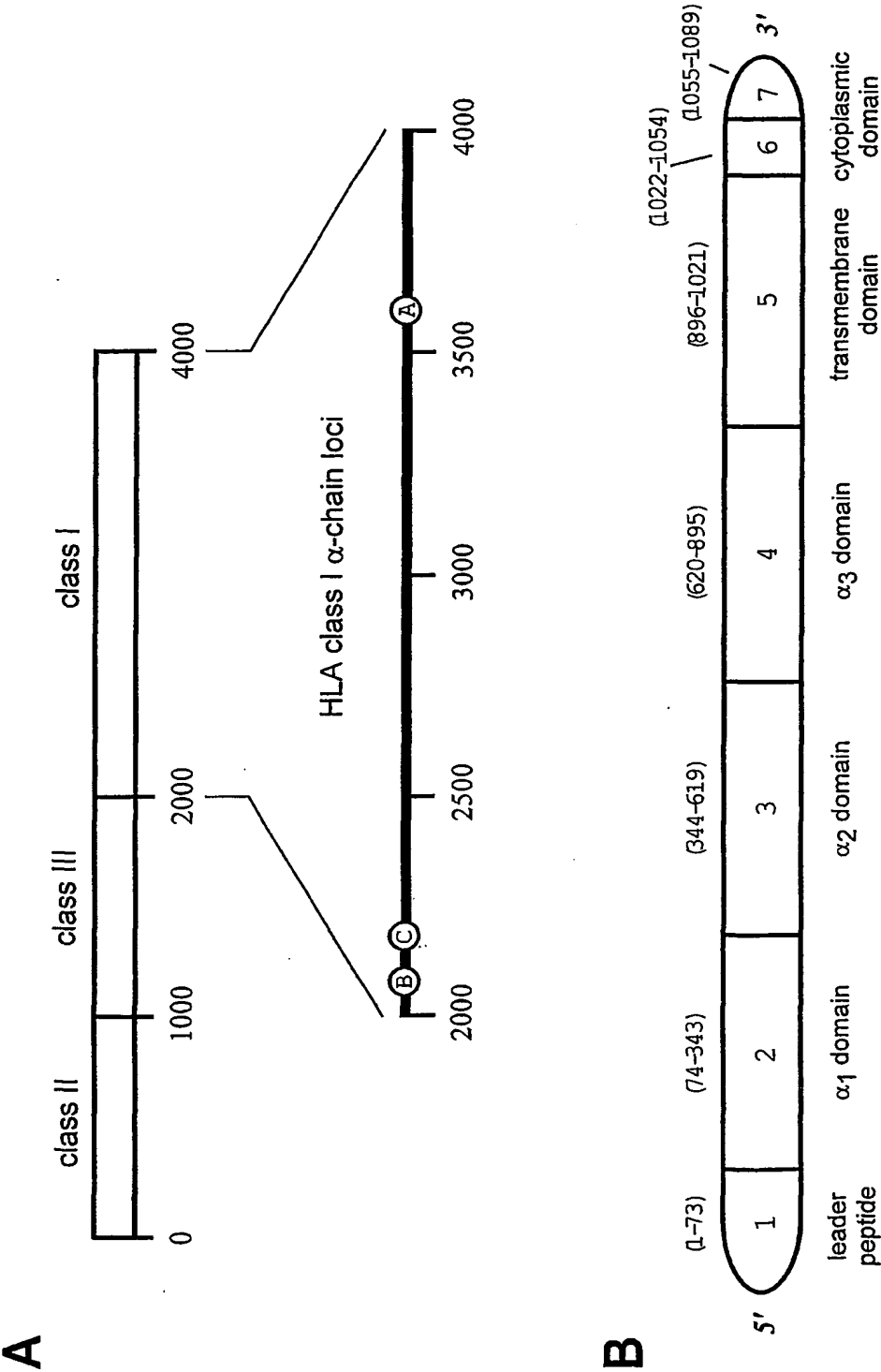


Fig. 1

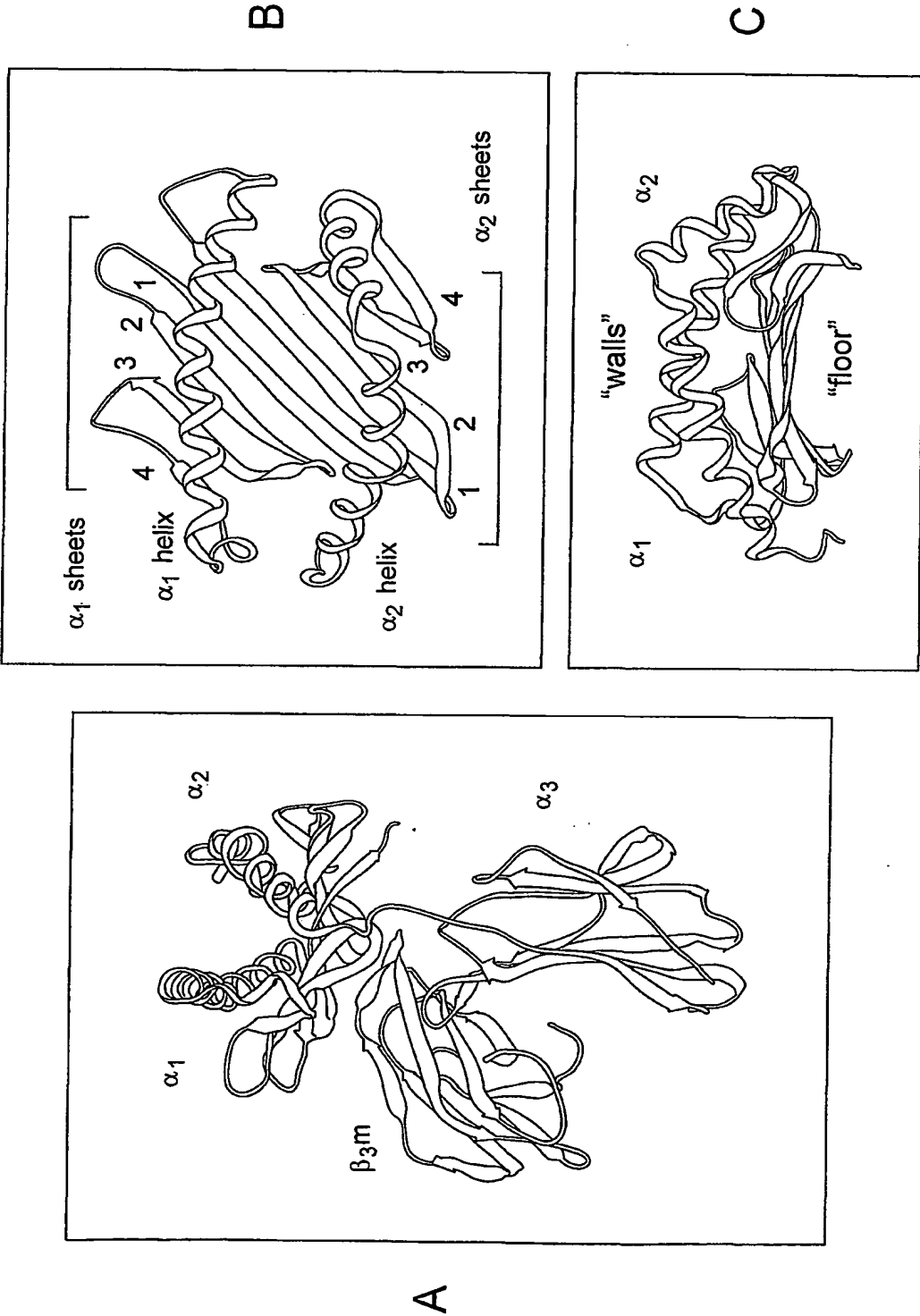


Fig. 2

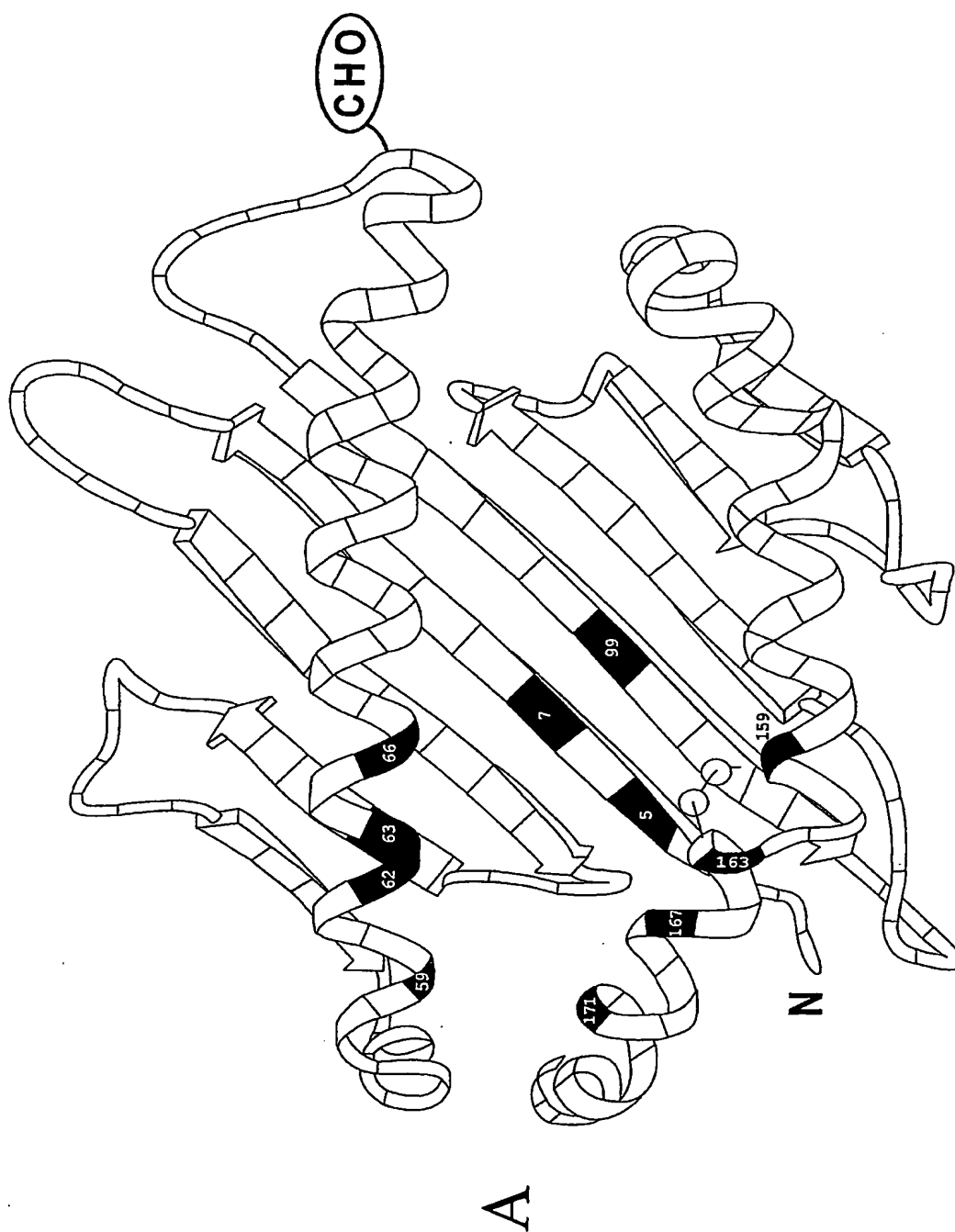


Fig. 3 1 of 6

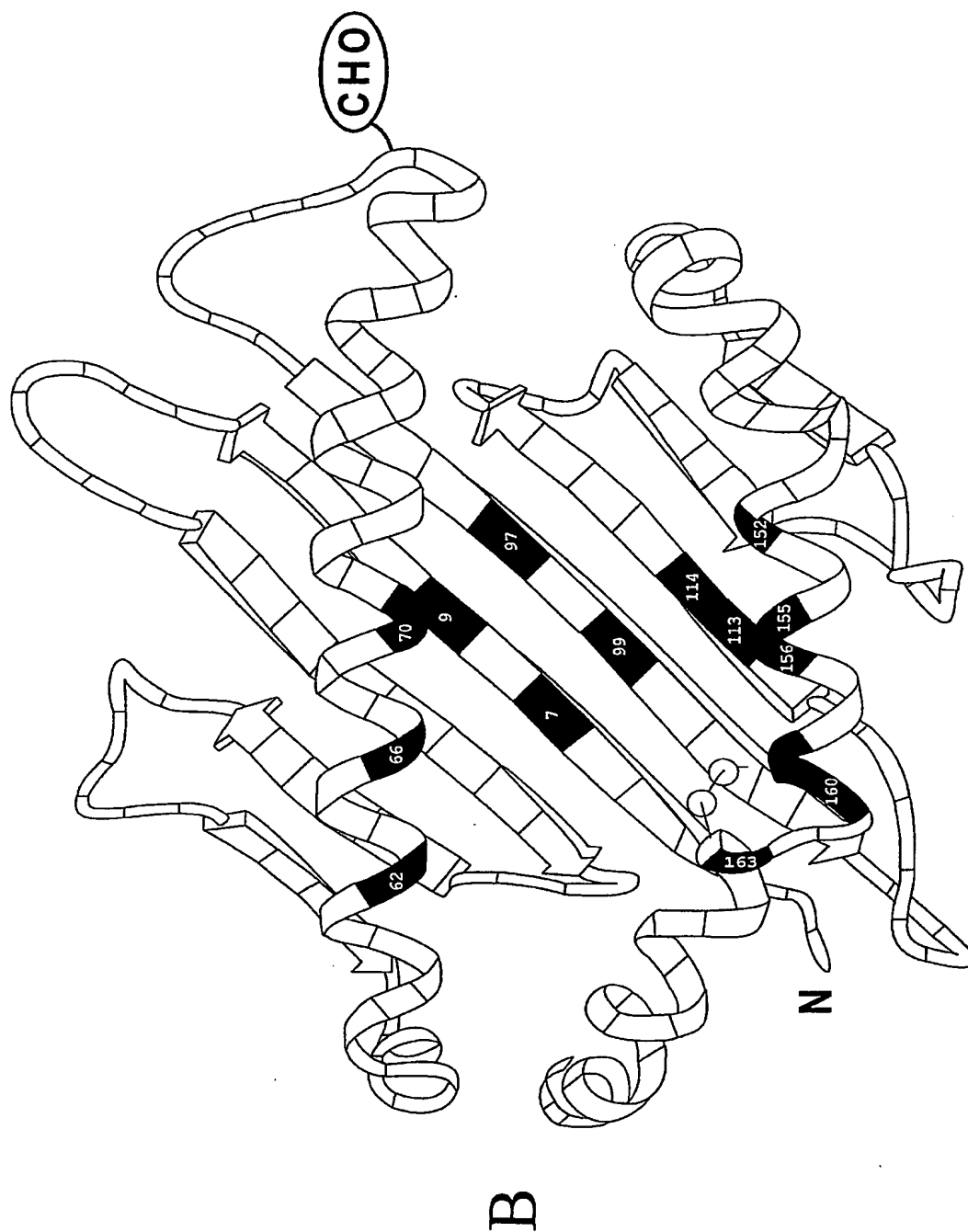


Fig. 3 2 of 6

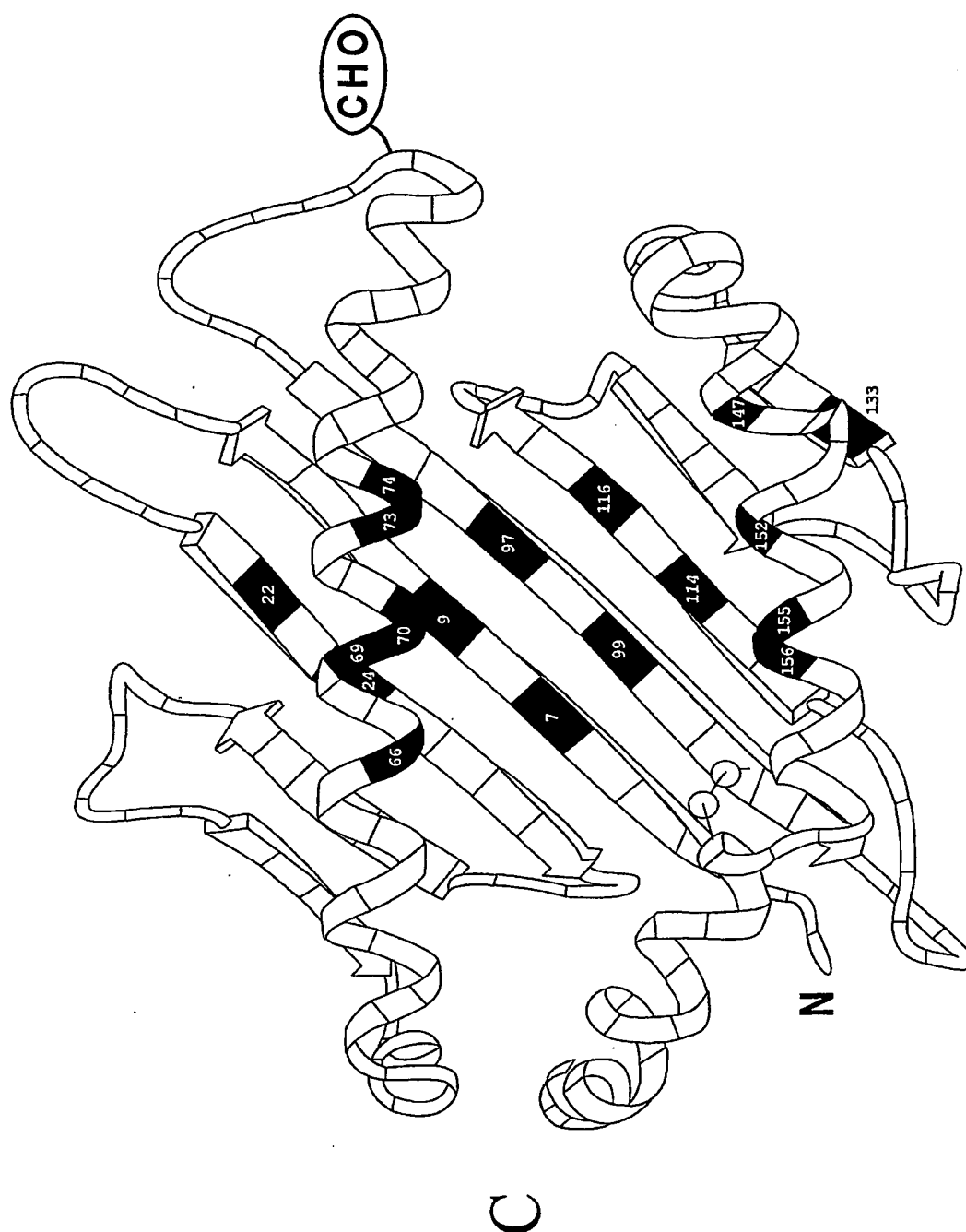


Fig. 3 3 of 6

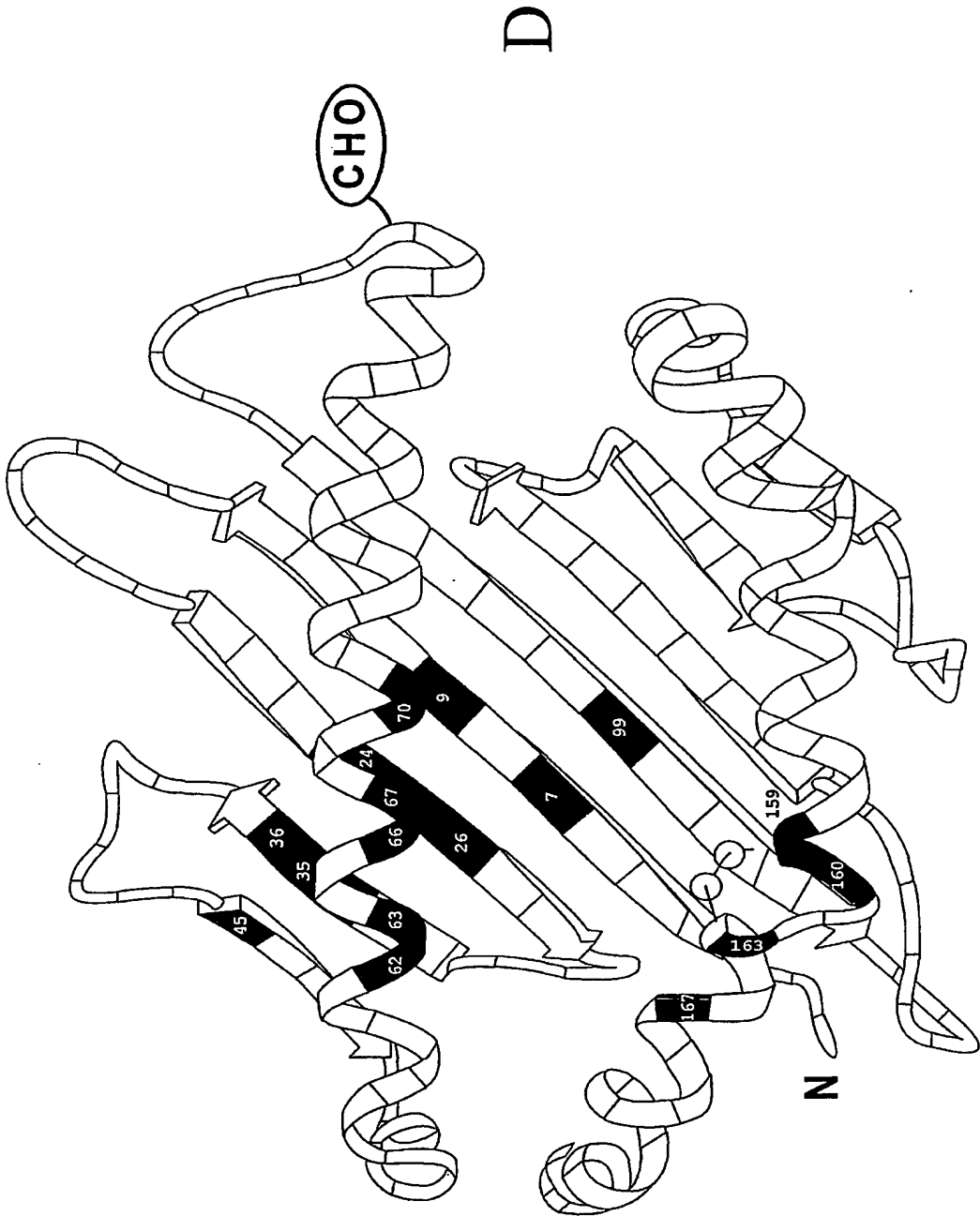


Fig. 3 4 of 6

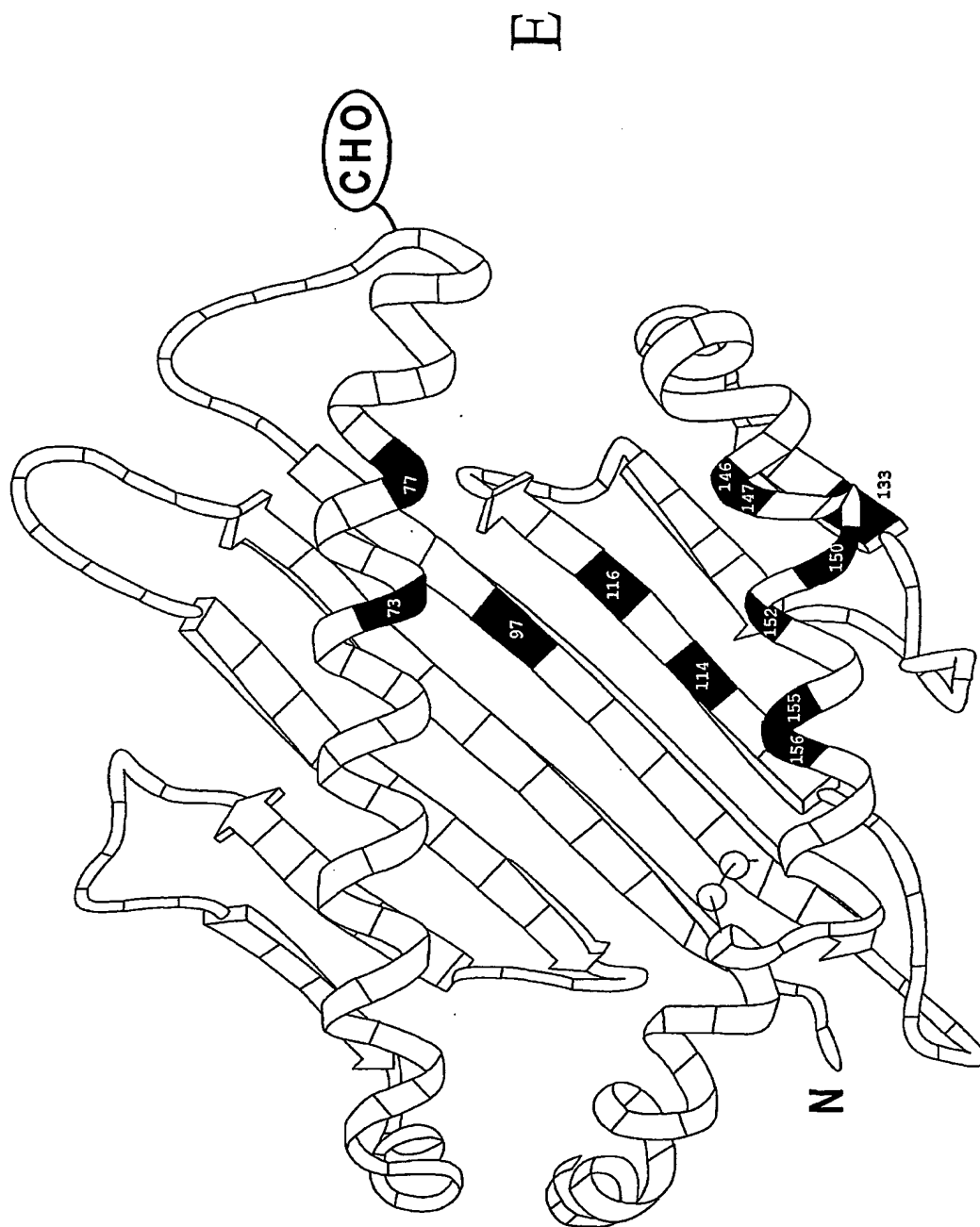


Fig. 3 5 of 6

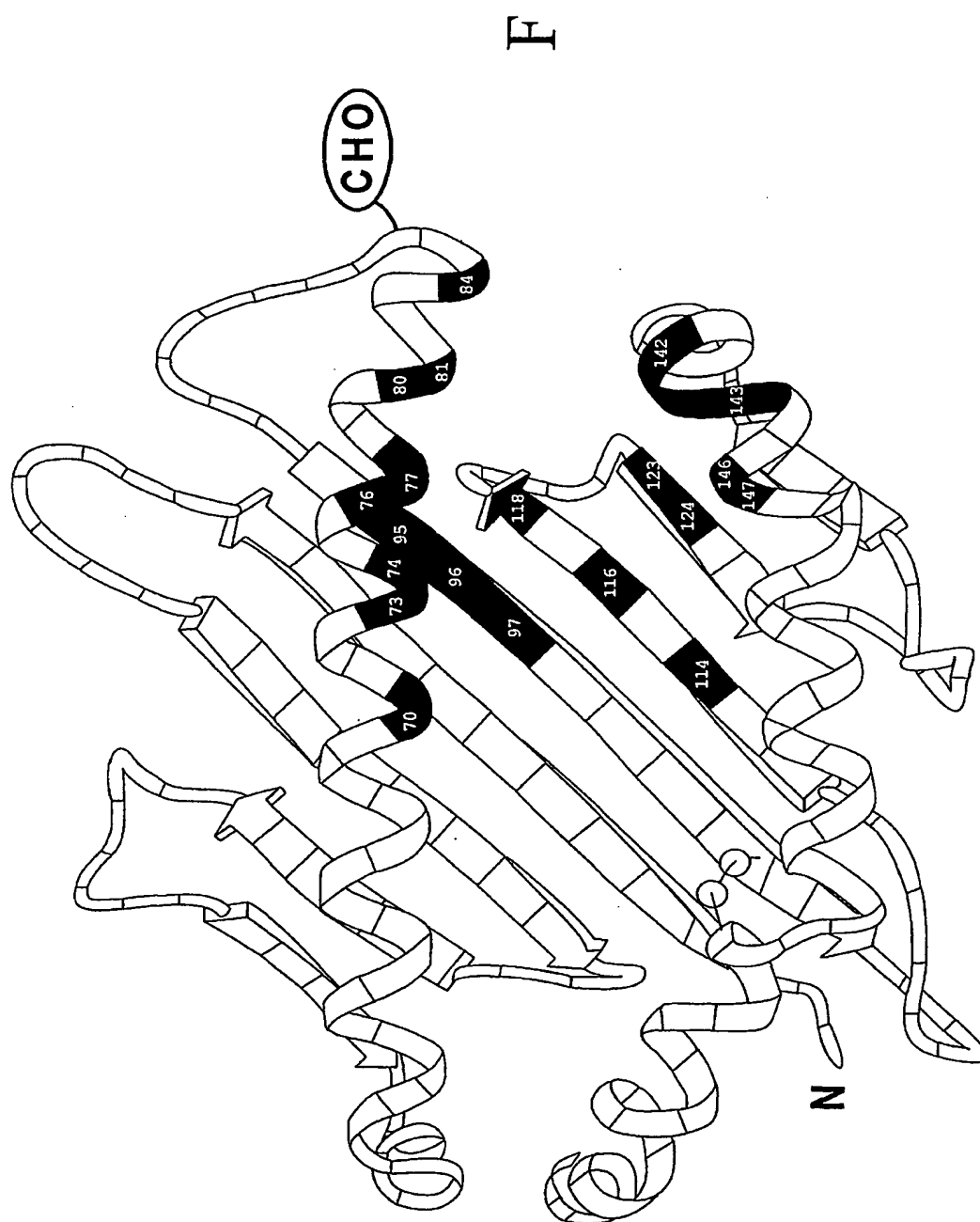


Fig. 3 6 of 6

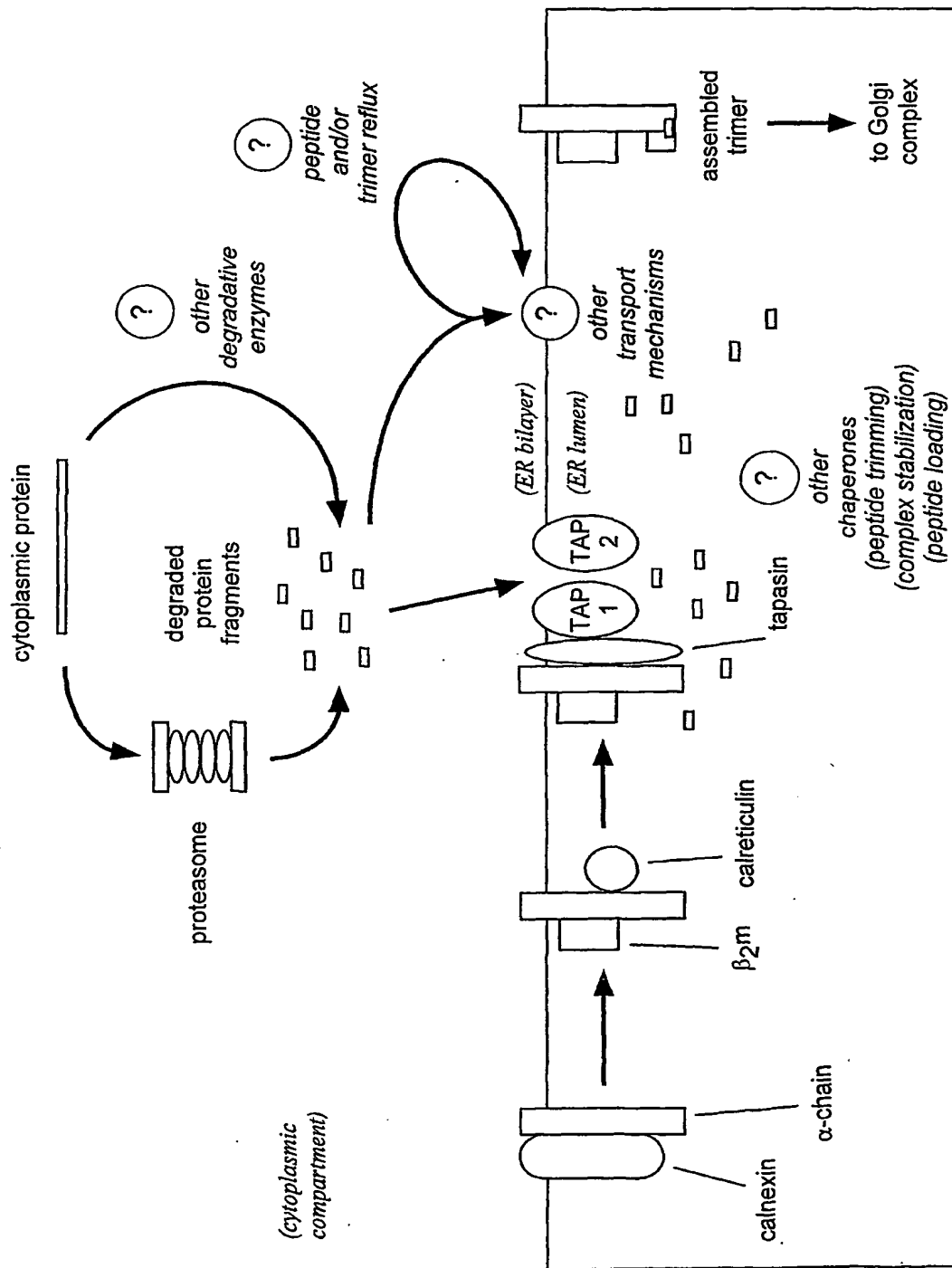


Fig. 4

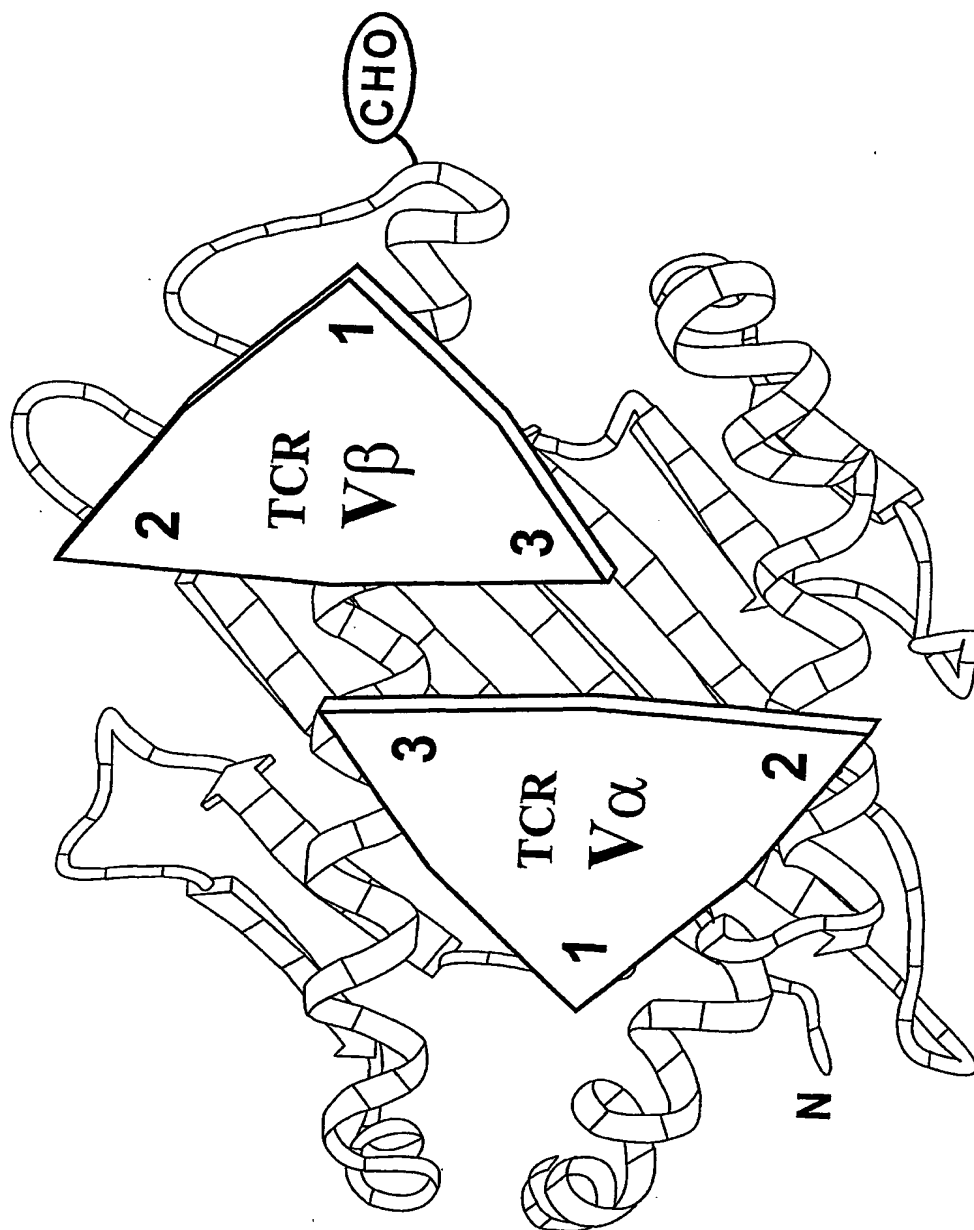


Fig. 5

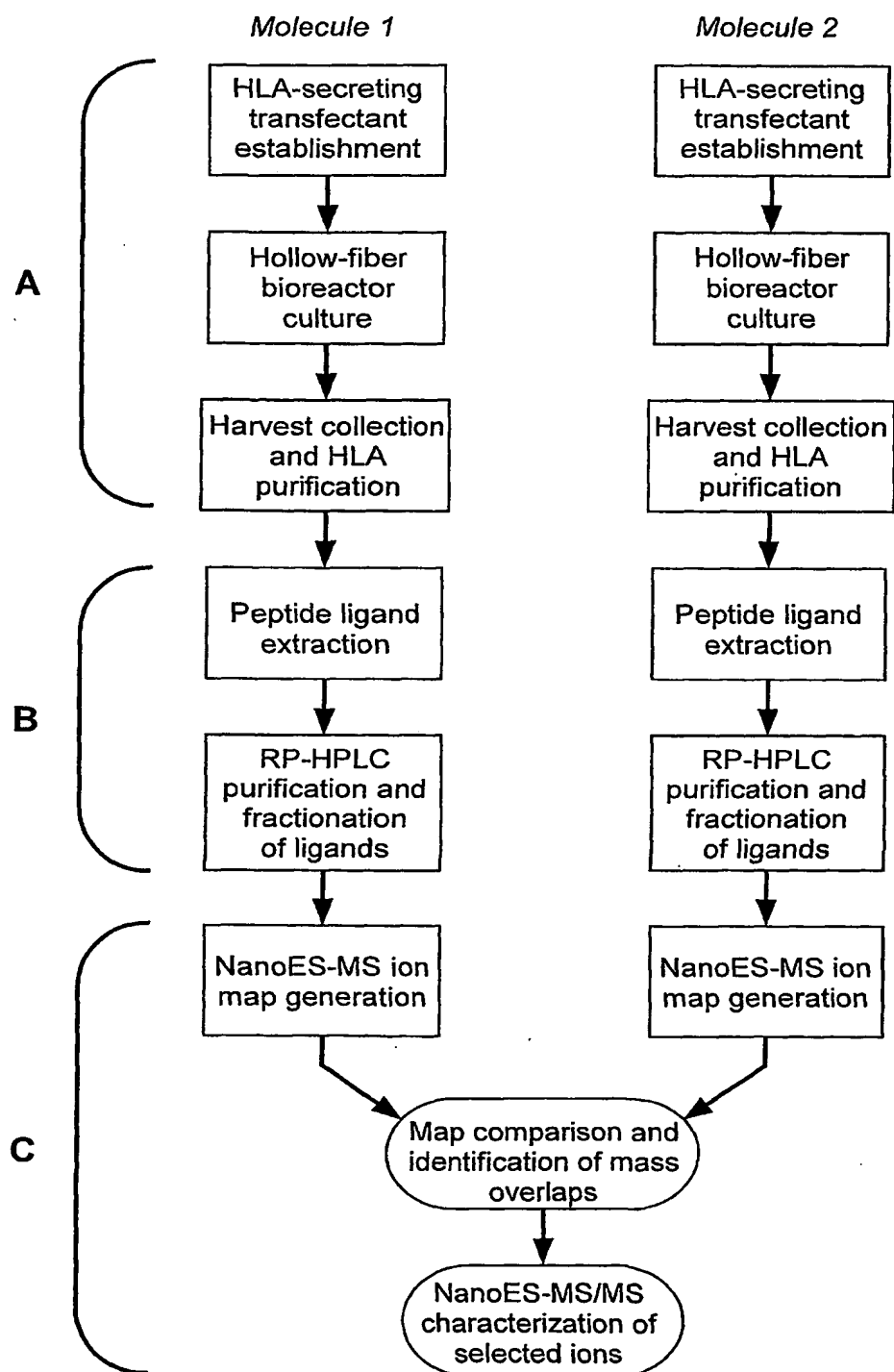


Fig. 6

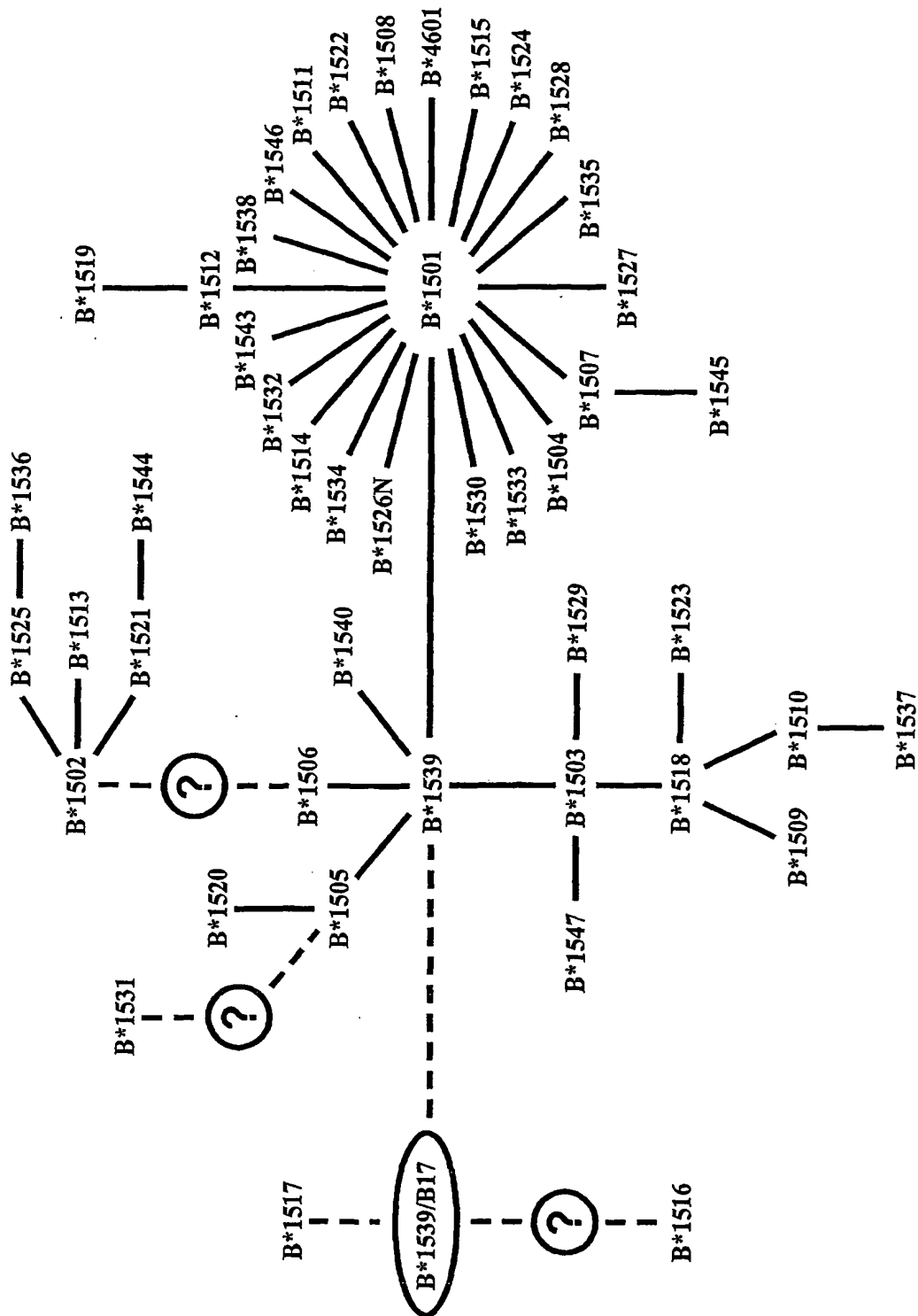


Fig. 7

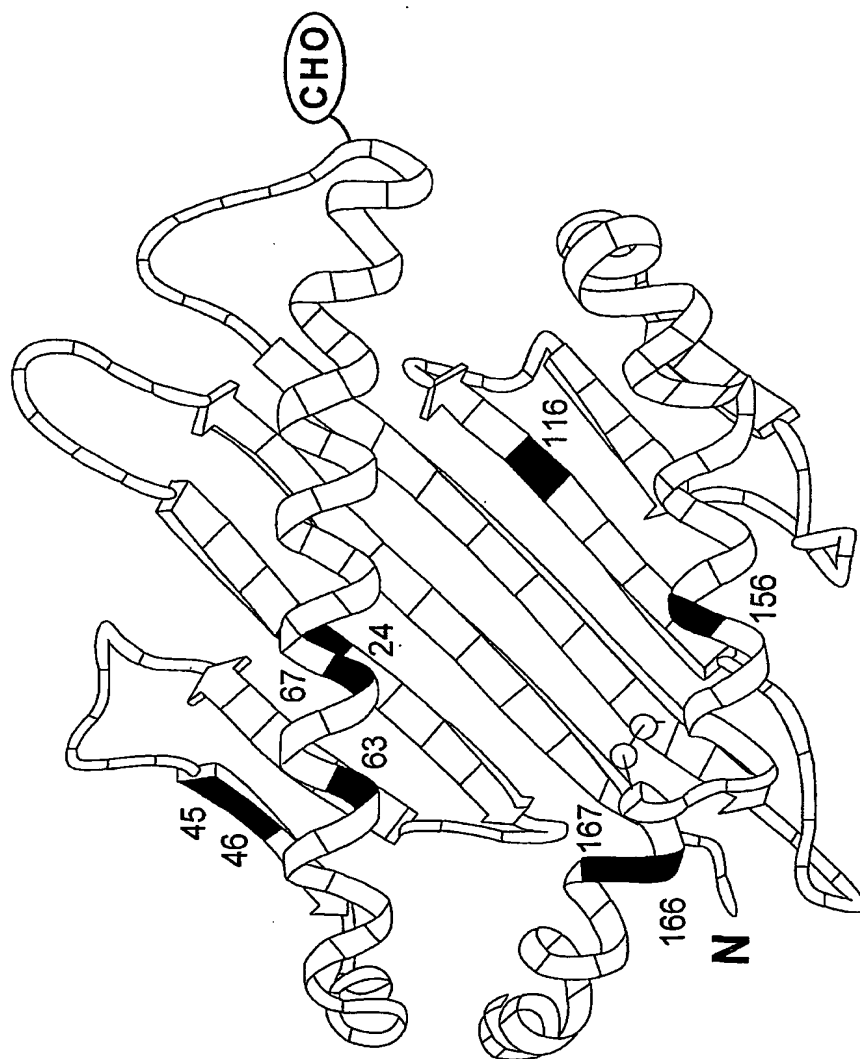


Fig. 8

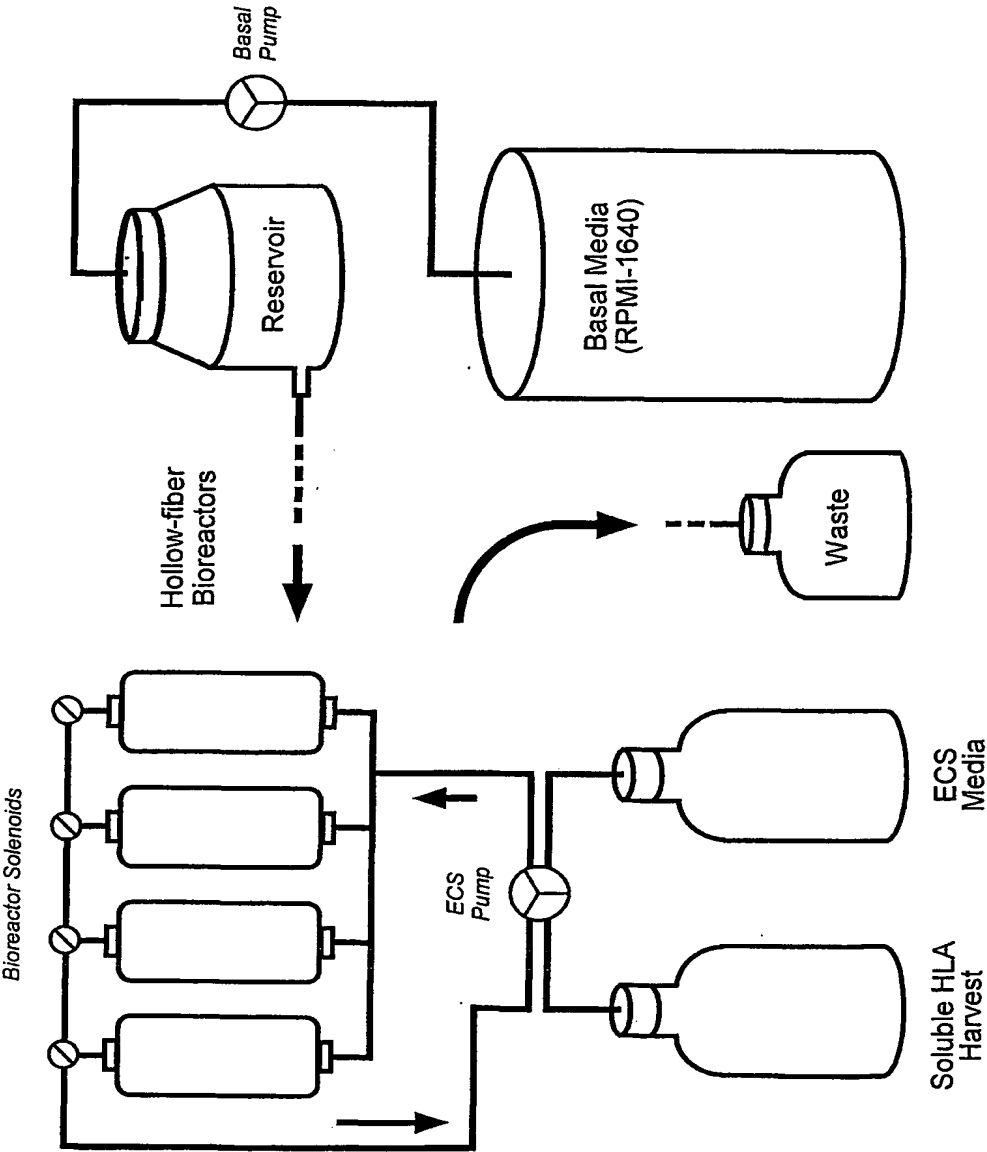


Fig. 9

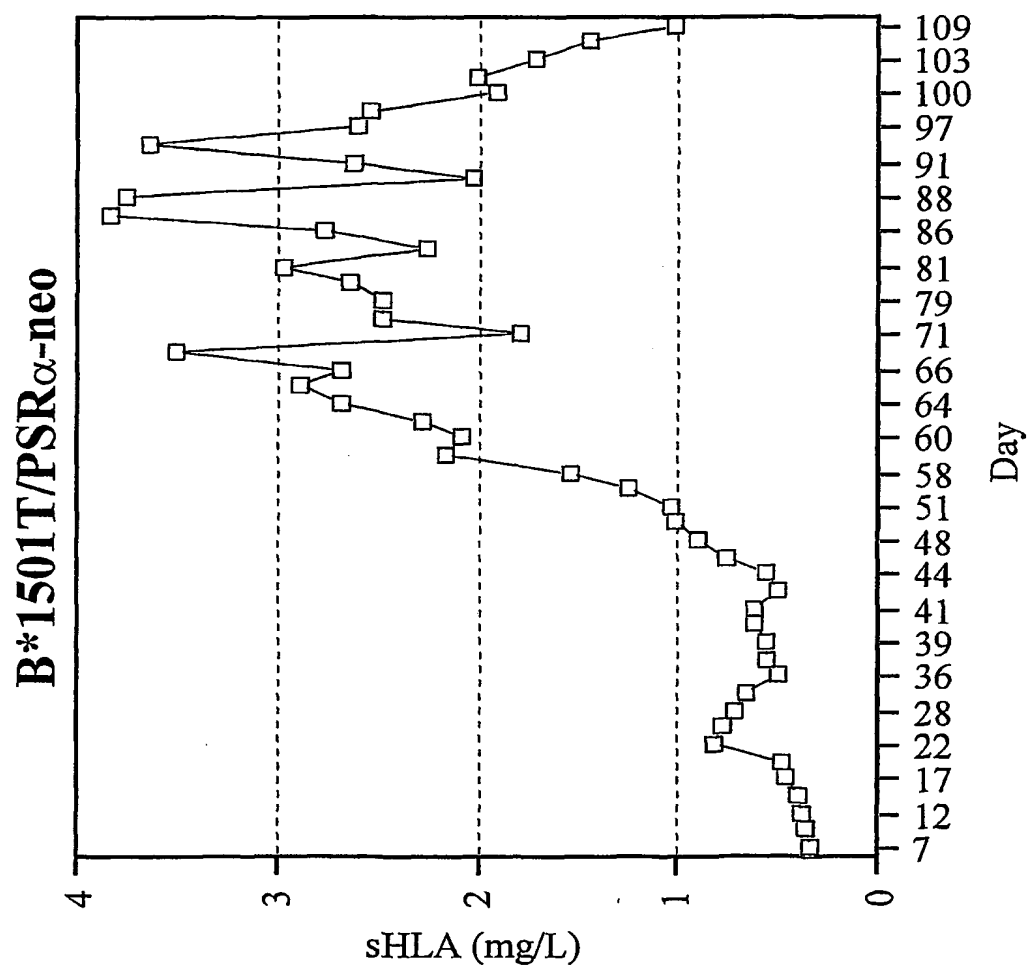


Fig. 10 1 of 2

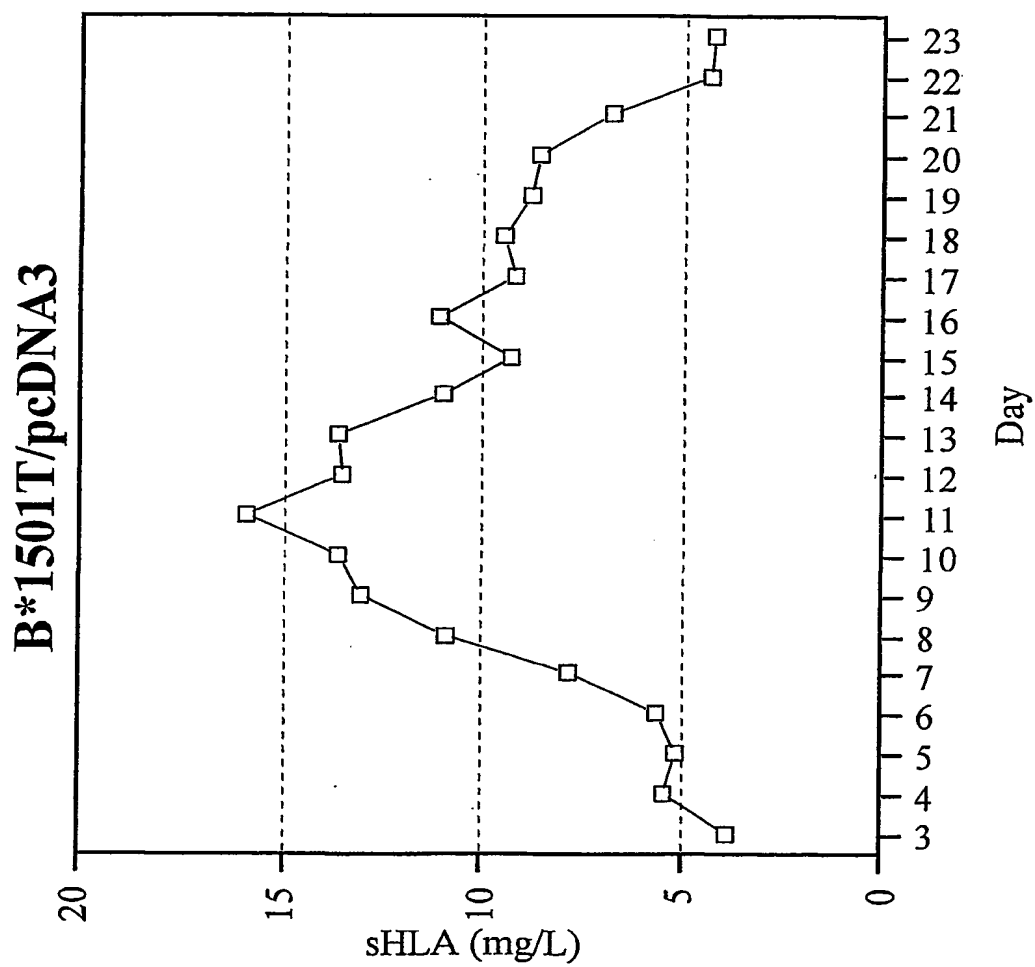


Fig. 10 2 of 2

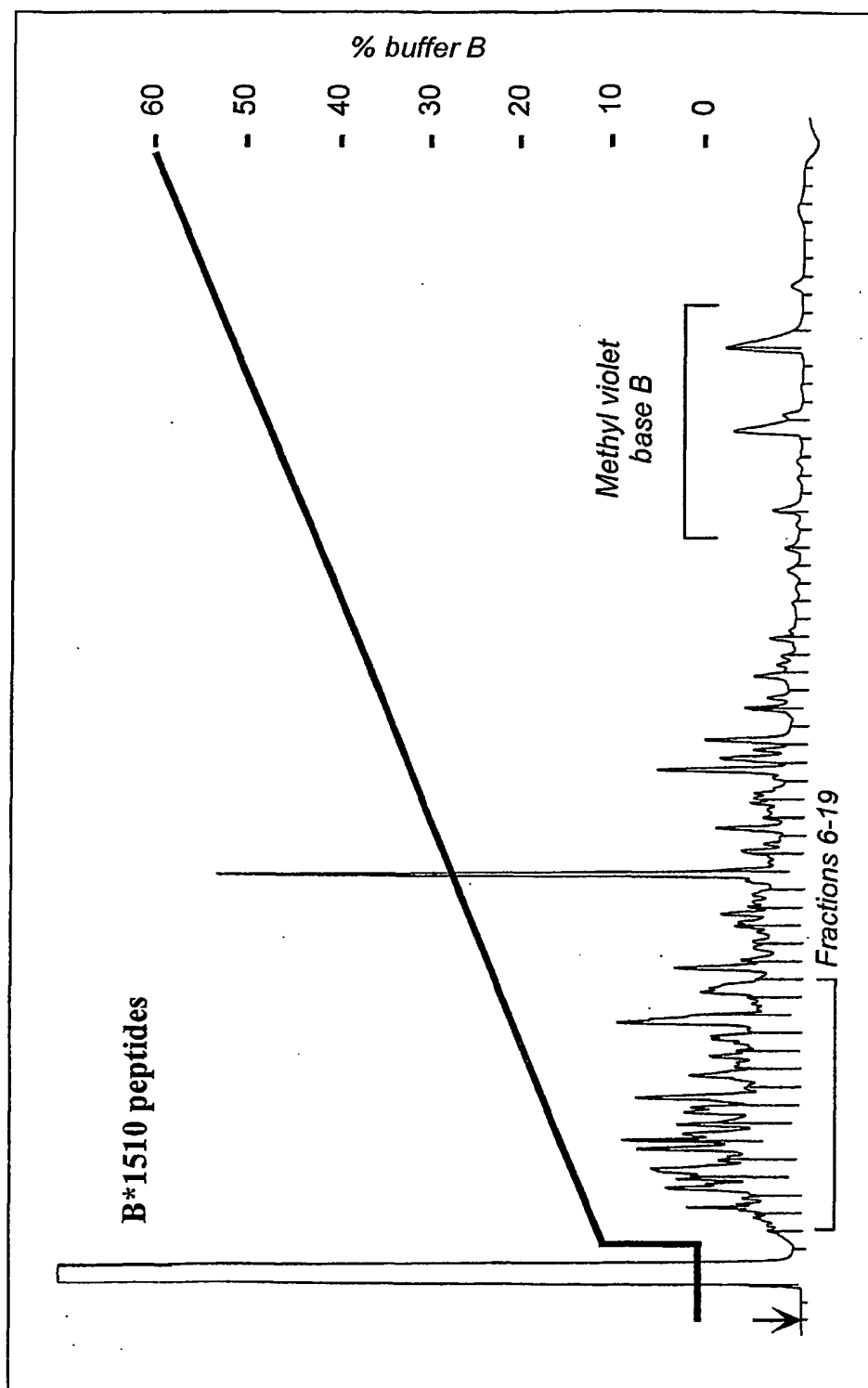


Fig. 11

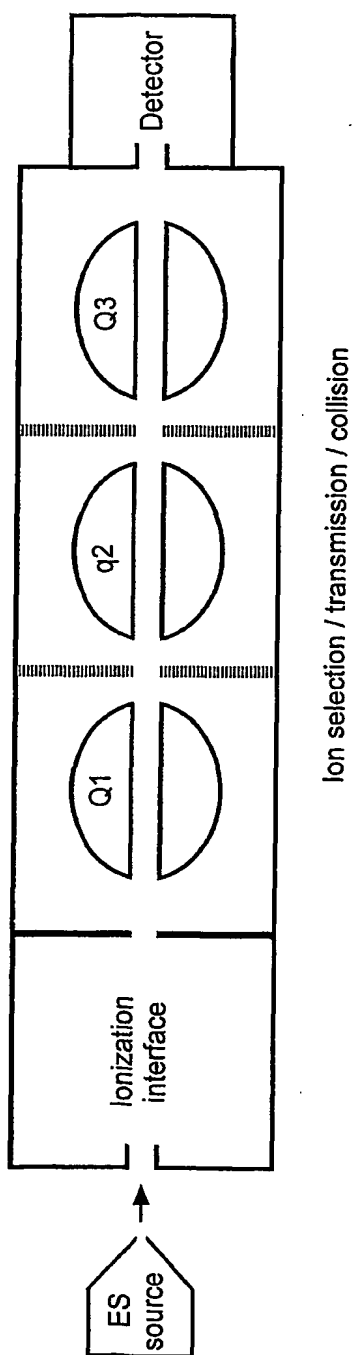
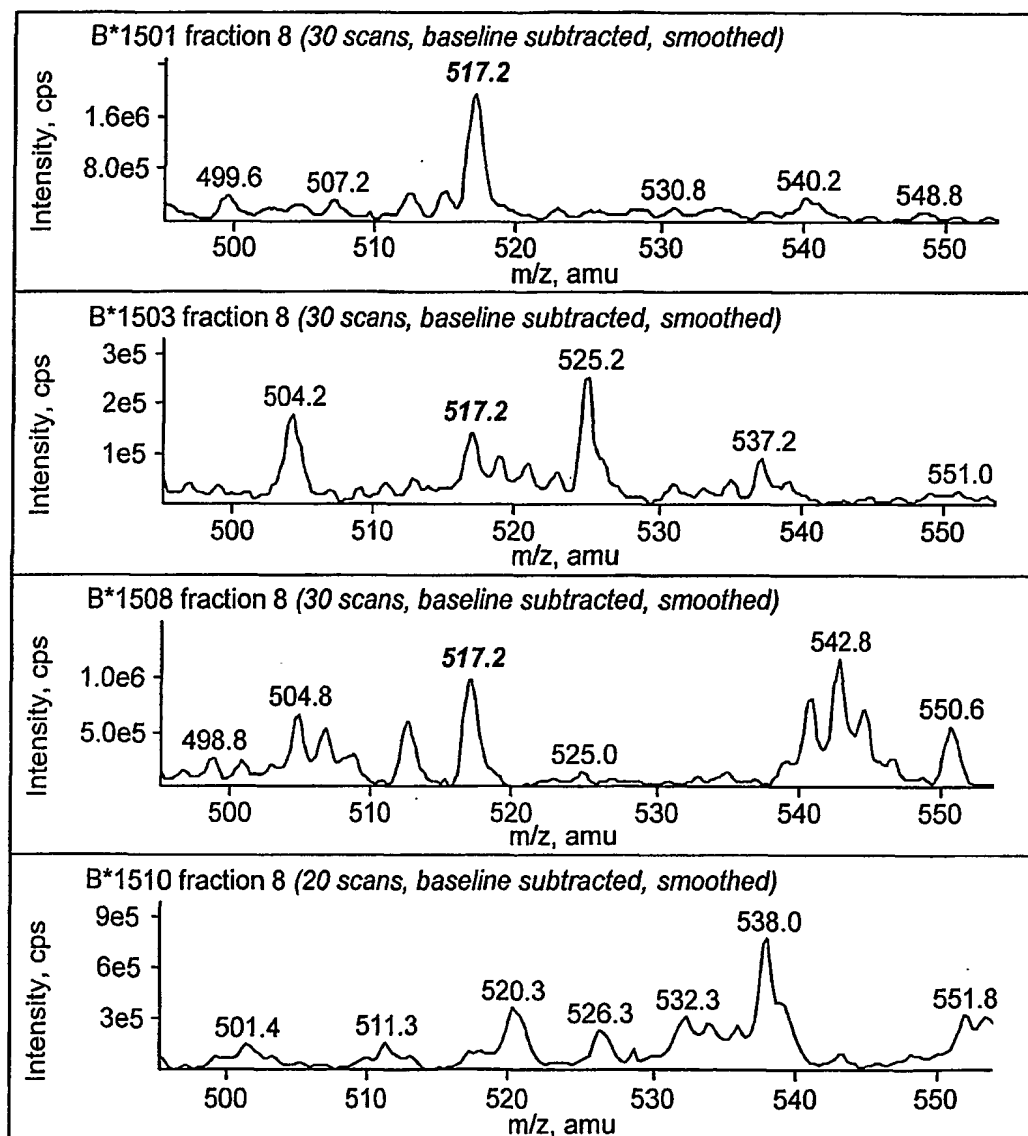
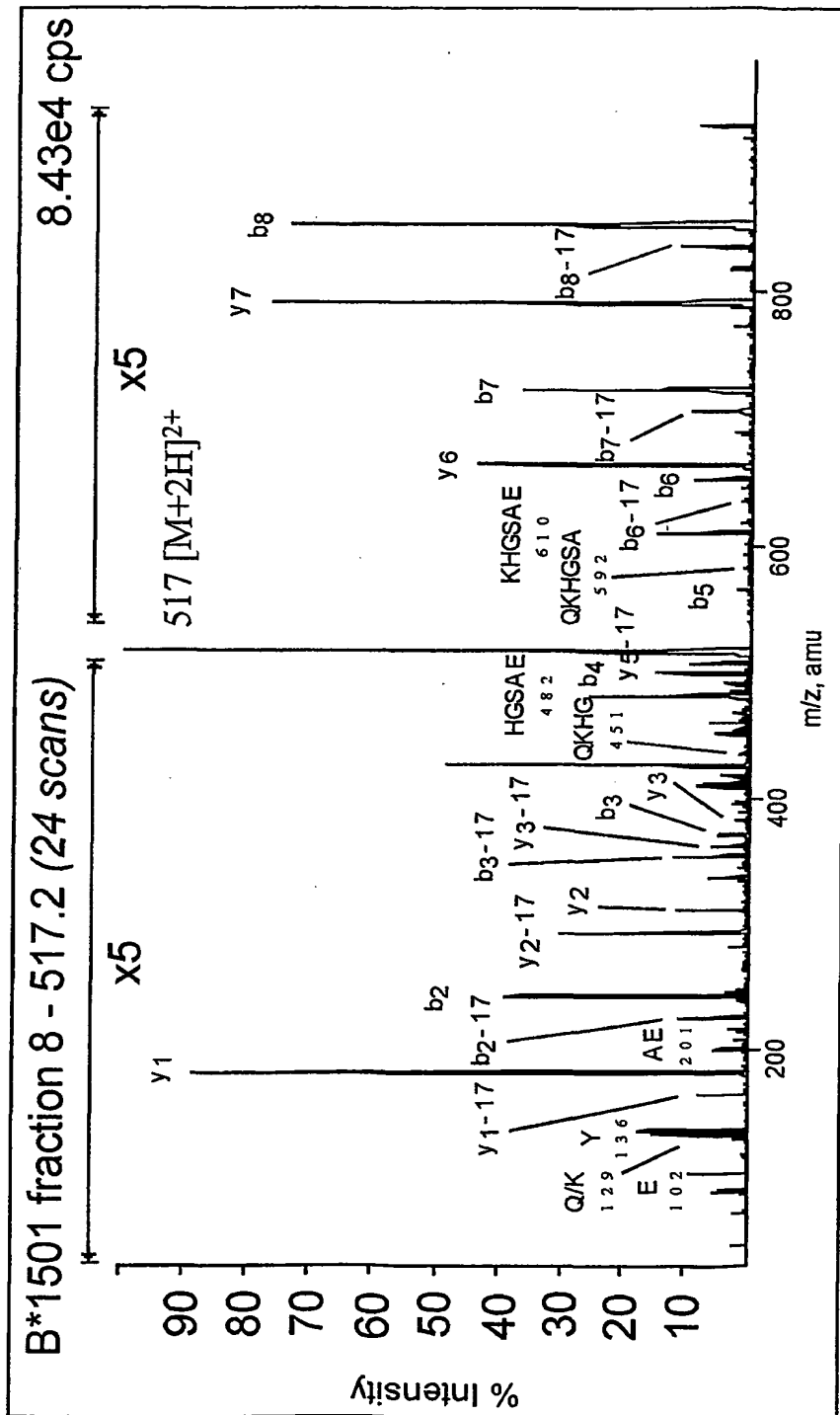


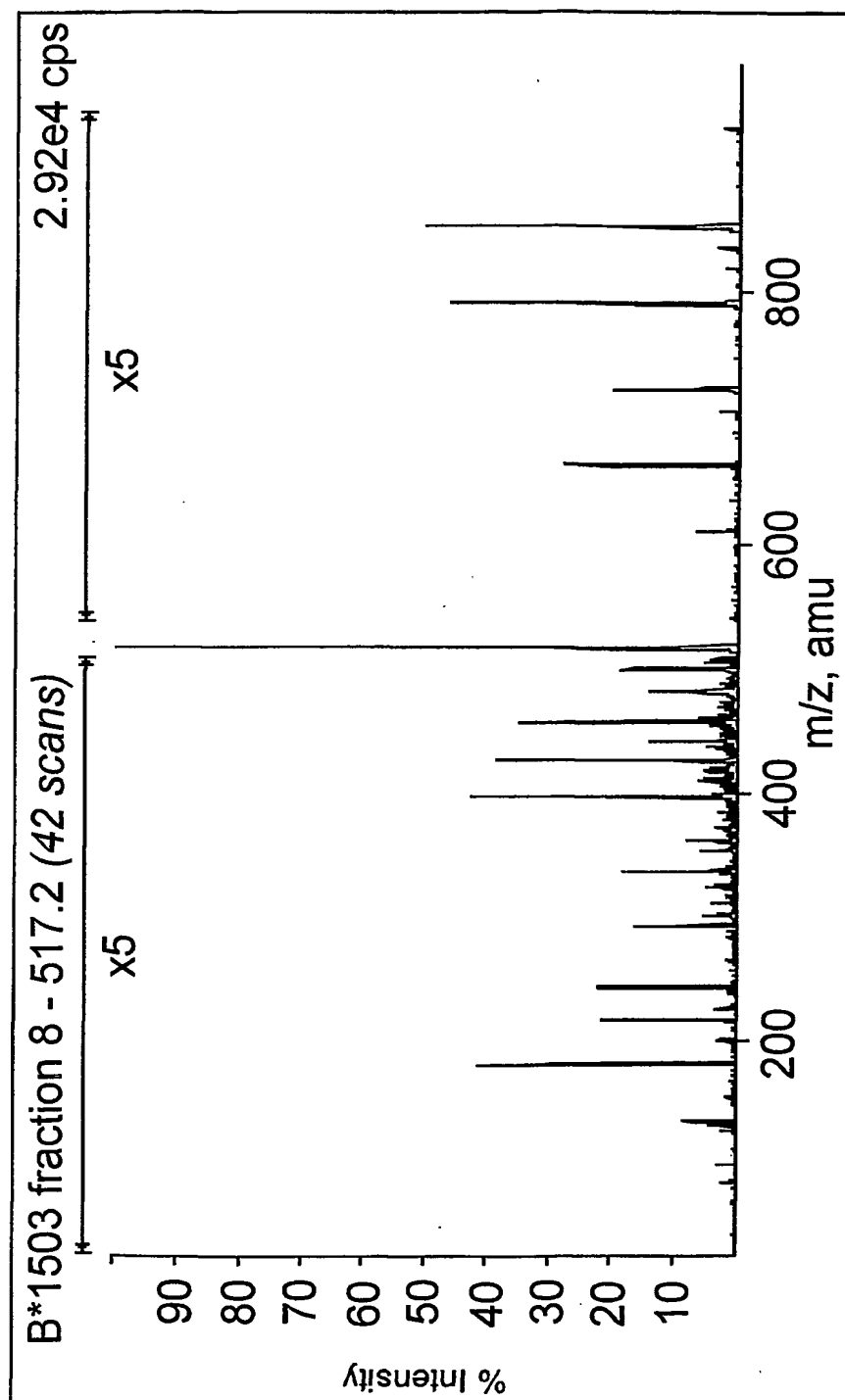
Fig. 12



Graph A

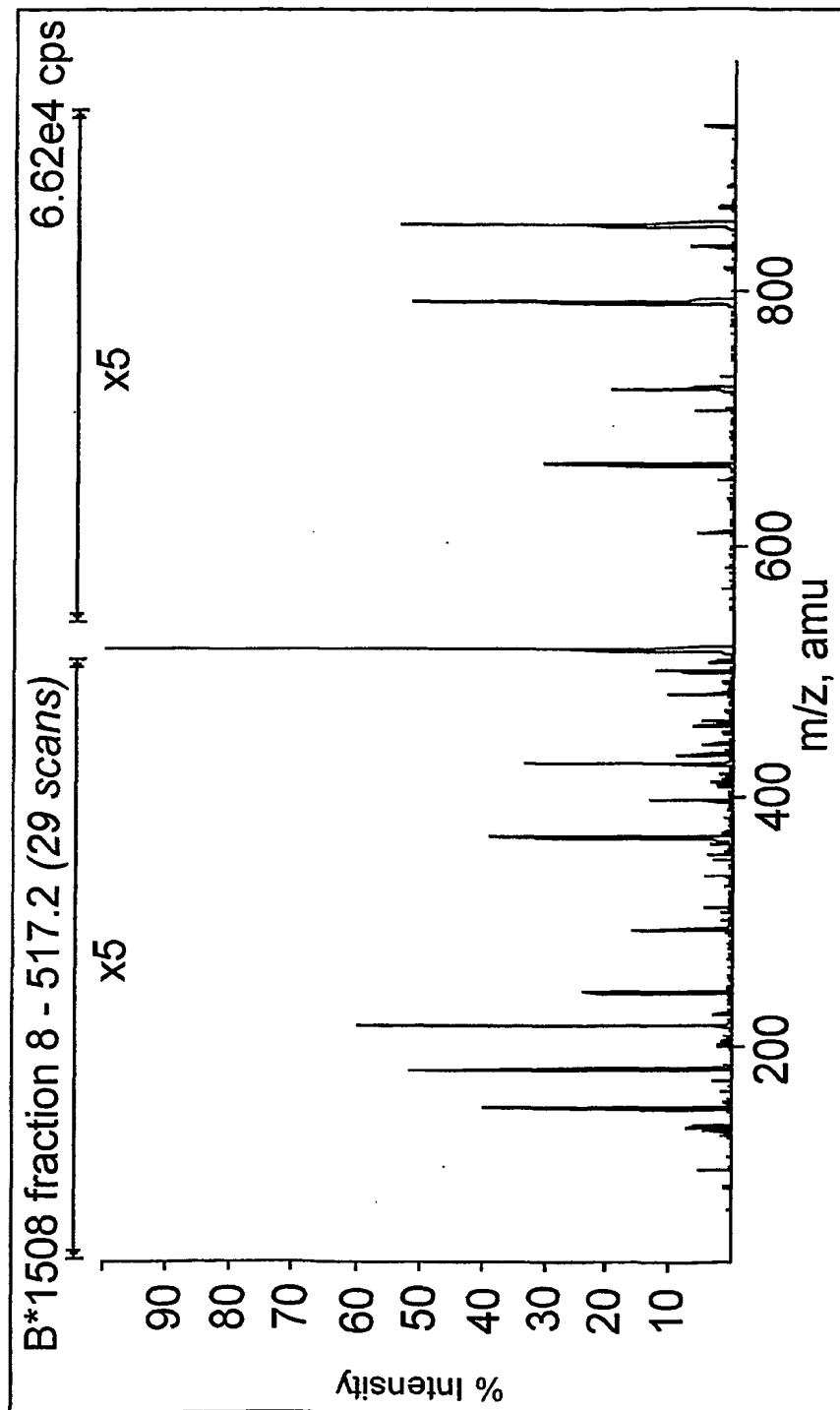
Fig. 13 1 of 5





Graph B 2 of 3

Fig. 13 3 of 5



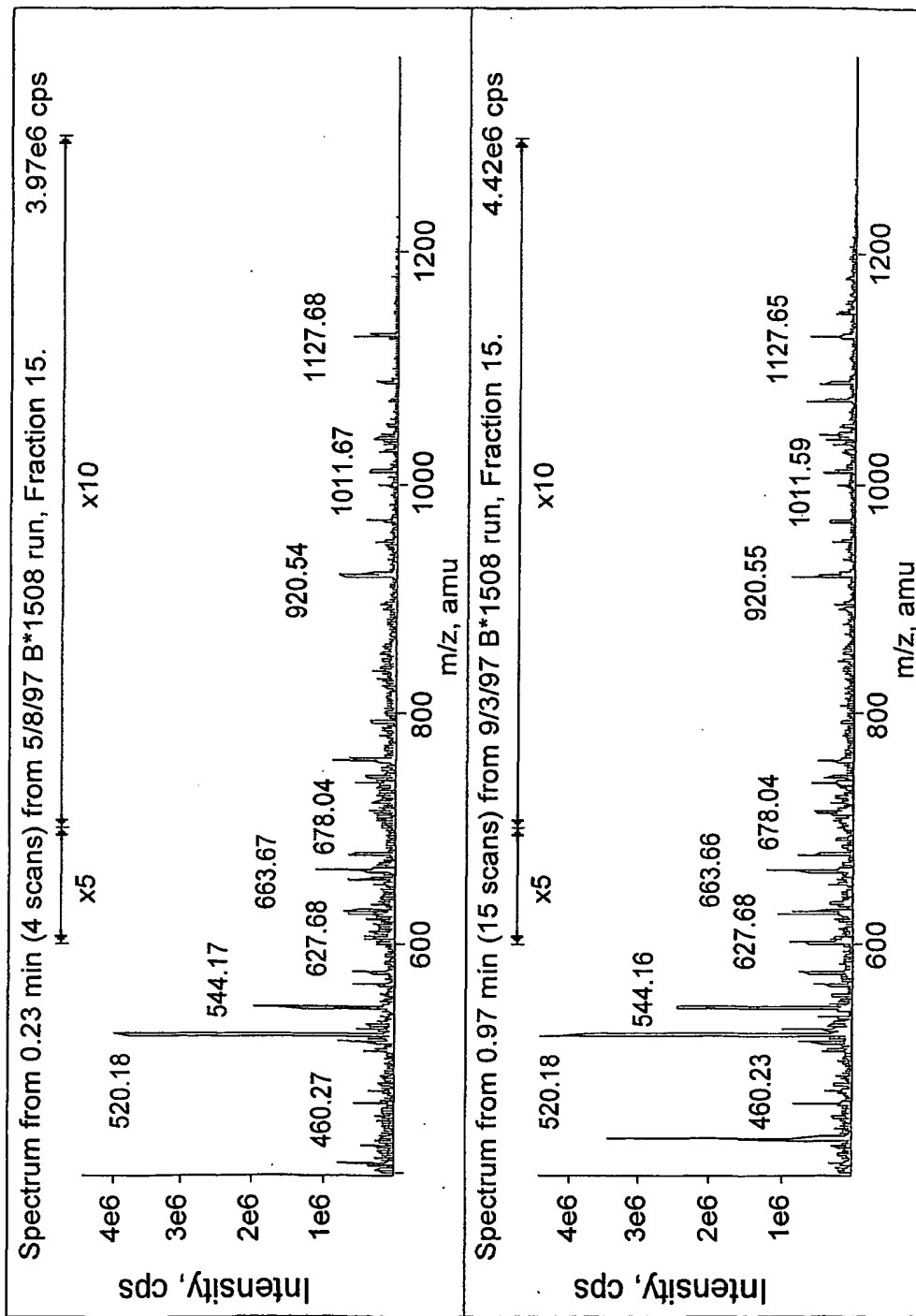
Graph B 3 of 3

Fig. 13 4 of 5

y-ions:	9	1033	N	115	1
	8	919	Q	243	2
	7	791	Z	371	3
	6	663	H	508	4
	5	526	G	565	5
	4	469	S	652	6
	3	382	A	723	7
	2	311	E	852	8
	1	182	Y	1015	9
b-ions:					

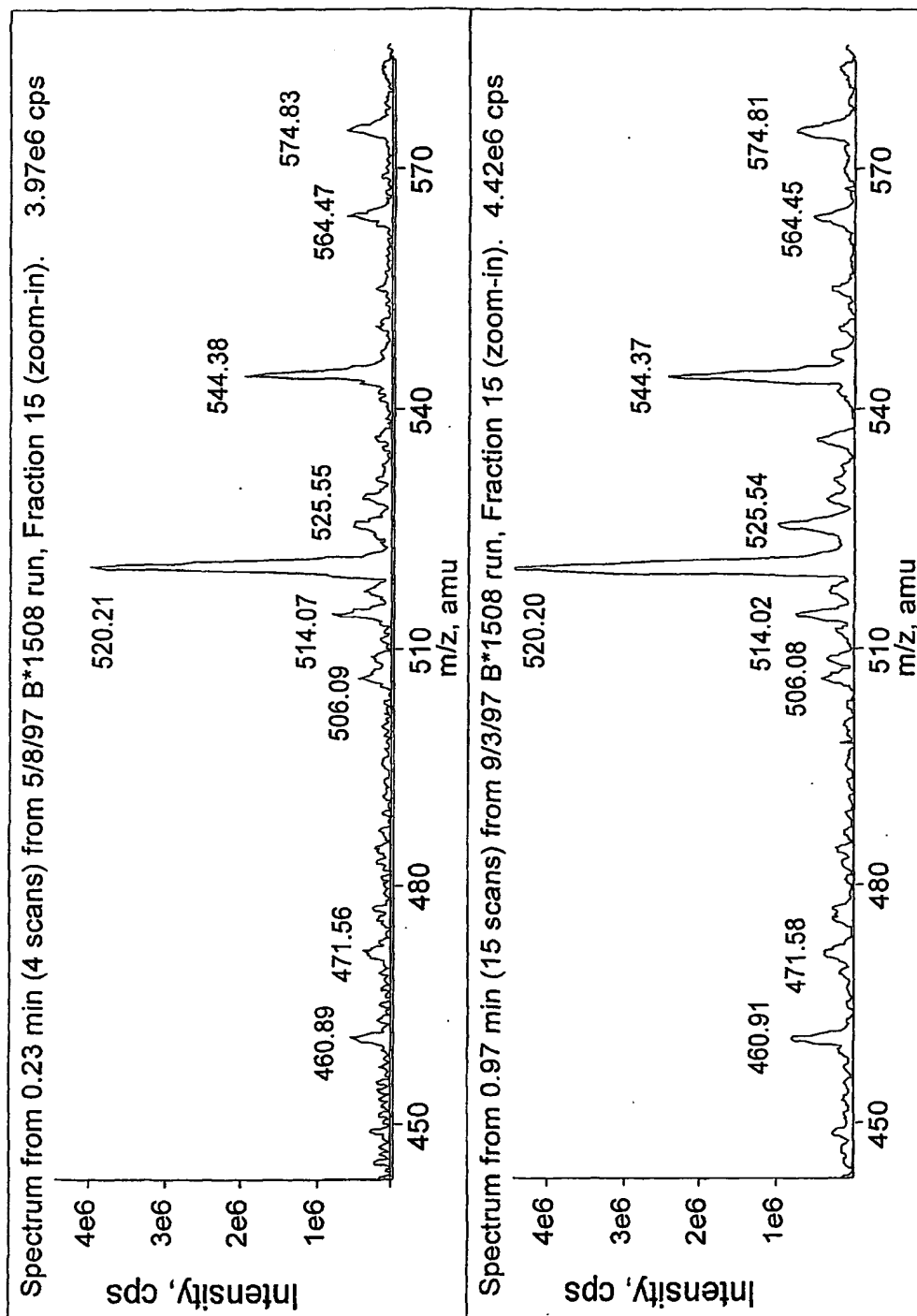
Chart C

Fig. 13 5 of 5



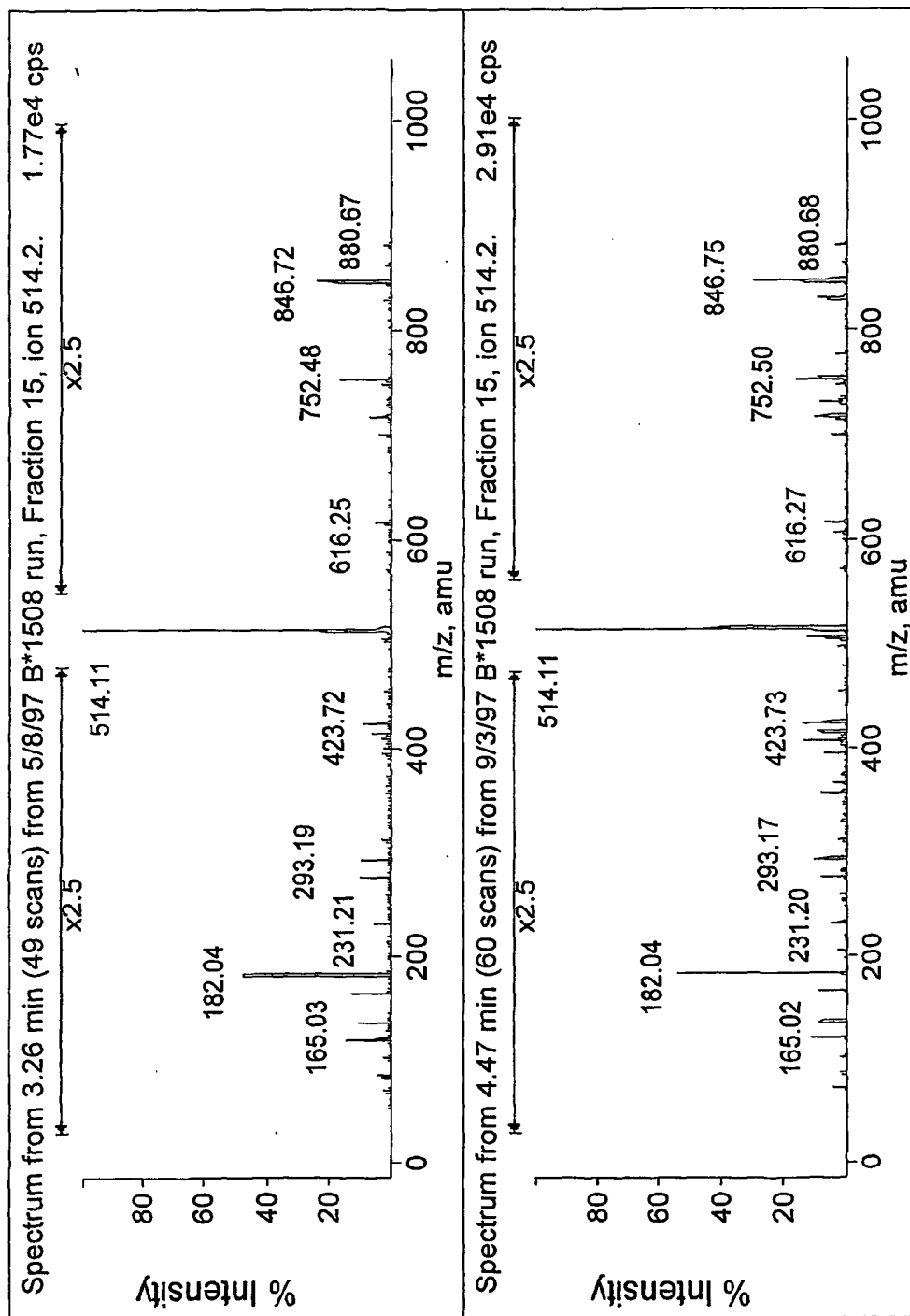
Graph A

Fig. 14 1 of 3



Graph B

Fig. 14 2 of 3



Graph C

Fig. 14 3 of 3

A (W6/32-purified B*1501 complexes)														
position:	1	2	3	4	5	6	7	8	9	10	11	12	13	14
dominant	-	Q _(8.0)	K _(8.0)	-	-	-	-	-	Y _(4.0)	-	-	-	-	-
			F _(5.0)											
			R _(4.0)											
strong	-	M _(3.0)	Y _(3.0)	P _(2.5)	G _(2.0)	-	-	-	F _(3.0)	-	-	-	-	-
		L _(2.0)	P _(2.0)	D _(2.5)										
		V _(2.0)	N _(2.0)	G _(2.0)										
			H _(2.0)	E _(2.0)										
B (BBM.1-purified B*1501 complexes)														
position:	1	2	3	4	5	6	7	8	9	10	11	12	13	14
dominant	-	Q _(7.5)	-	-	-	-	-	-	Y _(4.0)	-	-	-	-	-
strong	-	P _(3.5)	F _(2.5)	P _(2.2)	I _(2.0)	-	-	-	F _(3.0)	-	-	-	-	-
		L _(2.5)	K _(2.5)	D _(2.0)										
		V _(2.0)	R _(2.0)	G _(2.0)										
			P _(2.0)											
			N _(2.0)											

Fig. 15

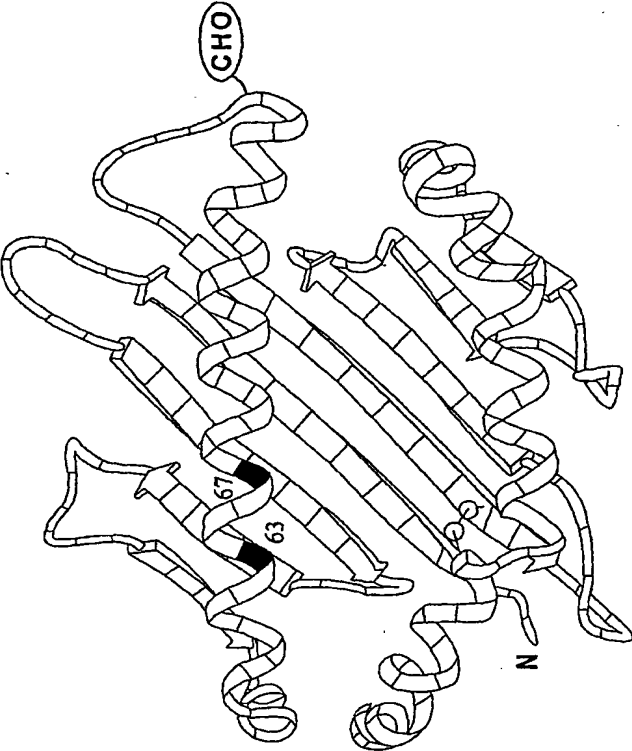


Fig. 16 1 of 5

B*1508									
	1	2	3	4	5	6	7	8	9
dominant	-	P	N	-	-	-	-	-	Y
		A	F	K	R	H	Y	I	
strong	-	-	-	E	H	R	-	-	F
				D	G	S			

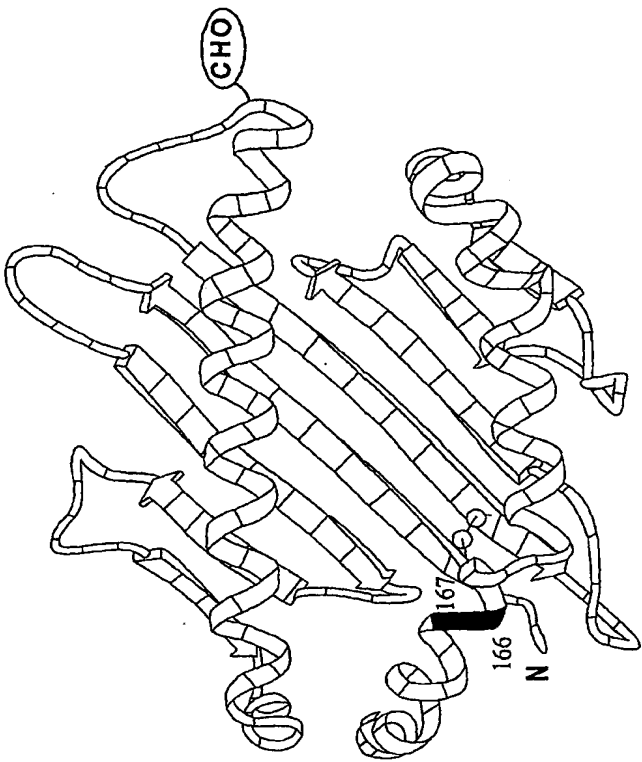


Fig. 16 2 of 5

B*1512

	1	2	3	4	5	6	7	8	9
dominant	-	Q	K	P	-	-	-	-	Y
			F	E					F
			R						
			H						
			V						
			N						
strong	-	L	I	D	T	-	-	-	Y
		M	A	G					

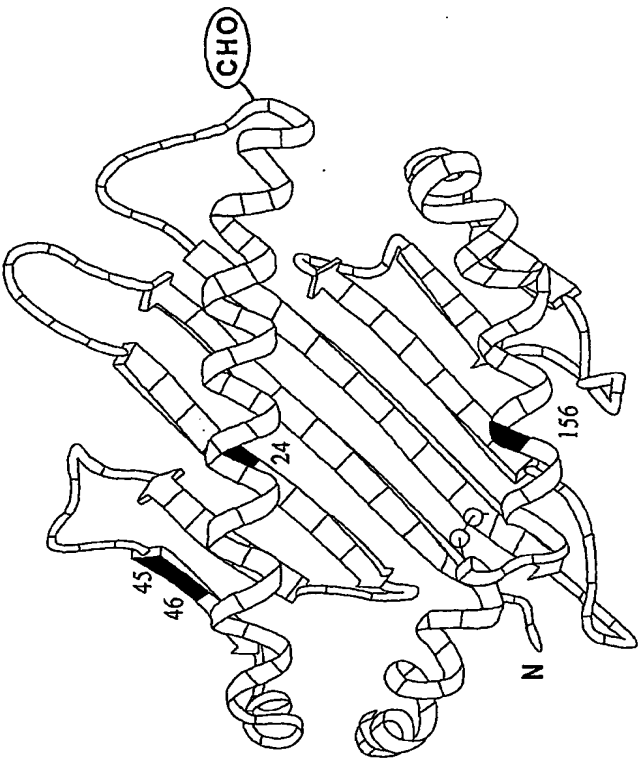


Fig. 16 3 of 5

B*1503									
	1	2	3	4	5	6	7	8	9
dominant	-	Q	F	P	-	-	-	-	Y
		K	Y						
			R	V	I	N	A		
strong	-	M	H	G	-	R	-	-	F
			L	E		V			
			P	D					

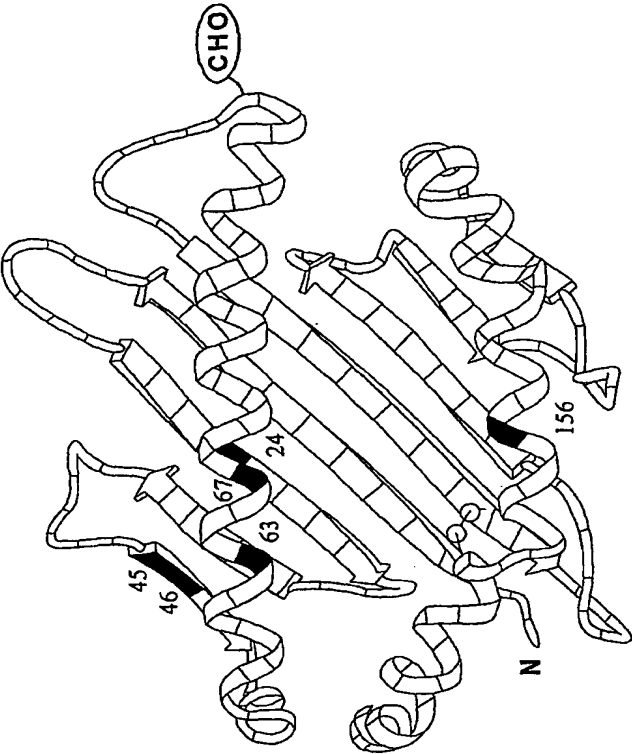


Fig. 16 4 of 5

B*1518

	1	2	3	4	5	6	7	8	9
dominant	-	H	F	-	-	-	-	-	Y
strong	-	-	I	P	-	V	I	-	F
			M	E					
			H						
			K						
			W						

B*1510

	1	2	3	4	5	6	7	8	9
dominant	-	H	Y	P	-	-	-	-	L
		A	I	K	S	F	L	V	T
strong	-	-	G	R	R	I	M	-	F
			D	D	V		L		
			E	K					
			M						
									N

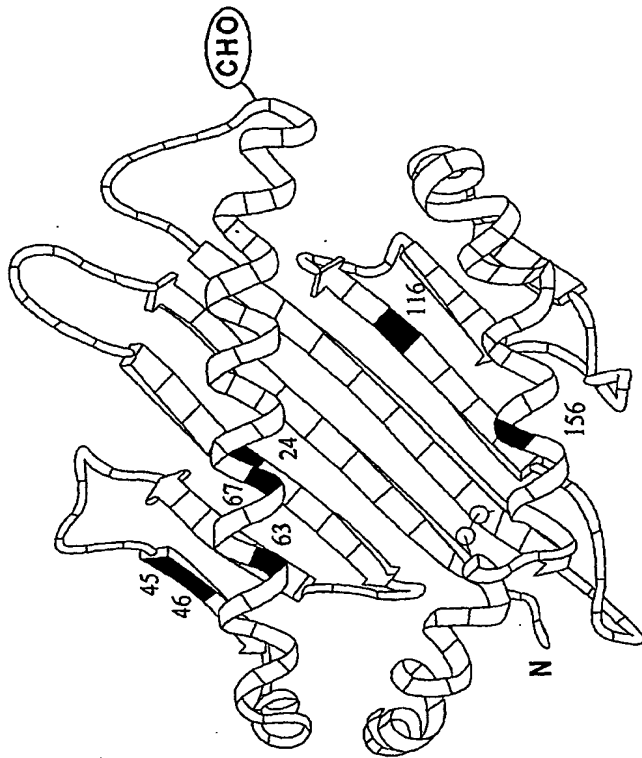


Fig. 16 5 of 5

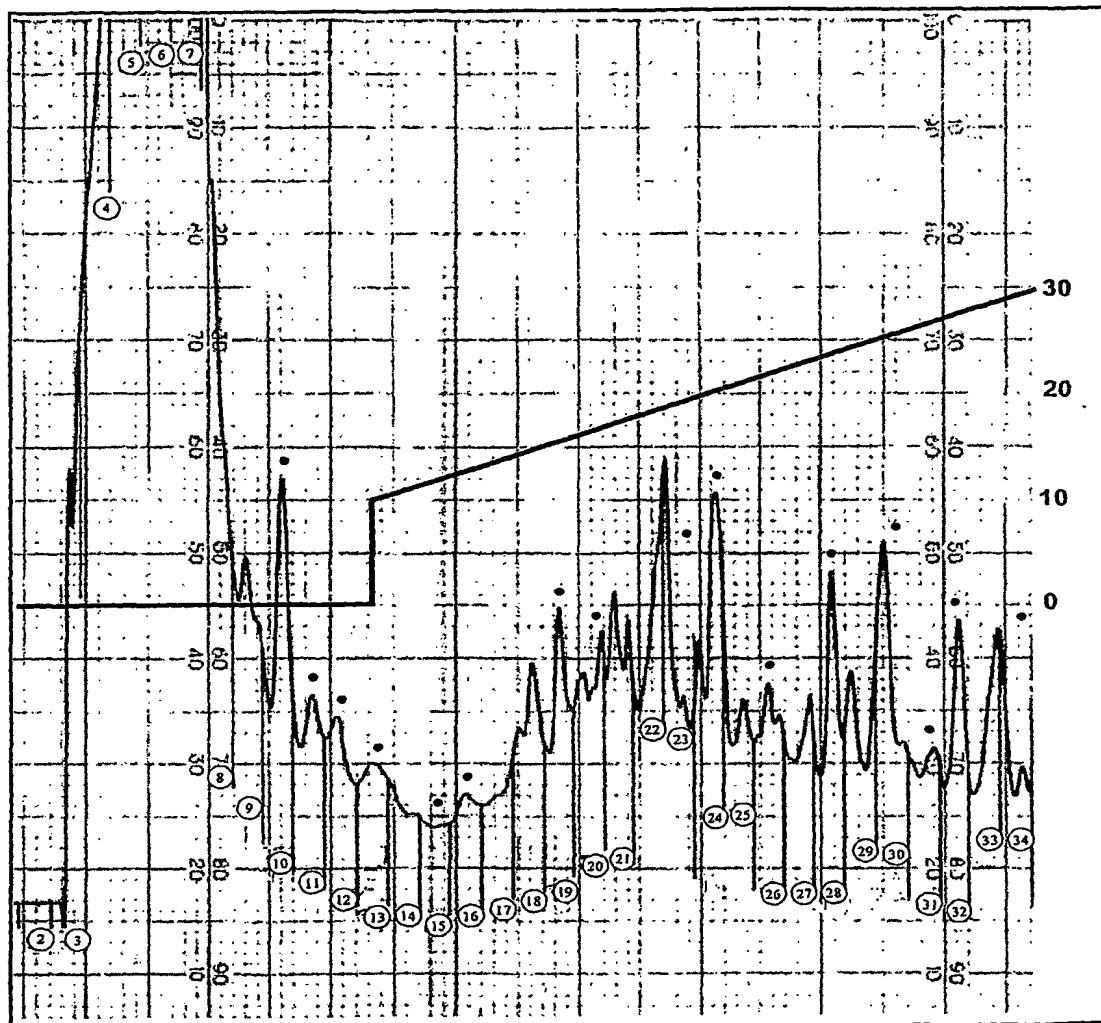
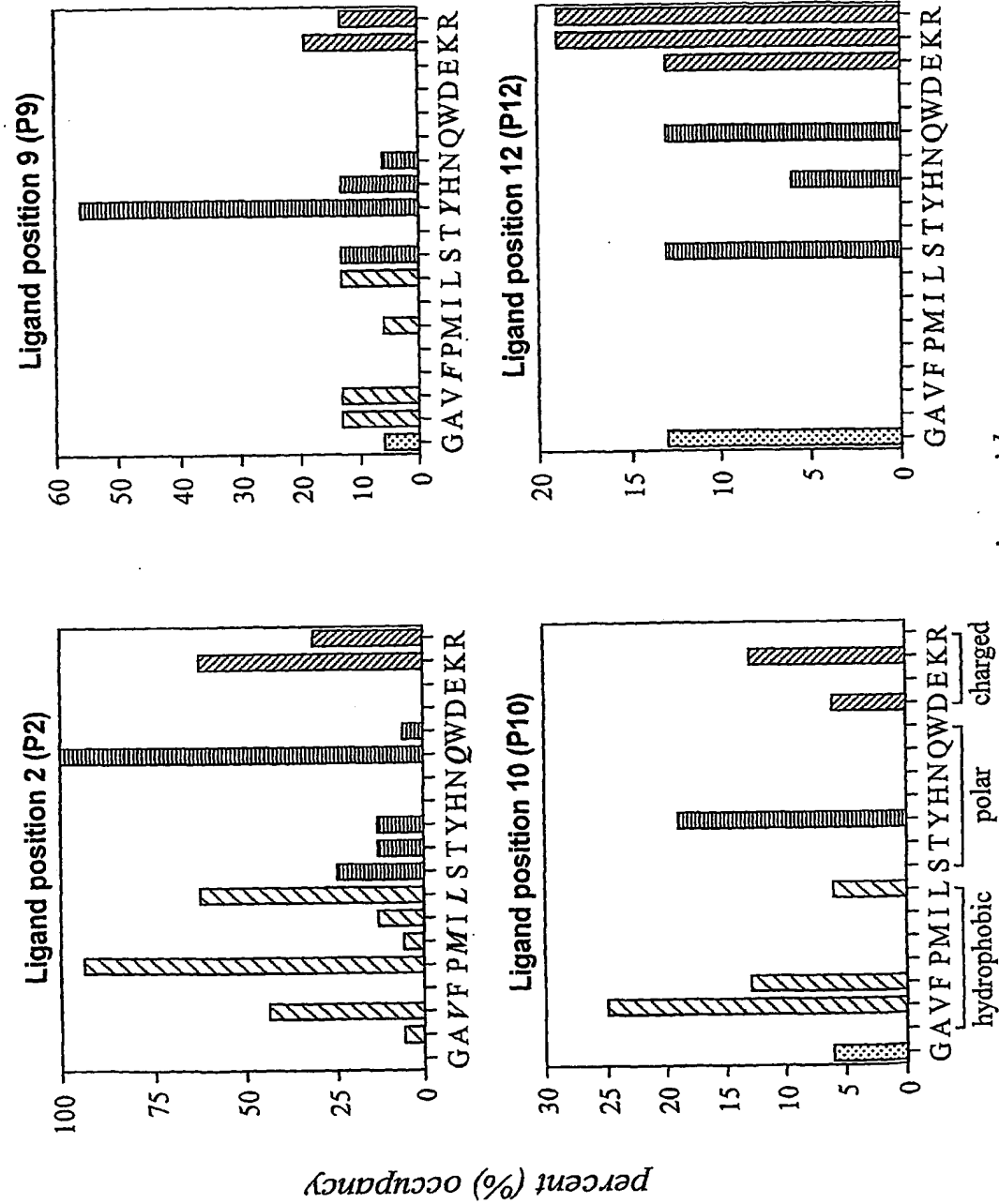


Fig. 17



amino acid
Fig. 18

Fraction 10												
Position:	2	3	4	5	6	7	8	9	10	11	12	
<i>Dominant</i>	P	-	-	-	-	-	-	-	-	-	-	-
	Q											
<i>strong</i>	W	K	G	-	-	-	-	-	-	-	-	-
		R	F									
		H										
<i>weak</i>	S	E	D	S	H	V	I	Y	K	V	E	
	I		N	W	S	T						
			M	P								
			I									

Fraction 15												
Position:	2	3	4	5	6	7	8	9	10	11	12	
<i>Dominant</i>	P	-	-	-	-	-	-	-	-	-	-	-
	Q											
<i>strong</i>	W	K	G	-	-	-	-	-	-	-	-	-
		R	F									
		H										
<i>weak</i>	S	E	D	S	H	V	I	Y	K	V	E	
	I		N	W	S	T						
			M	P								
			I									

Fraction 28												
Position:	2	3	4	5	6	7	8	9	10	11	12	
<i>Dominant</i>	K	-	-	-	-	-	-	-	-	-	-	-
<i>strong</i>	Q	H	G	L	R	I	D	K	V	E	-	
	V	N		S			N					
	P	R		H								
<i>weak</i>	-	P	E	P	V	P	Q	Y	-	F	S	
		F	E	E	A	M	E	F	-	P	K	
		K			F		A	H				
		D					W					

Fraction 31										
Position:	2	3	4	5	6	7	8	9	10	
<i>Dominant</i>	-	K	-	W	-	-	-	-	-	-
<i>strong</i>	S	H	N	R	S	F	-	G	F	
	Q			M	Y	V				
	P									
<i>weak</i>	L	Y	A	-	L	-	Y	Y	-	
		L			K		S			
		V					K			

Fraction 38												
Position:	2	3	4	5	6	7	8	9	10	11	12	
<i>Dominant</i>	K	-	-	-	-	-	-	-	-	-	-	-
<i>strong</i>	Q	H	G	L	R	I	D	K	V	E	-	
	V	N		S			N					
	P	R		H								
<i>weak</i>	-	P	E	P	V	P	Q	Y	-	F	S	
		F	E	E	A	M	E	F	-	P	K	
		K			F		A	H				
		D					W					

Fig. 19

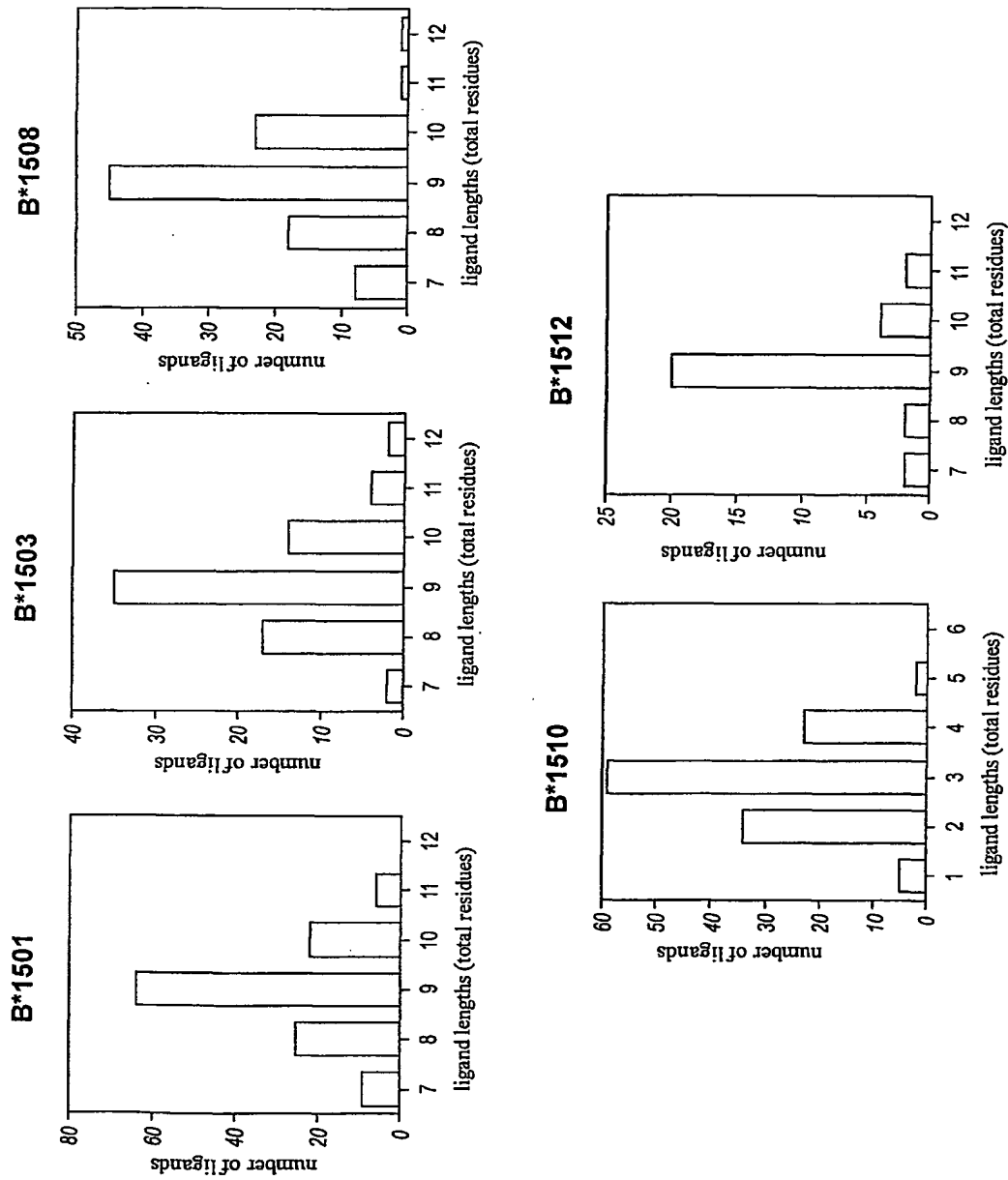


Fig. 20

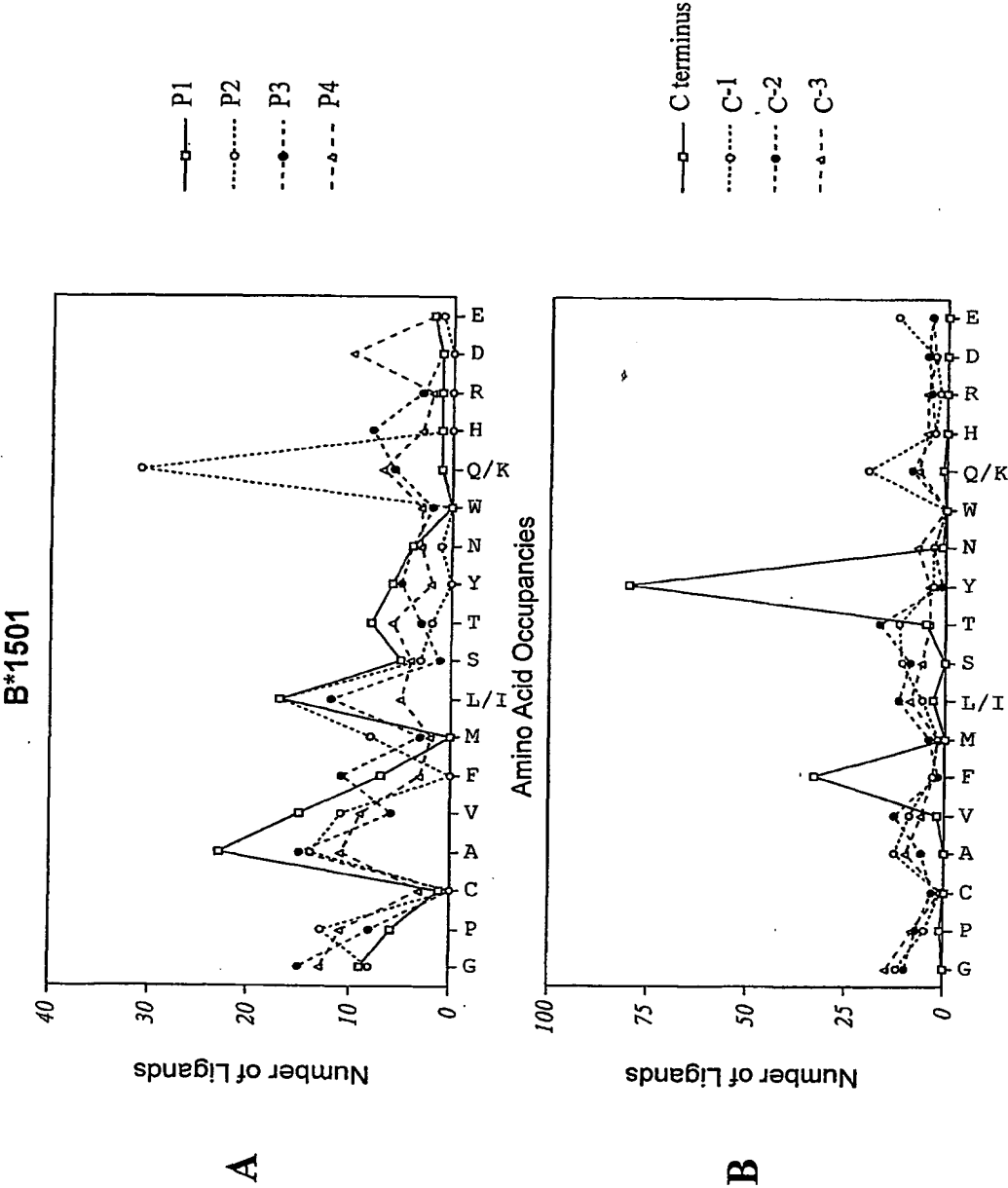
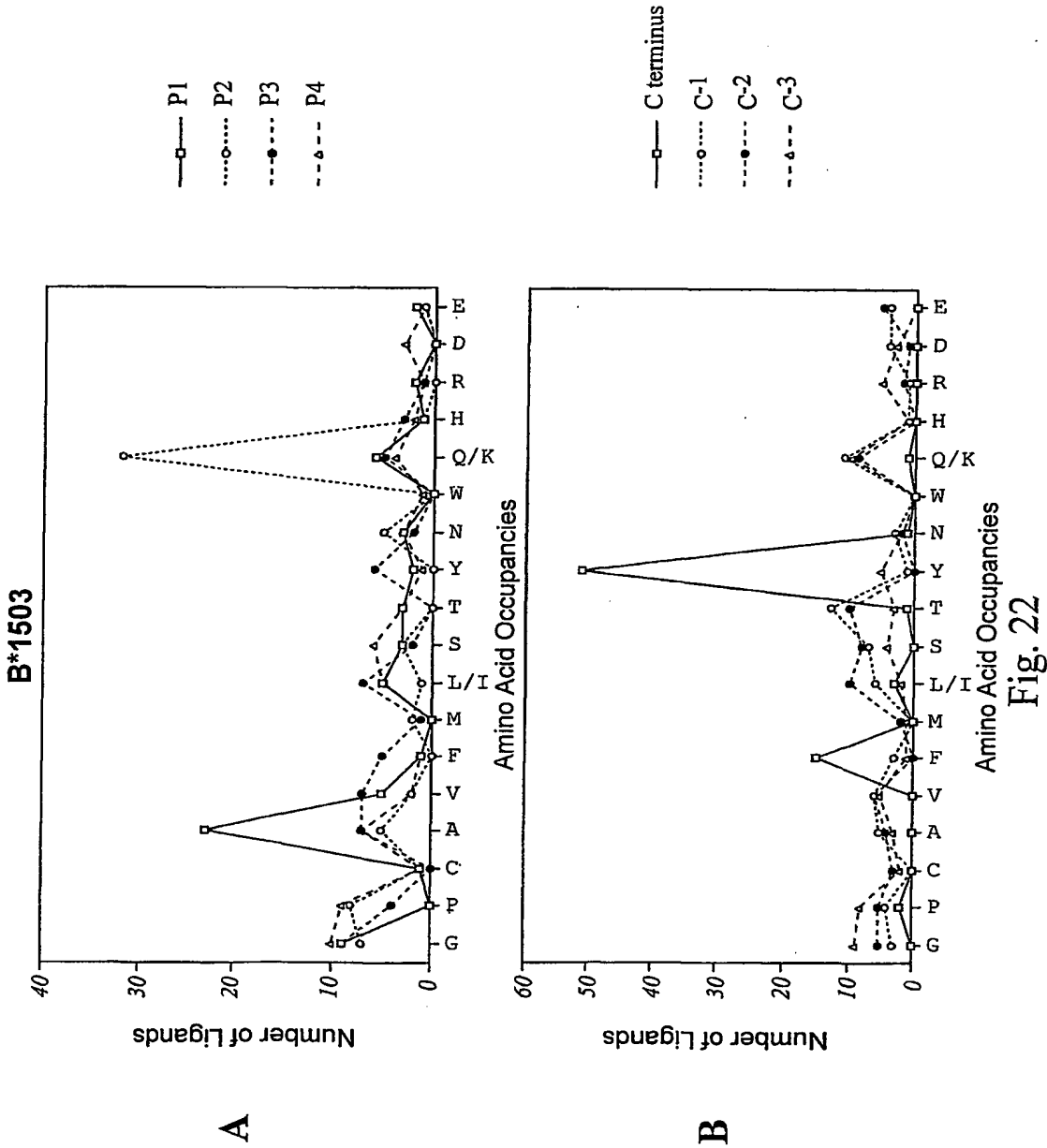


Fig. 21



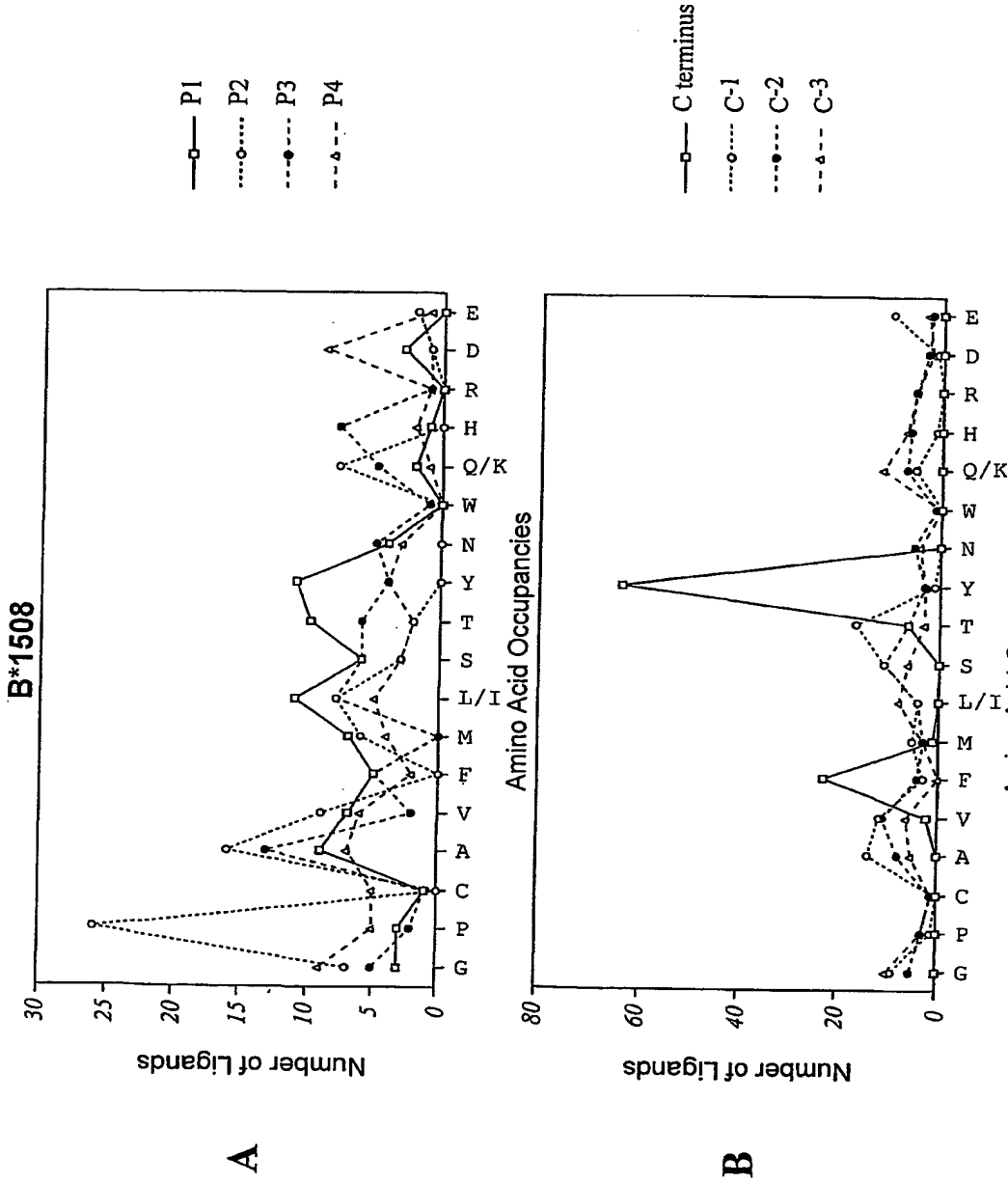


Fig. 23

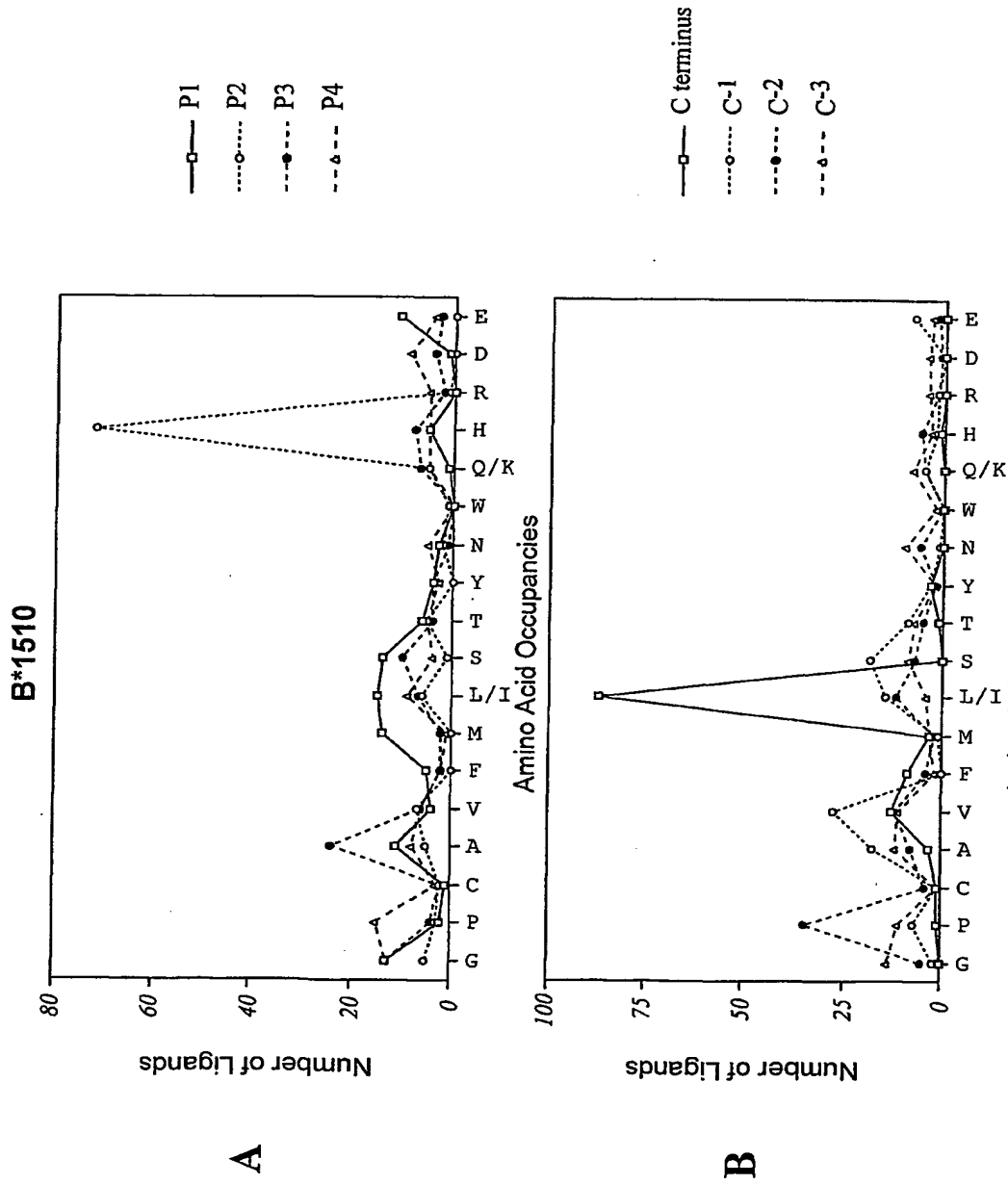


Fig. 24

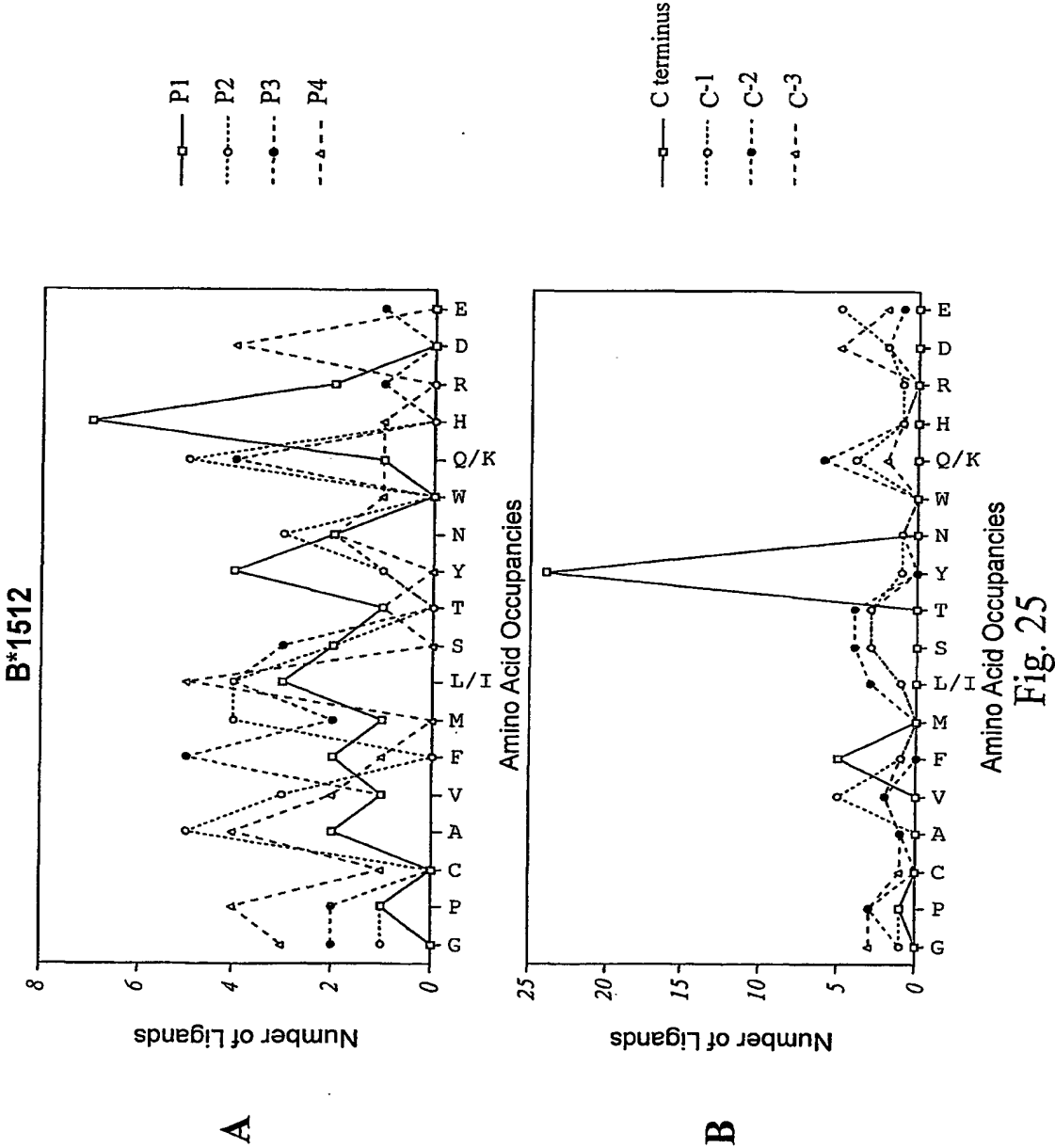


Fig. 25

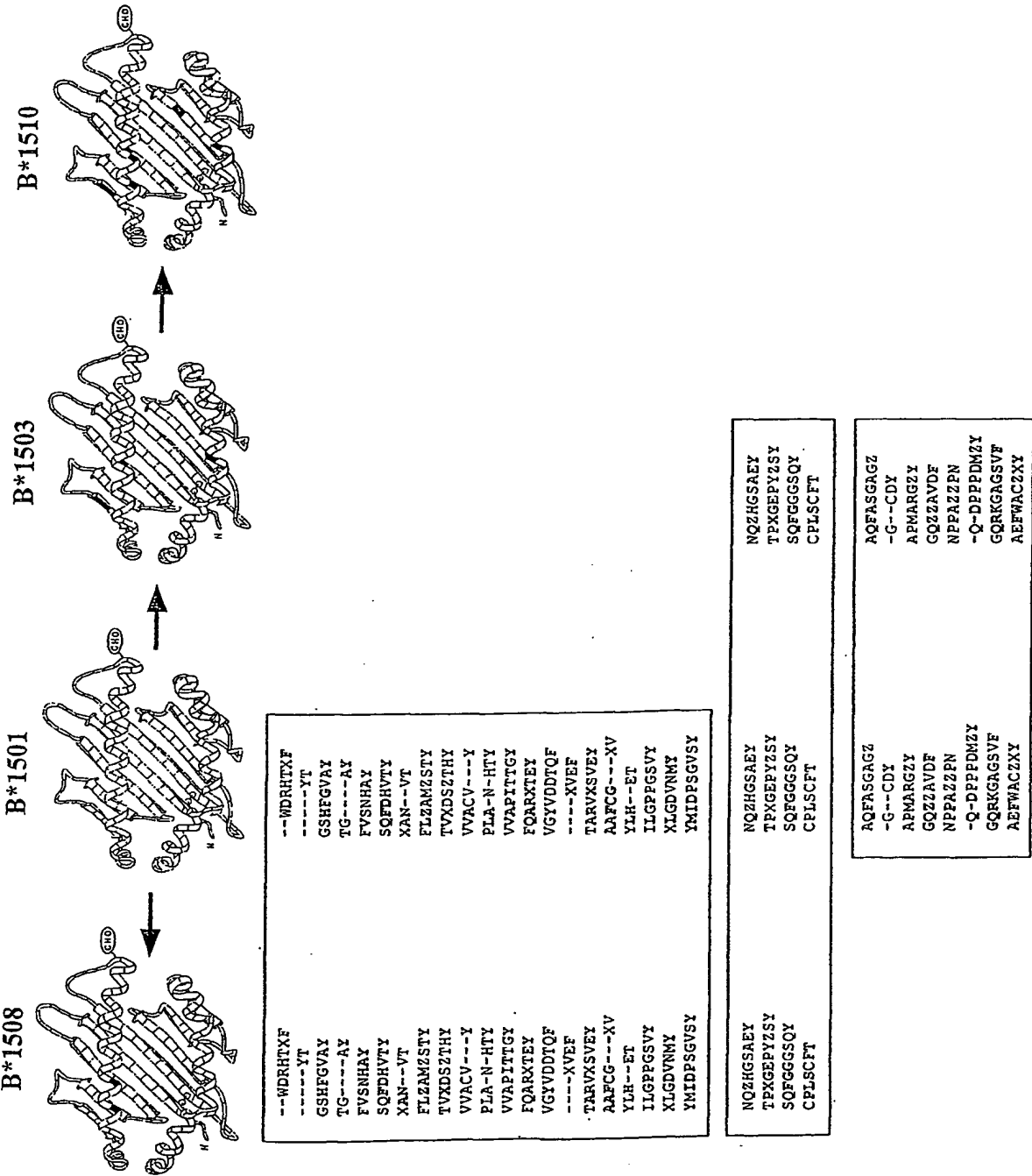


Fig. 26

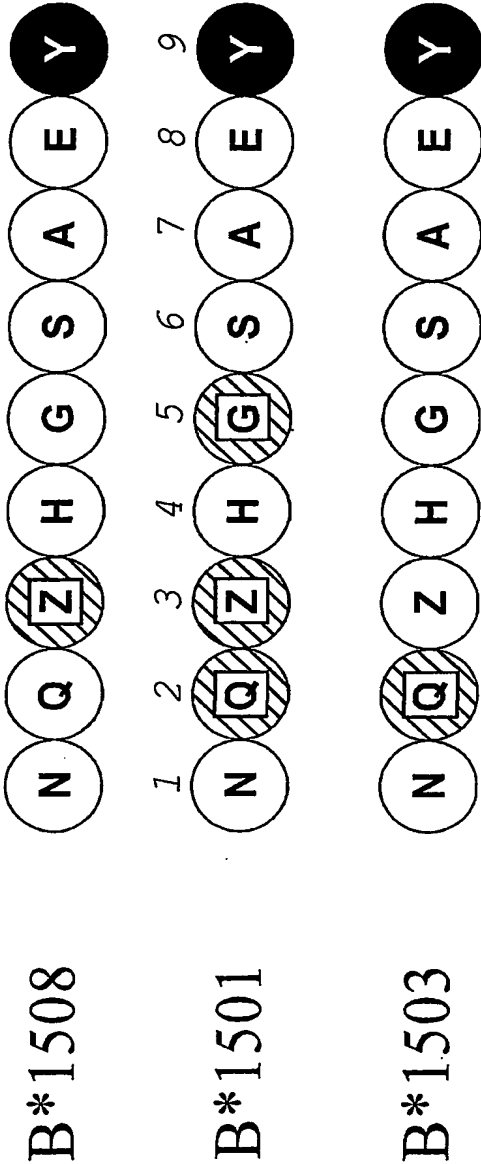


Fig. 27

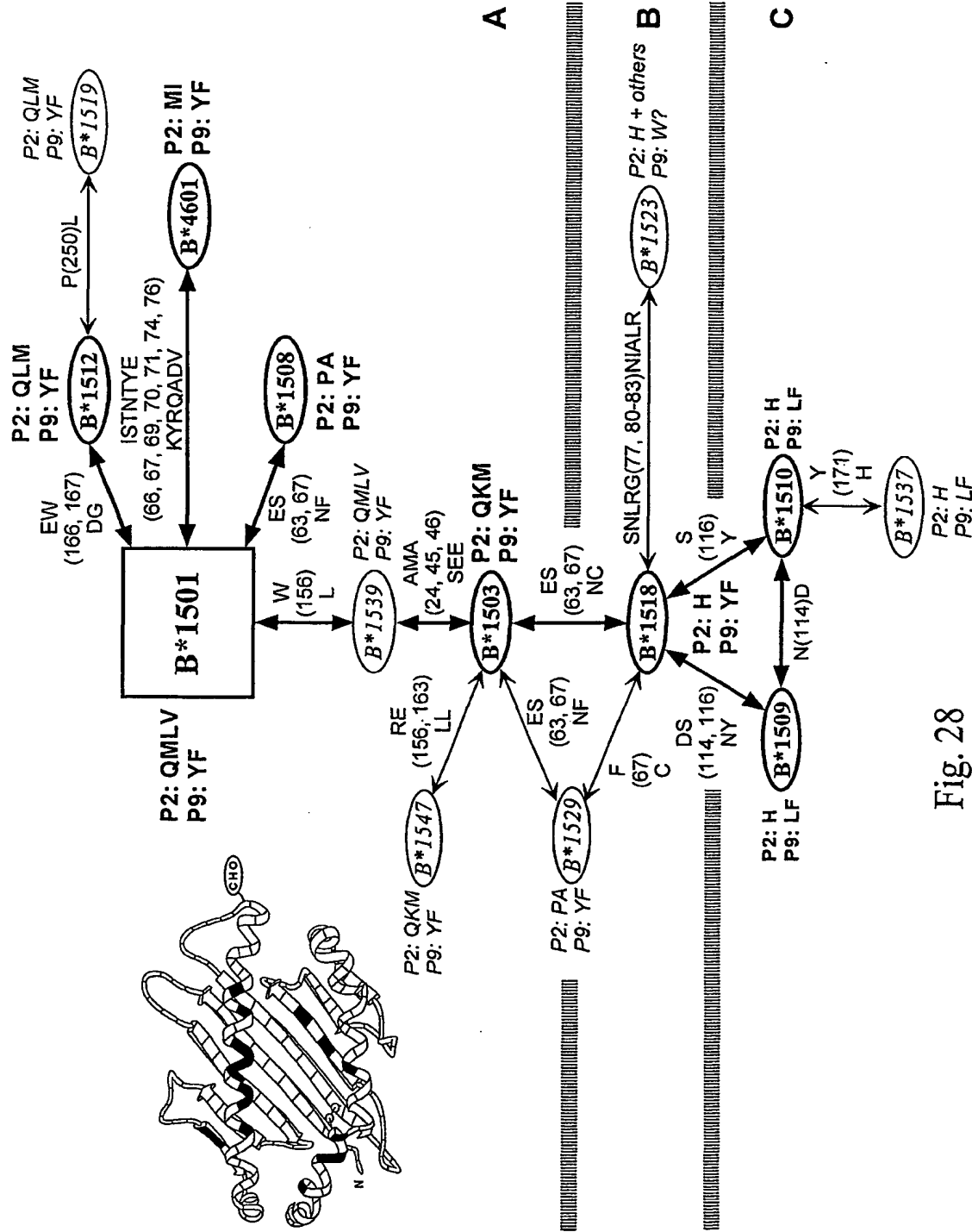
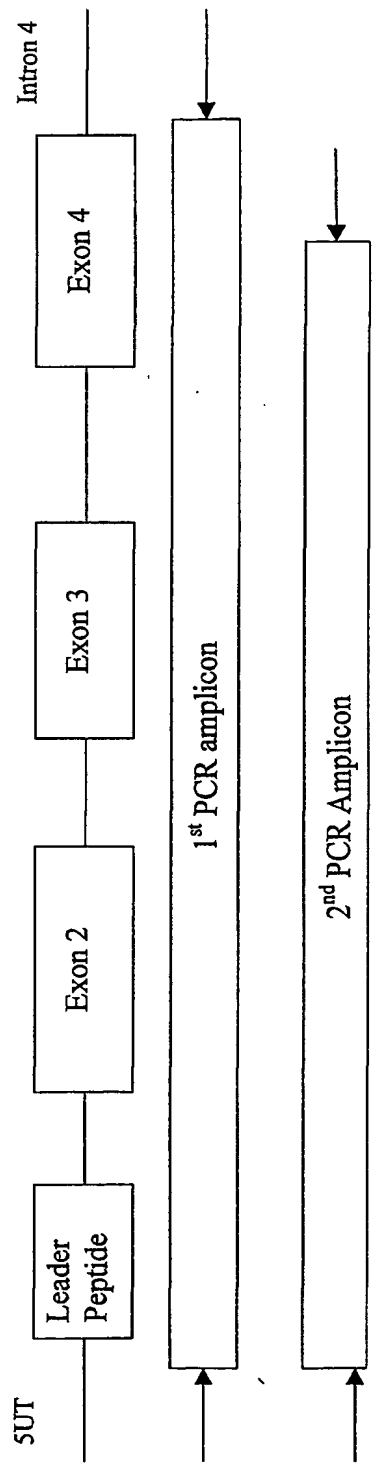


Fig. 28



PCR Strategy
Fig. 29



FIG. 30

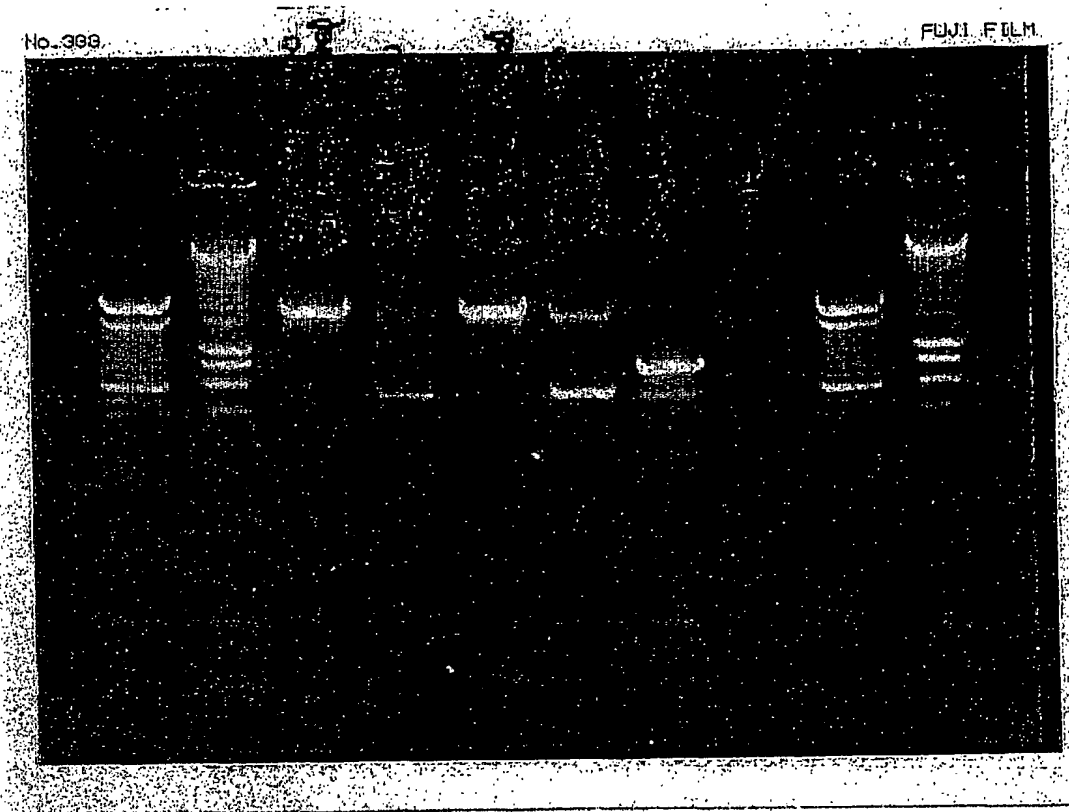


FIG. 31



FIG. 32

GDNA FIG. 5

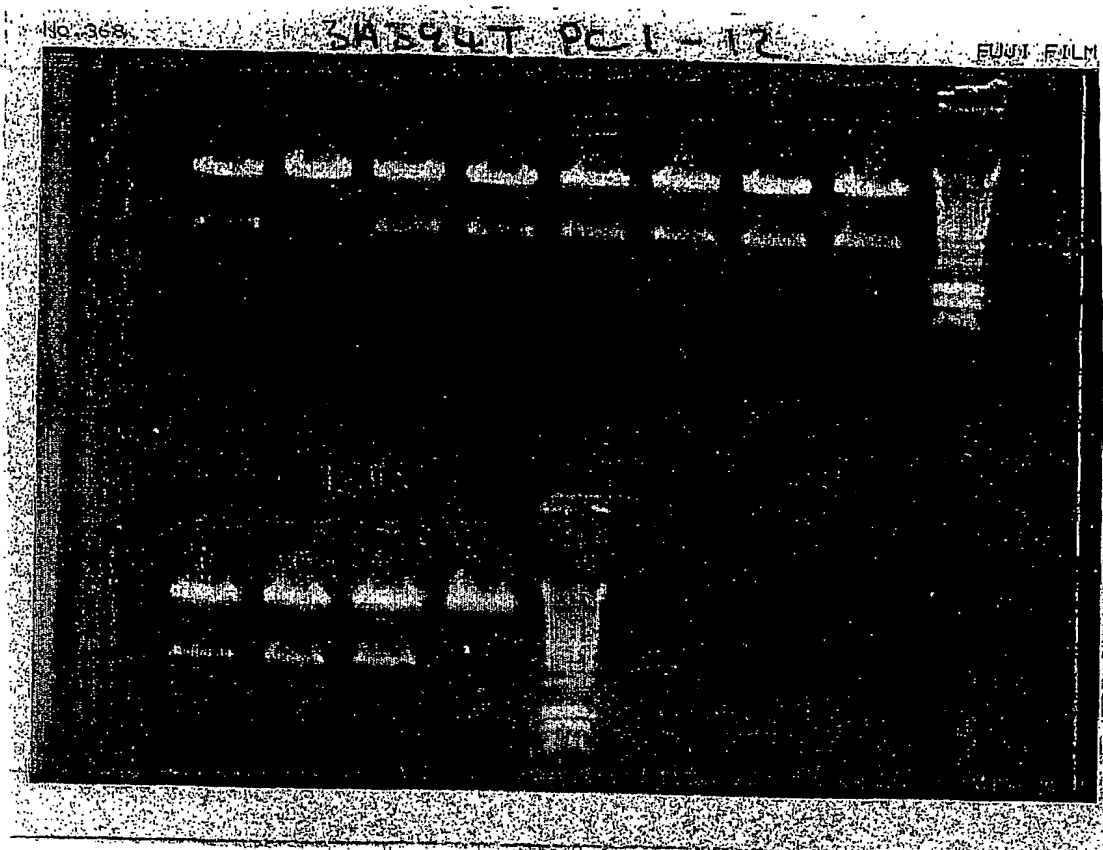


FIG. 33

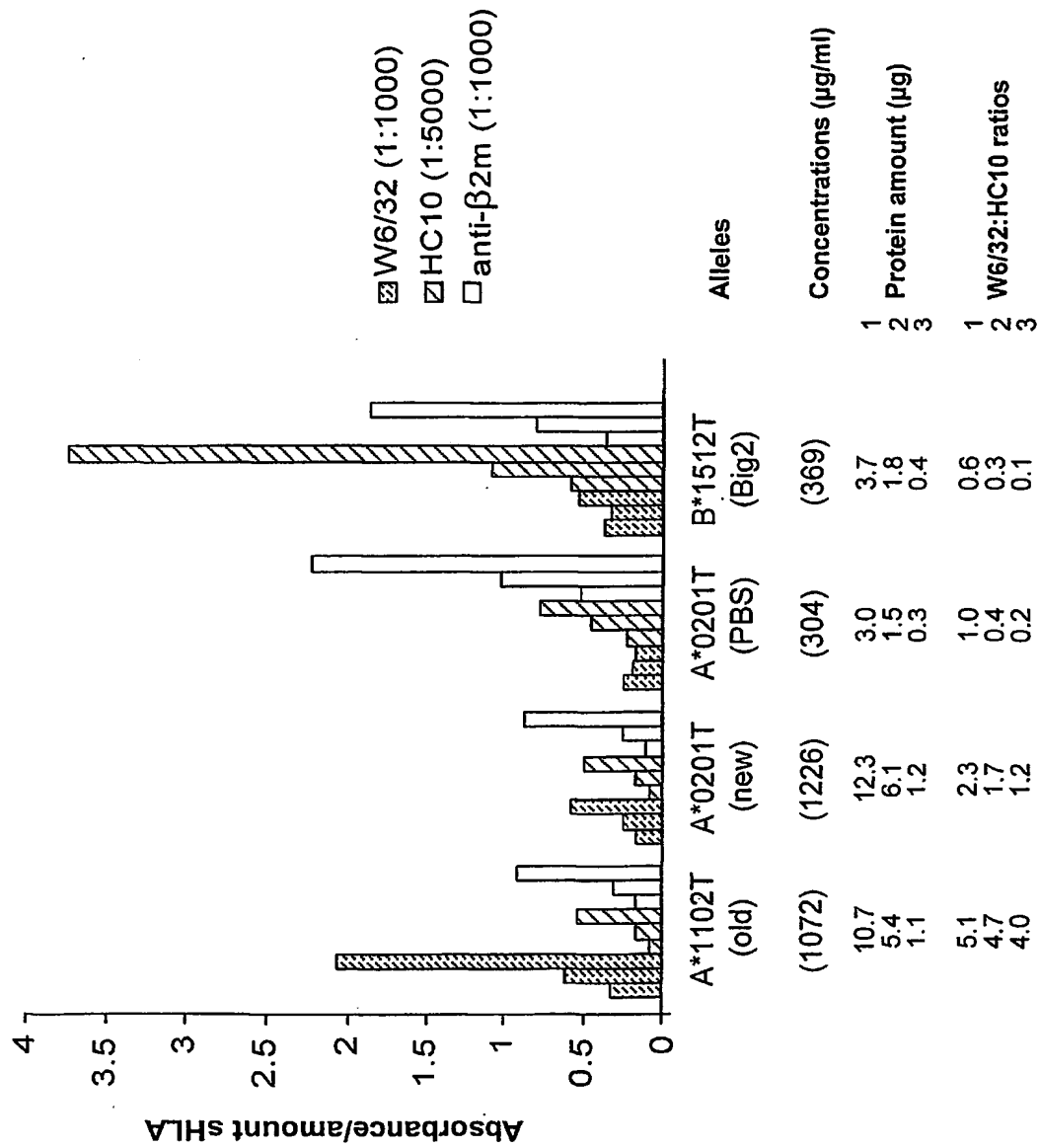


Fig. 34

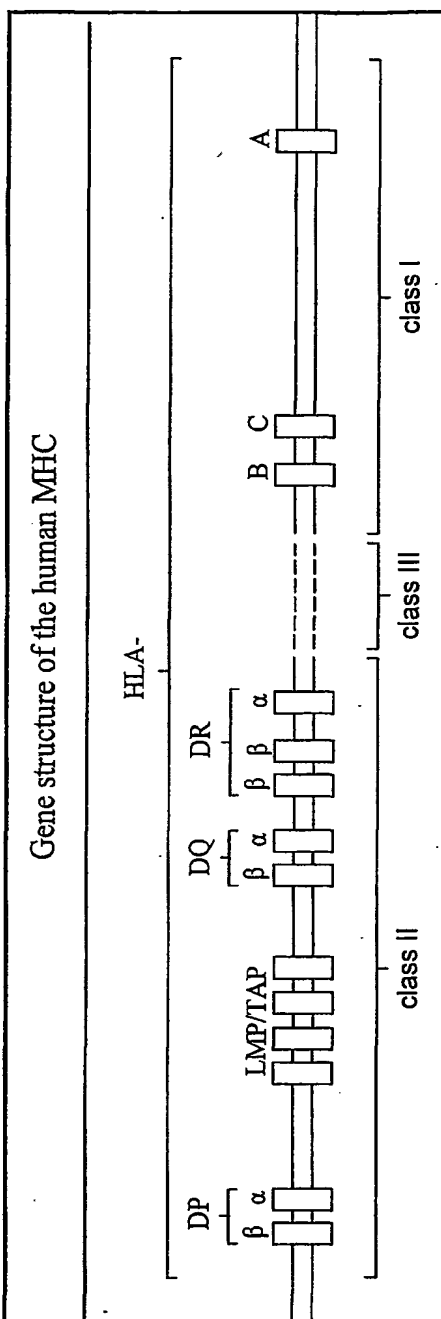


Fig. 35

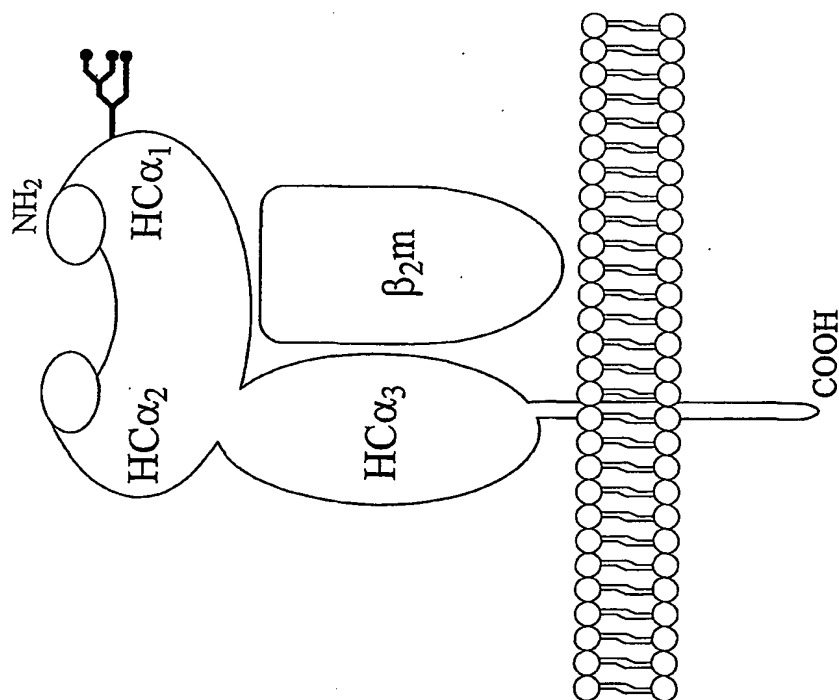


Fig. 36

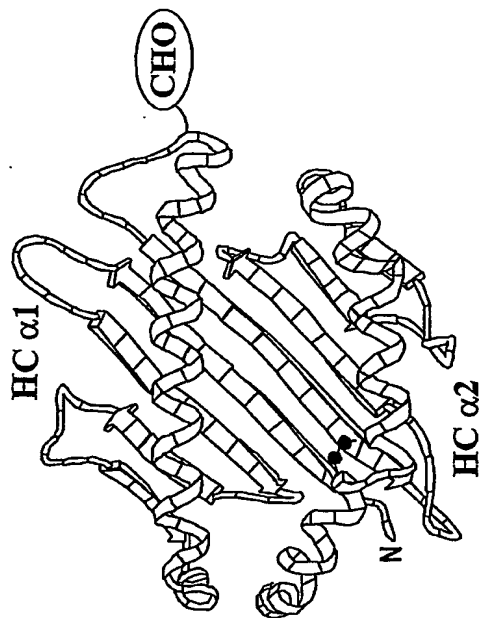


Fig. 37

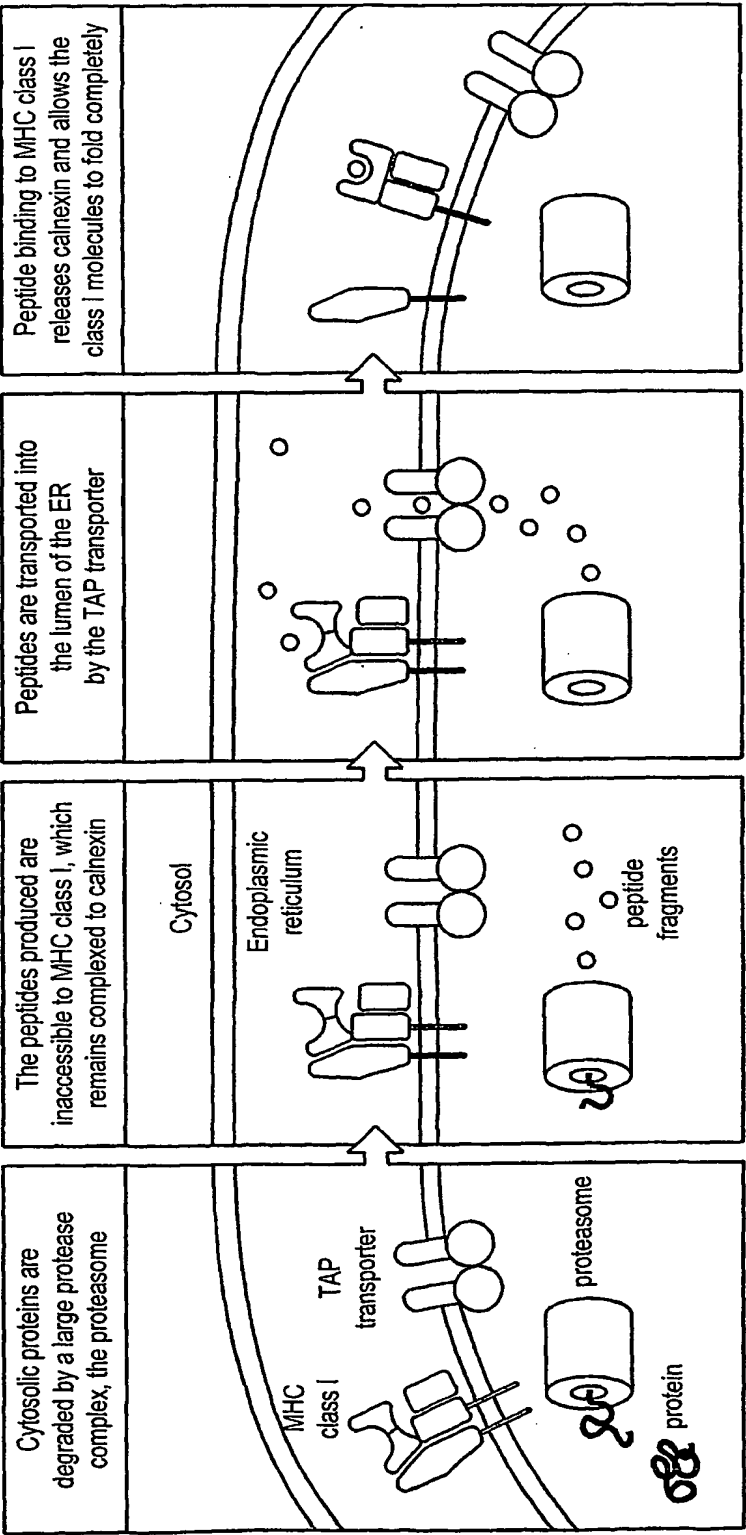


Fig. 38

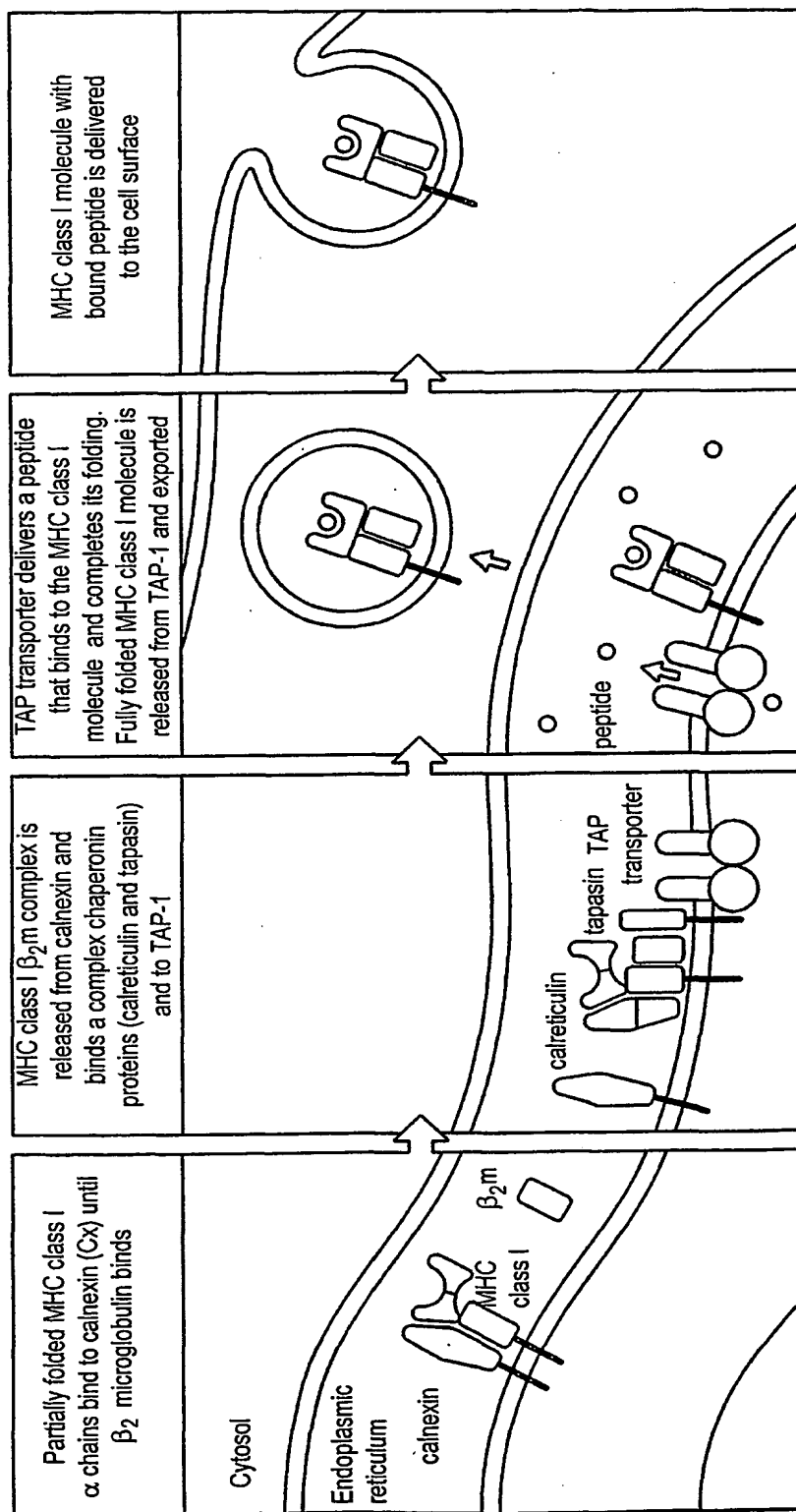


Fig. 39

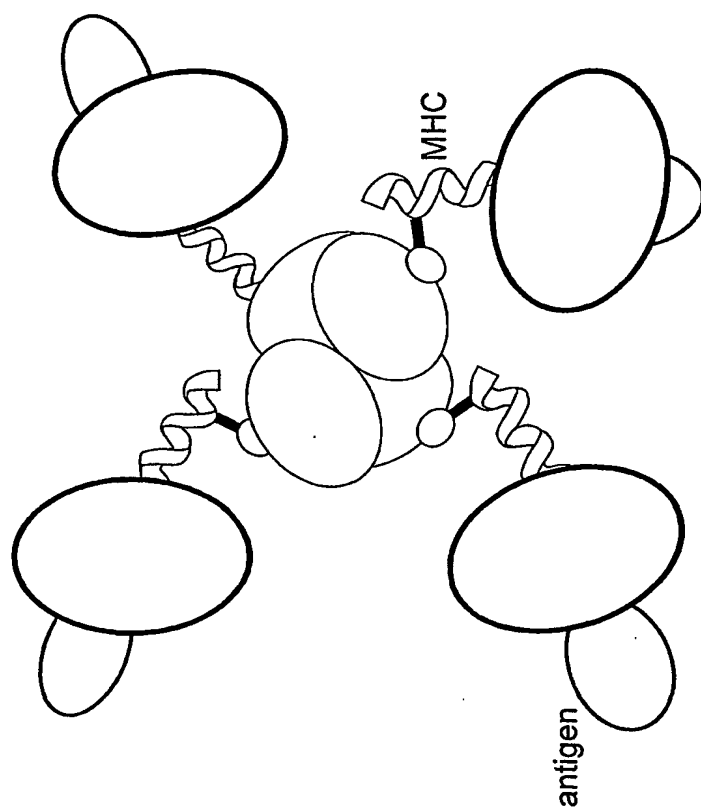


Fig. 40

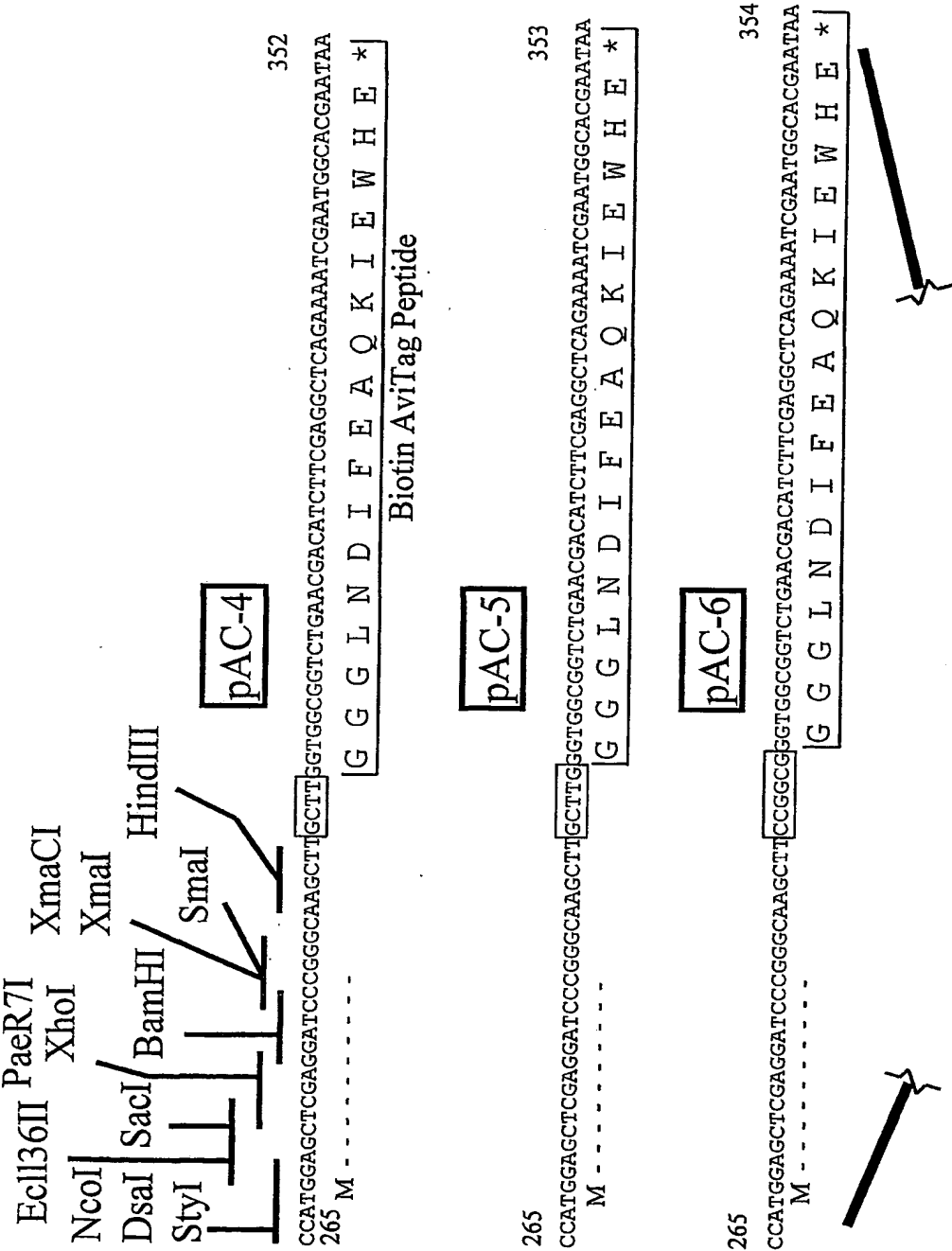


Fig. 41 1 of 3

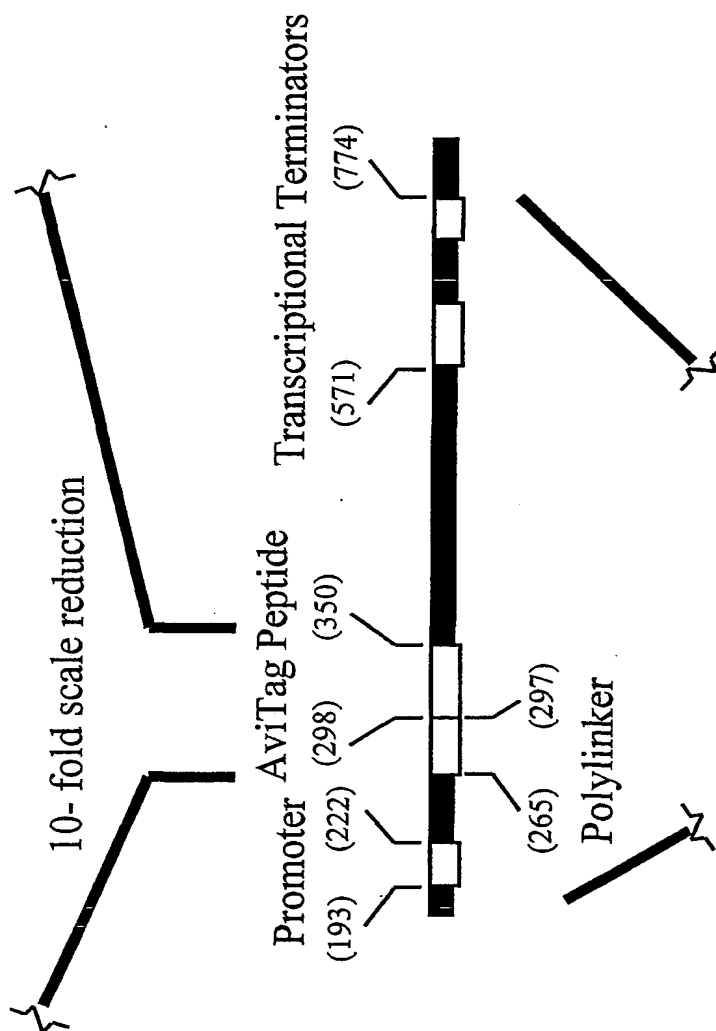


Fig. 41 2 of 3

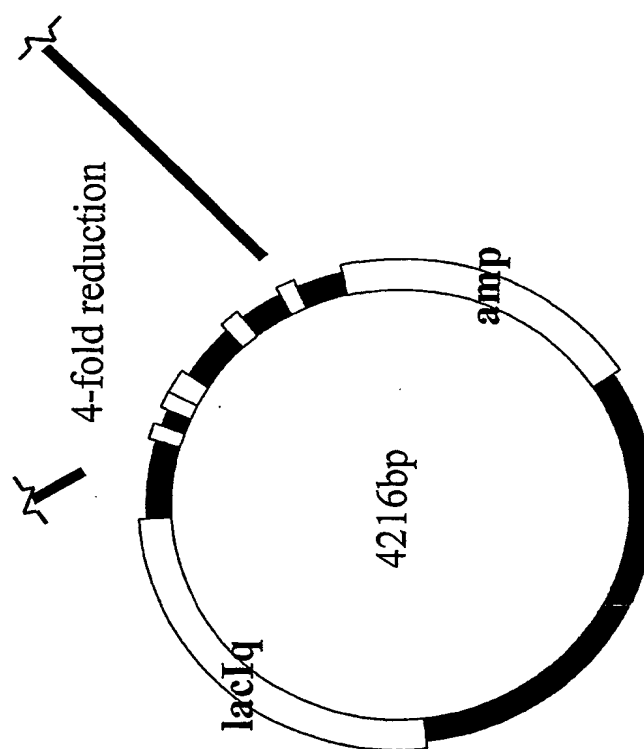


Fig. 41 3 of 3

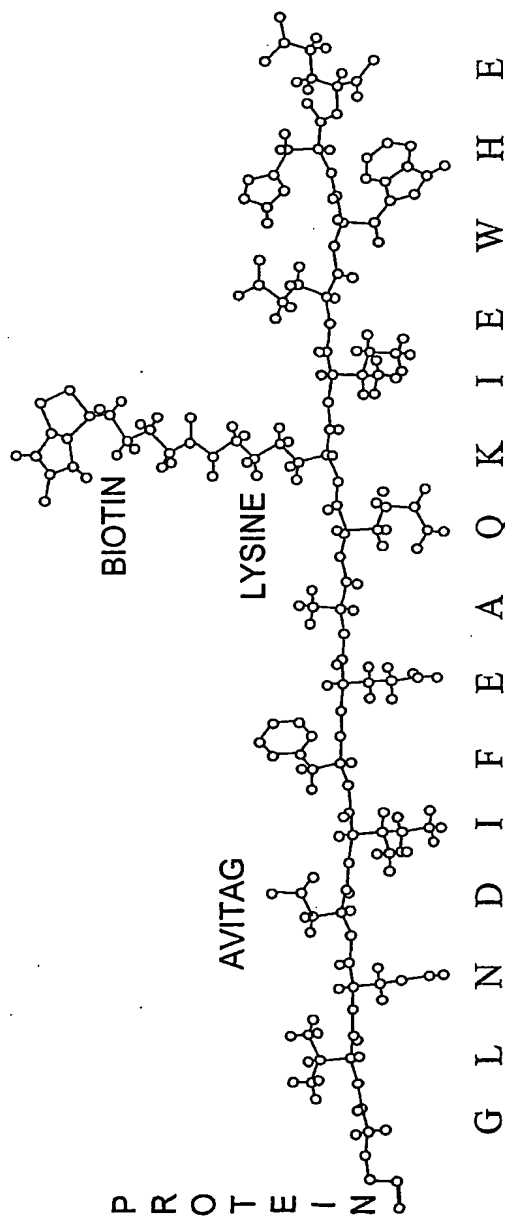


Fig. 42

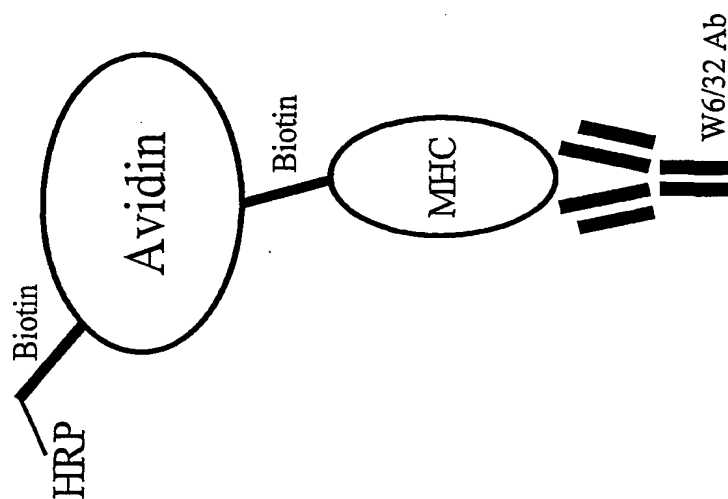


Fig. 43

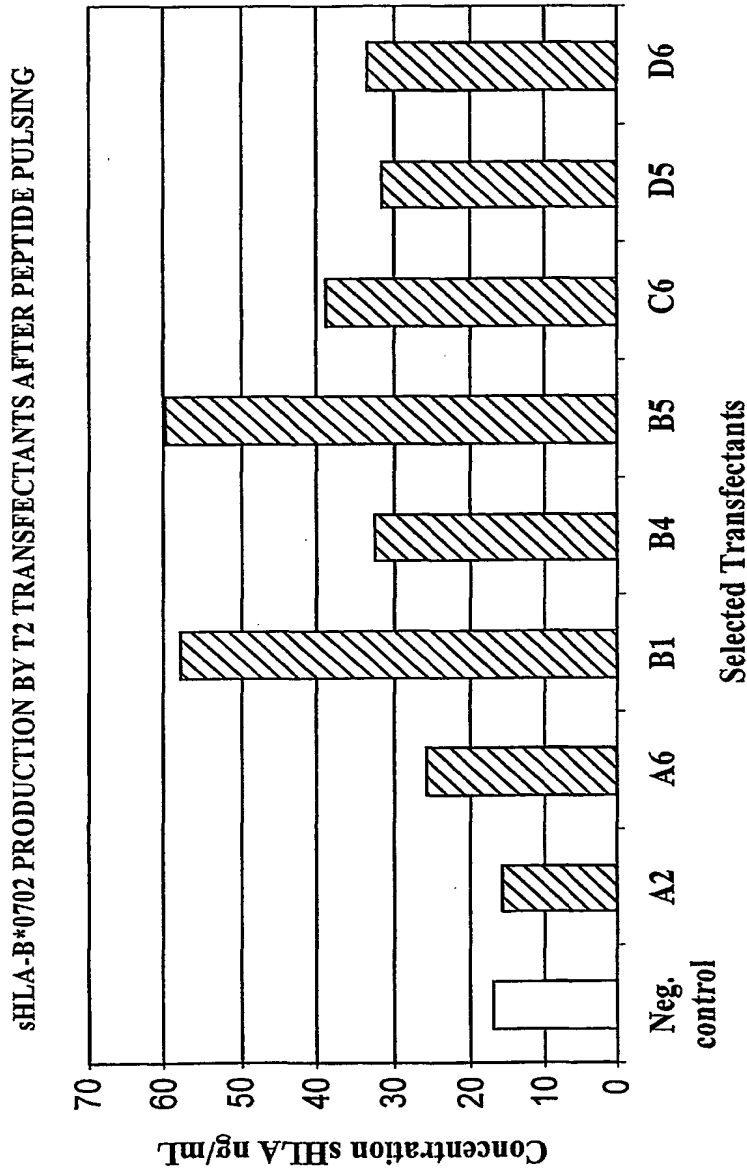


Fig. 44

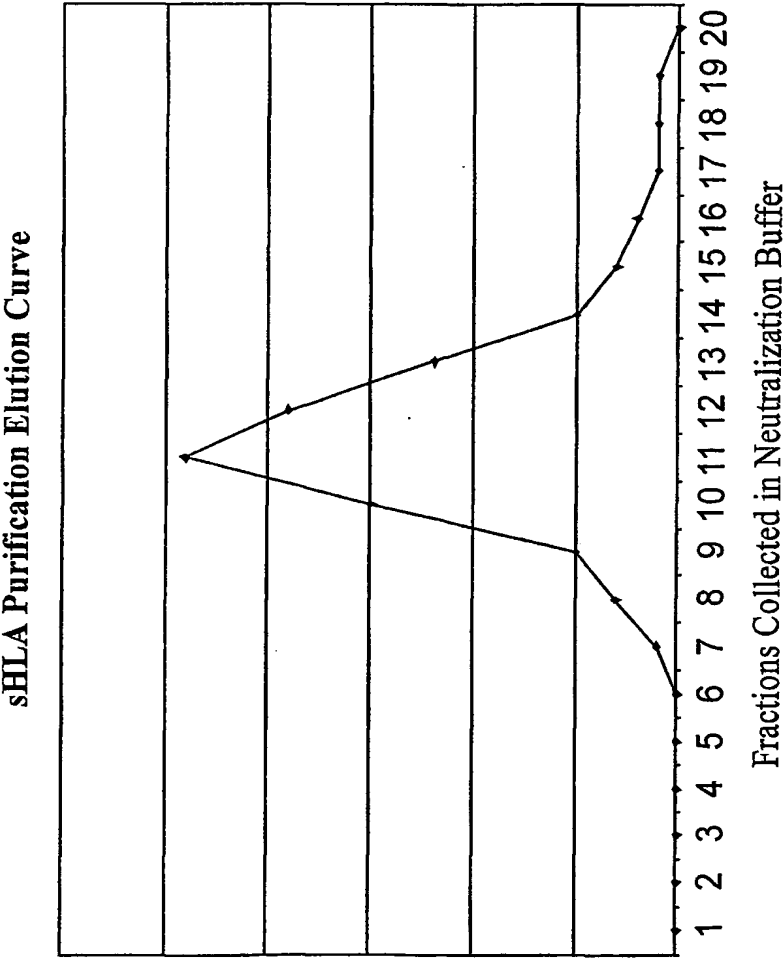


Fig. 45

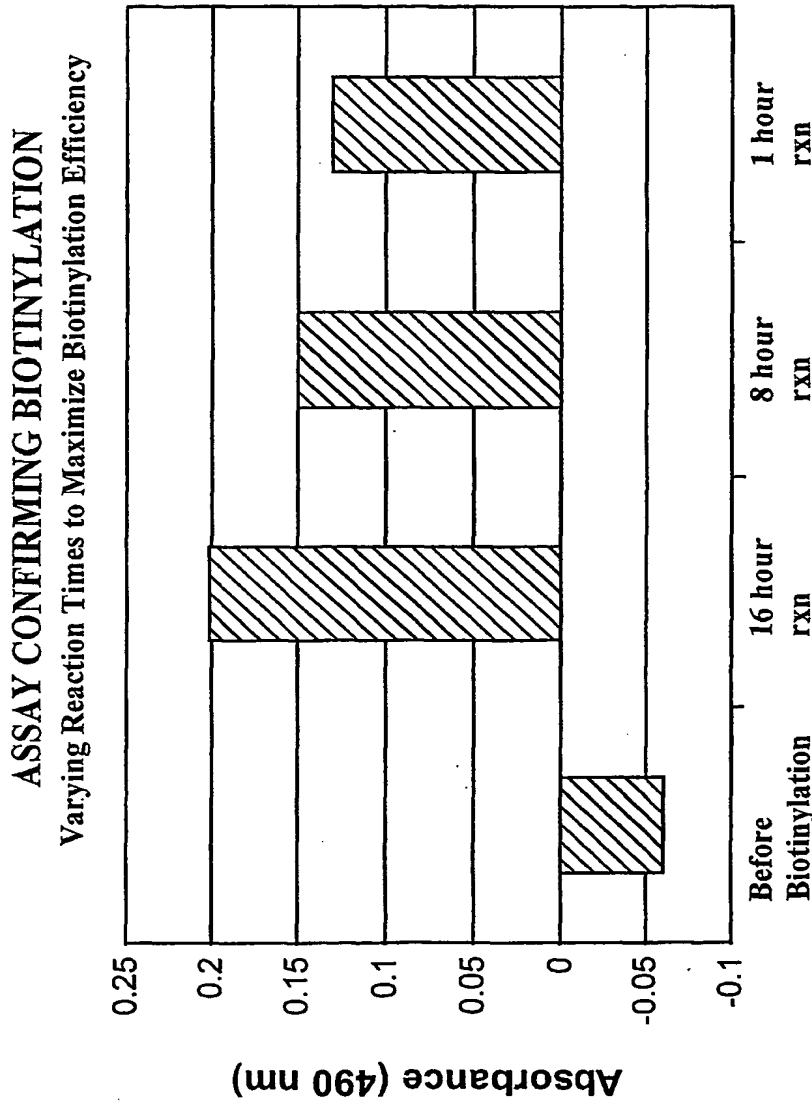


Fig. 46

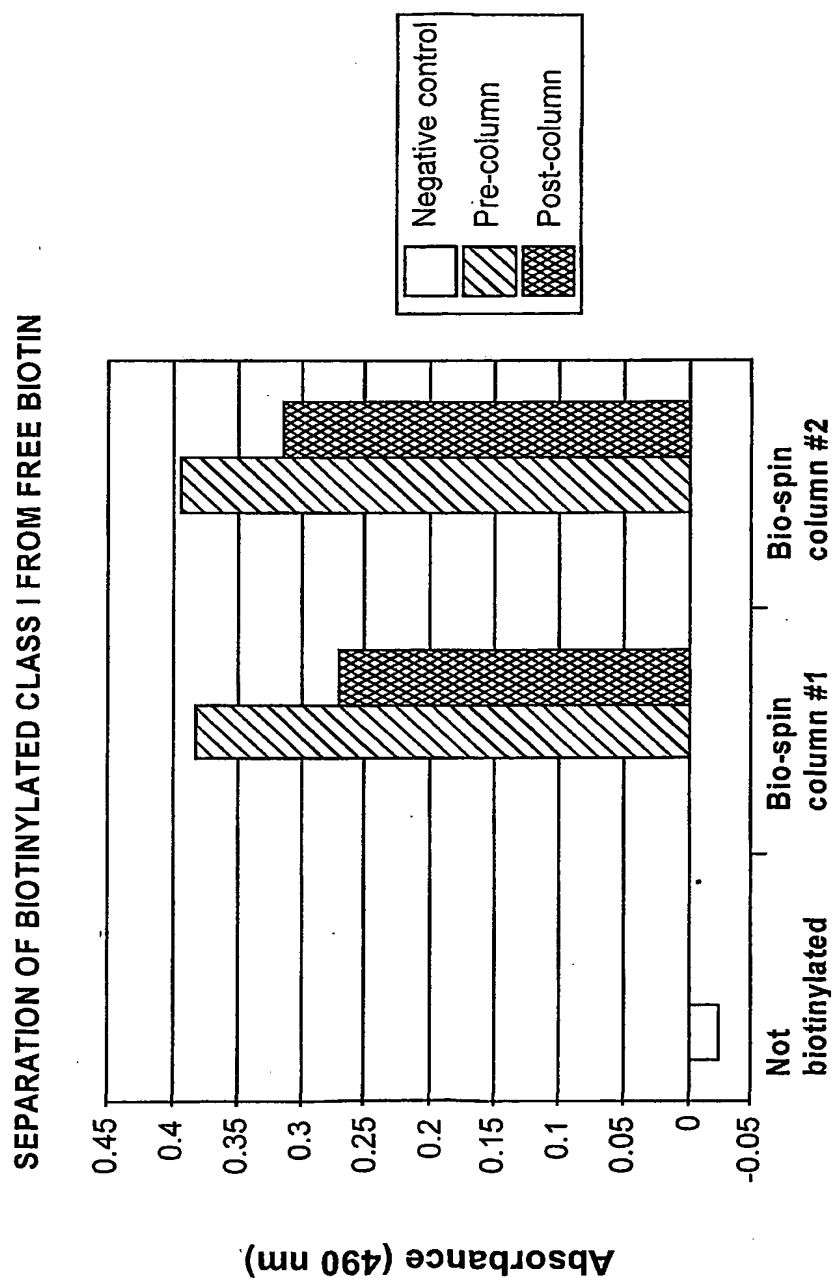


Fig. 47